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Case Report: Clinical and pathological features of a metastasizing malignant mixed epithelial and stromal tumor of the kidney in a Siberian tiger (*Panthera tigris altaica*)

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We describe a case of mixed epithelial and stromal tumor (MEST) of the kidney with widespread metastases in an eight-year-old male Siberian tiger (*Panthera tigris altaica*) from the ZOO Ljubljana. The tiger presented with hematuria, and after unsuccessful treatment attempts with antibiotics, an ultrasound examination followed by laparotomy was performed. During the laparotomy, a large encapsulated lesion weighing 9.3 kg was found in the location of the left kidney and surgically removed. Histopathological examination of the lesion revealed an encapsulated tumor with a biphasic growth pattern consisting of mesenchymal and epithelial components. Numerous mitoses, hemorrhages, and necrosis were found in the mesenchymal component, which resembled ovarian stroma, leading to the diagnosis of malignant MEST of the kidney. After 3 months, the tiger clinically deteriorated and was euthanized after diagnostic imaging revealed multiple metastases. At necropsy, numerous metastases were found at the site of the previously excised tumor, at the laparotomy scar, in the peritoneum, liver, pancreas, omentum, mesentery, lungs, and sternal lymph nodes. Microscopically, the tumor lesions consisted only of the malignant mesenchymal component. To our knowledge, this is the first report of malignant MEST of the kidney with peritoneal seeding and distant metastases in animals.

KEYWORDS

tiger, kidney, tumor, mixed epithelial and stromal tumor, pathology

1 Introduction

Tumors in captive tigers are uncommon. Mostly single cases of tumors such as pancreatic neuroendocrine tumor (1), Sertoli cell tumor (2), mesothelioma (3, 4), malignant peripheral nerve sheath tumor (5), meningioma (6), maxillary epithelial odontogenic tumor (7), pancreatic adenocarcinoma and Brunner's gland adenoma (8), mandibular squamous cell carcinoma (9), Leydig cell tumor (10), cutaneous metastatic melanoma (11) and mammary carcinoma (12) have been reported to date. In addition, there are some retrospective studies on tumors in captive wild felids, which also report the occurrence of tumors in tigers (13–17)

with the most common being mammary adenocarcinomas (13, 15), uterine leiomyomas (14) and thyroid adenomas (14, 15).

Mixed epithelial and stromal tumor (MEST) of the kidney is a rare biphasic tumor consisting of epithelial and stromal components (18, 19). Due to its rarity and overlapping microscopic features, the tumor has been described under various names, such as adult mesoblastic nephroma, cystic hamartoma of the renal pelvis, leiomyomatous renal hamartoma, and more recently as renal epithelial and stromal tumor (REST) (20). According to data from human medicine, MEST are usually unilateral, confined to the kidney and renal pelvis (21) and occur predominantly in women (19, 22), typically during perimenopause (22). Microscopically, MEST show a biphasic growth pattern with cysts of different sizes and glands embedded in a spindle cell stroma that resembles ovarian stroma (22). Most MEST are benign tumors, but there are also rare cases with malignant transformation of the epithelial or stromal component (20, 23–30) and aggressive clinical behavior (30–34). Only a few cases have recurred (31, 32) or have either metastasized to the lymph nodes, liver and lungs or have metastasized systemically (22, 33, 35, 36).

To our knowledge, three reports of MEST have been published in veterinary medicine, in a ringtail lemur (*Lemur catta*) (37), a beagle dog (38) and a mouse (39). However, all cases exhibited benign clinicopathologic features and had not metastasized.

The aim of our study is to describe a case of malignant MEST in a Siberian tiger (*Panthera tigris altaica*) with very aggressive clinical behavior with peritoneal seeding and distant metastases that led to euthanasia 3 months after initial diagnosis based on excisional biopsy. We report the history, gross and histopathologic findings of the excisional biopsy, imaging results, and necropsy findings.

2 Case description

An eight-year-old intact male Siberian tiger (*Panthera tigris altaica*) weighing 180 kg from the ZOO Ljubljana was presented with hematuria. The animal showed no other clinical signs of disease and did not respond to initial antibiotic treatment for suspected urinary bladder infection. As the animal was not trained for voluntary ultrasound examination, it was performed under general anesthesia. During the examination, an abdominal mass was detected on ultrasonography, which warranted an immediate laparotomy during the same general anesthesia. Anesthesia was induced with 300 mg ketamine (Ketamidor 100 mg/mL, VetViva Richter GmbH, Austria), 15 mg midazolam (Midazolam Accord 5 mg/mL, Accord Healthcare, Poland), 10 mg butorphanol (Butomidor 10 mg/mL, VetViva Richter GmbH, Austria), and 8 mg medetomidine (Domitor 1 mg/mL, Orion Corporation, Finland), administered intramuscularly. The dosage of the drugs was based on standard species-specific protocols. Following induction of anesthesia, an intravenous catheter was placed and the tiger was intubated with an endotracheal tube. Despite apparent sedation, a positive palpebral reflex was noted, indicating insufficient anesthetic depth. Therefore, an additional bolus of ketamine was administered intravenously to deepen anesthesia. Anesthesia was maintained with isoflurane administered via anesthetic circuit. Before extubation, reversal agents were administered subcutaneously: atipamezole (Antisedan 5 mg/mL, Orion Corporation, Finland) to antagonize the effect of medetomidine, and flumazenil (Anexate 100 µg/mL, Cheplapharm Arzneimittel GmbH, Germany) to reverse the effect of midazolam. The laparotomy revealed a very large encapsulated lesion at the location of the left kidney, which was then surgically removed. During the same procedure, after inspection of the abdominal cavity, a splenectomy was performed due to a nodule on the spleen. No other lesions suggestive of metastases were found in the abdominal cavity and thoracic radiographs were unremarkable. The tiger made a full recovery, and hematologic and biochemical parameters were within normal limits on the day of surgery.

The entire encapsulated lesion and spleen were immediately chilled and submitted to the Institute of Pathology, Wild Animals, Fish and Bees, of the Veterinary Faculty University of Ljubljana.

The lesion from the location of the left kidney measured 35 × 26 × 12 cm, weighed 9.3 kg, and was well circumscribed by a gray-white, 5-mm-thick capsule. After partial removal of the capsule, the lesion's surface was gray-red, lobulated, and had both solid areas with an arboriform architecture and cystic areas (Figure 1). The cut surface of the lesion was gray-red, solid, cystic and bosselated with numerous hemorrhages and necrotic areas. The spleen had a well-demarcated, exophytic, nodular, gray-red lesion measuring 2 × 1.8 × 1.4 cm with a red cut surface.

Representative samples of both lesions were fixed in 10% neutral buffered formalin, routinely processed for microscopic examination, sectioned at 4 µm and stained with hematoxylin and eosin (H&E) and examined under a light microscope. Immunohistochemistry was performed according to the manufacturer's instructions to confirm the origin of the neoplastic cells and to investigate a possible hormonal influence on tumor pathogenesis. The procedure was performed on selected 4-µm-thick paraffin sections. Details of the utilized primary antibodies and immunohistochemistry protocols used are provided in Table 1.

FIGURE 1
The excisional biopsy of the lesion in the left kidney of a Siberian tiger (*Panthera tigris altaica*), diagnosed with malignant mixed epithelial and stromal tumor (MEST). The well-circumscribed tumor was encapsulated by a grey-white capsule (blue arrows). The surface of the tumor after partial removal of the capsule was grey-red, lobulated and with solid and cystic areas. The solid areas displayed an arboriform architecture. Bar = 5 cm.

TABLE 1 Details of the primary antibodies and immunohistochemical protocols.

Primary antibody, clone, and catalogue number	Manufacturer	Antigen retrieval	Antibody dilution	Time and temperature of incubation of the primary antibody	Detection system	IHC staining procedure
Multicytokeratin, AE1/AE3, (NCL-L-AE1/AE3-601)	Novocastra, Germany	CC1, 64 min, 95 °C	1/50	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
GATA3, L50-823, (390 M-16)	Cellmarque, USA	CC1, 32 min, 100 °C	1/50	32 min, 37 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Estrogen receptors, SP1, (05278406001)	Ventana Roche, USA	CC1, 64 min, 95 °C	RTU	16 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Progesterone receptors, 1E2, (05277990001)	Ventana Roche, USA	CC1, 64 min, 95 °C	RTU	12 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
CD31, JC70A, (M0823)	Dako Agilent, USA	CC1, 64 min, 95 °C	1/15	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
CD34, QBEnd-10, (M0751)	Dako Agilent, USA	CC1, 64 min, 95 °C	1/20	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
FVIII (poly), (A0082)	Dako Agilent, USA	/	1/800	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Desmin, DE-R-11, (05267005001)	Ventana Roche, USA	CC1, 64 min, 100 °C	RTU	20 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
αSMA, 1A4, (202 M-96)	Cellmarque, USA	CC1, 64 min, 95 °C	1/100	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Calretinin, SP65, (05992184001)	Ventana Roche, USA	CC1, 56 min, 100 °C	RTU	20 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Vimentin, EPR3776, (ab92547)	Abcam, UK	CC1, 56 min, 100 °C	1/600	20 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
CK20, SP33, (05587760001)	Ventana Roche, USA	CC1, 56 min, 100 °C	RTU	20 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Pax8, SP348, (ab227707)	Abcam, UK	CC1, 64 min, 100 °C	1/100	32 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
CD10, 56C6, (NCL-L-CD10-270)	Novocastra, Germany	CC1, 56 min, 100 °C	1/15	32 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
MelanA, A103, (05278350001)	Ventana Roche, USA	CC1, 72 min, 100 °C	RTU	32 min, 36 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
HMB45, Hmb45, (M0634)	Dako Agilent, USA	CC1, 64 min, 100 °C	1/20	32 min, 36 °C	Ventana, UltraView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Carbon anchidrase (CAIX), EP161, (379R-14)	Cellmarque, USA	CC1, 56 min, 95 °C	1/100	20 min, 37 °C	Ventana, OptiView (Ventana Medical Systems Inc., Tucson, AZ, USA)	Automated
Uroplakin III, AU1, (696168)	Progen, Germany	Citrate buffer, pH 6.0, MW (1,100 W), 20 min	1/200	60 min, 23 °C	DAKO REAL™ EnVision Detection System Peroxidase/DAB+, Rabbit/Mouse (Dako, Denmark)	Manual
p63, (CM163A)	Biocare Medical, USA	Citrate buffer, pH 6.0, MW (1,100 W), 20 min	1/200	60 min, 23 °C	DAKO REAL™ EnVision Detection System Peroxidase/DAB+, Rabbit/Mouse (Dako, Denmark)	Manual

RTU, ready to use.

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Microscopic examination of the lesion from the location of the left kidney revealed a multinodular tumor composed of epithelial and stromal components, the majority of which was well demarcated from the renal parenchyma by a fibrous capsule.

The epithelial component displayed tubular and cystic structures lined by cuboidal to columnar neoplastic epithelial cells resembling urothelium with umbrella cells, with the cystic spaces containing serous or proteinaceous fluid (Figures 2A,B). The neoplastic epithelial cells exhibited mild anisocytosis, had a small to moderate amount of pale eosinophilic cytoplasm, round to oval nuclei exhibiting moderate anisokaryosis, and a single prominent nucleolus or an inconspicuous nucleolus. Mitoses were rare, with 1 or less than 1 mitosis per 10 highpower field (2.37 mm^2). The tubular structures exhibited positive immunolabeling for CKAE1/AE3, GATA3 and PAX8, while the stromal component was negative (Figure 3). Despite multiple attempts to optimize the protocol, no specific immunolabeling for estrogen and progesterone receptors, calretinin, CD20, p63, CD31, CD34 and uroplakin III was detected in either the tumor or the positive controls.

The stromal component consisted predominantly of neoplastic spindle cells reminiscent of ovarian stroma and multifocal groups of polygonal to round neoplastic stromal cells. The neoplastic spindle cells formed interwoven bundles and were multifocally condensed around neoplastic epithelial structures and blood vessels. In addition, multifocal papillary-like structures of neoplastic spindle cells were observed protruding into the lumina of the cystic structures and covered by neoplastic epithelial cells. The stromal cells exhibited moderate anisocytosis, with small to moderate amounts of eosinophilic cytoplasm. The nuclei exhibited moderate anisokaryosis, were large, oval to round with coarse chromatin, prominent nucleoli and frequent mitotic figures, 90 per 10 high-power fields (2.37 mm^2), including atypical mitoses (Figures 2C,D). This component also demonstrated extensive necrosis and hemorrhage, with expansile rather than infiltrative tumor growth toward the surrounding renal tissue. Invasion of blood and lymphatic vessels was not observed. Stromal component expressed diffuse positive immunolabeling for CD10, vimentin, and desmin, and focal imunolabeling for alpha smooth muscle actin (αSMA), consistent with smooth muscle differentiation. Immunohistochemical stains for Melan-A, HMB-45, carbonic anhydrase IX (CAIX) and von Willebrand factor/factor VIII complex (FVIII) were negative in both the epithelial and stromal components. Interstitial fibrosis and tubular atrophy were observed in the remaining renal parenchyma.

Based on the gross, microscopic and immunohistochemical features of the tumor, a malignant MEST of the kidney was diagnosed. The nodular lesion in the spleen was diagnosed as hematoma.

After 3 months, the tiger clinically deteriorated. He developed lethargy and cachexia, with marked weight loss and muscle wasting despite normal food intake. Radiographic and ultrasound imaging revealed multiple metastases in the lungs and abdominal cavity. Based on the poor prognosis and in accordance with ethical and welfare considerations, the tiger was euthanized *ad tabulam* using 40 mL of T-61 (1 mL/ 5 kg) (1 mL containing embutramide 200 mg, mebezonium iodide 50 mg, and tetracaine hydrochloride 5 mg; Intervet International BV, Boxmeer, Netherlands) administered intravenously after it had already been under general anesthesia (combination of medetomidine (40 mcg/kg), ketamine, butorphanol

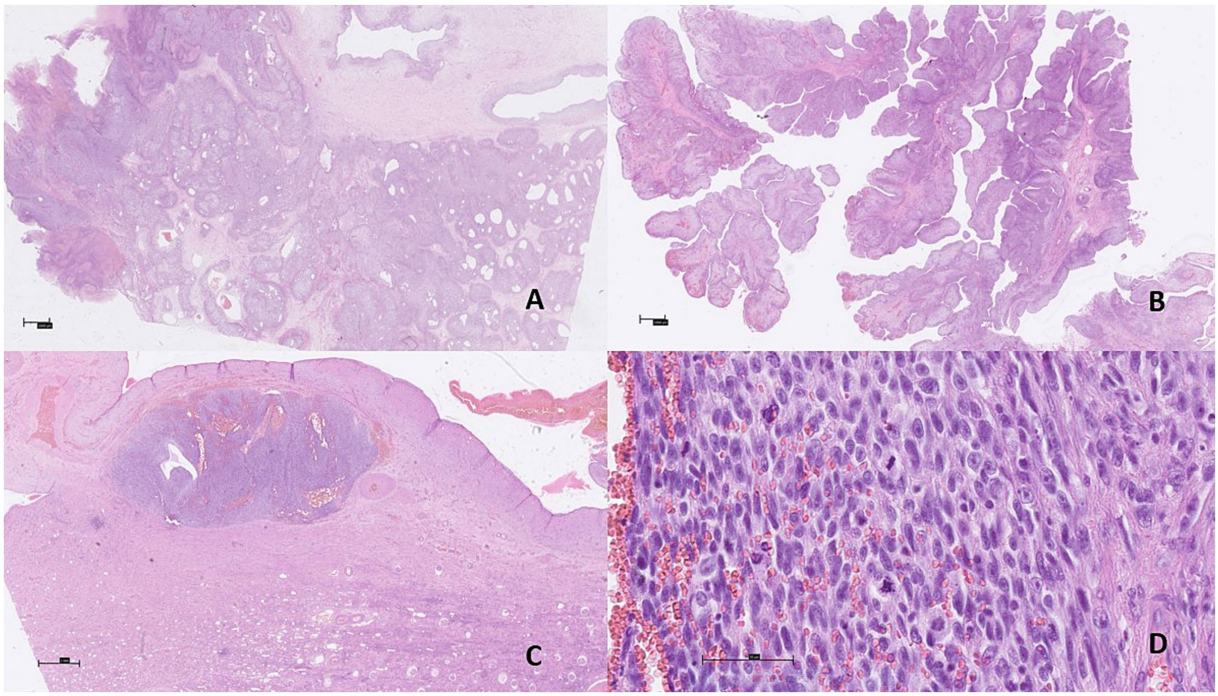

FIGURE 2
Microscopic findings of malignant mixed epithelial and stromal tumor (MEST) in the left kidney of a Siberian tiger (*Panthera tigris altaica*). The tumor was composed of epithelial and stromal components. **(A,B)** The epithelial component displayed papillary, tubular and cystic structures lined by epithelial cells. H&E, bar = 1,000 µm. **(C)** Focus of densely packed pleomorphic spindle stromal cells. H&E, bar = 1,000 µm. **(D)** High mitotic activity and atypical mitoses in the spindle stromal cells. H&E, bar = 50 µm.

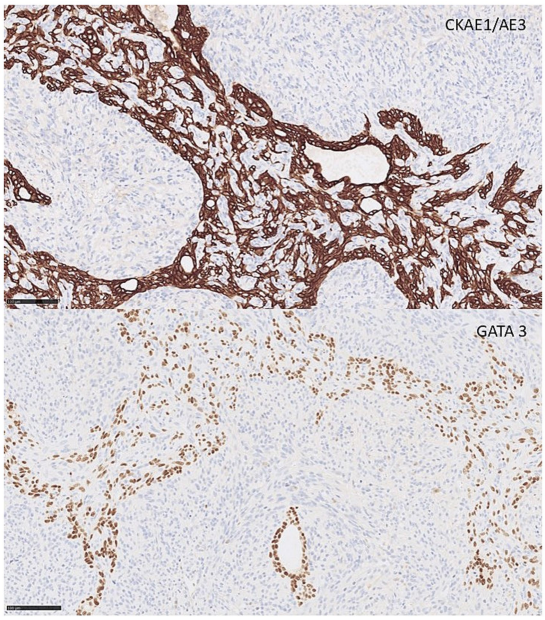

FIGURE 3
Microscopic findings of malignant mixed epithelial and stromal tumor (MEST) in the left kidney of a Siberian tiger (*Panthera tigris altaica*). Epithelial cells lining tubular structures were immunolabeled for GATA3 and CK AE1/AE3, while the stromal component showed immunolabeling for vimentin and αSMA. Immunohistochemistry, bar = 100 μm.

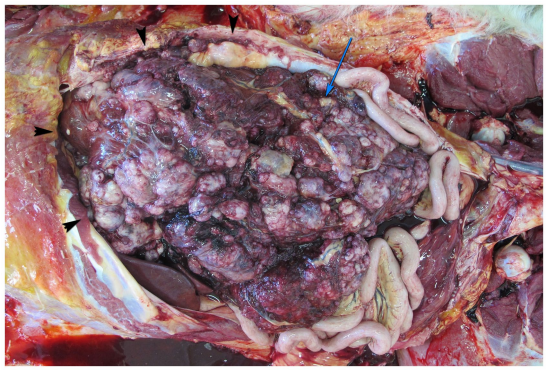

FIGURE 4
Necropsy findings in a Siberian tiger (*Panthera tigris altaica*) with malignant mixed epithelial and stromal tumor (MEST). Abdominal cavity with widespread peritoneal seeding. Tumors of different sizes are scattered in the abdominal cavity at the site of the previously excised tumor (blue arrow) and in the omentum. Arrowheads indicate the costal arch.

and midazolam 0.1 mg/ kg) for follow-up diagnostic procedures, and immediately submitted for necropsy.

At necropsy, numerous tumors of various sizes were found at the site of the previously excised tumor of the left kidney, in the area of the laparotomy scar, in the parietal peritoneum, in the serosa of the stomach and urinary bladder, in the liver, pancreas, omentum and mesentery (Figure 4). The tumor at the site of the previously excised renal tumor was the largest and measured 25 × 17 × 16 cm, the tumor in the area of the laparotomy scar measured 18 × 9 × 3 cm, while the tumors in the parenchymal organs, the omentum and the mesentery measured up to 9 × 2.5 × 2 cm. The tumors were round to oval, multinodular, gray-red, smooth, poorly demarcated and firm, with red-gray, smooth, bosselated cut surfaces. These tumors and the finding of two liters of watery, slightly opaque fluid in the abdominal cavity were consistent with peritoneal seeding. Numerous tumors ranging in size from 2 mm to 10 × 6 × 6 cm, grossly resembling the tumors in the abdominal cavity, were also scattered throughout the lung lobes. The sternal lymph nodes were severely enlarged and measured up to 8 × 4 × 4 cm. The tiger was in poor body condition. Moderate eccentric hypertrophy of the left ventricle with mild epicardial fibrosis, moderate endocardial fibrosis and severe dilatation of the right ventricle were also noted. In addition, there was 0.5 liters of yellow serous fluid in the thoracic cavity.

Samples of tumors from all the above listed organs and sites as well as samples from the right kidney, intestine, adrenal glands, mesenteric lymph nodes, heart, and brain were fixed in 10% buffered formalin, routinely embedded in paraffin, sectioned at 4 μm, stained with H&E, and examined under a light microscope. Microscopically, the tumors were poorly demarcated, nonencapsulated, infiltrative, densely cellular, and consisted exclusively of a malignant mesenchymal (stromal) component that resembled the stromal component of the primary tumor of the left kidney. The mitoses were numerous and often atypical. Numerous necroses and hemorrhages were scattered throughout the tumors.

The case was diagnosed as malignant MEST of the left kidney with widespread peritoneal seeding and distant metastases of the mesenchymal (stromal) component of the tumor to the lungs and sternal lymph nodes.

3 Discussion

MEST are rare in humans and account for 0.2% of all renal tumors (30). Most MEST are benign, but in very rare cases malignant transformation occurs (23–30, 40) and aggressive clinical behavior has been described (30–34). MEST with benign histopathologic features but extension into the inferior vena cava has also been described (41).

The gross, microscopic and immunohistochemical features of the tumor described in this tiger closely resemble those described in other cases of malignant MEST in humans (20, 23–30, 34). The epithelial component showed positive imunolabeling for cytokeratins and GATA3, while expression of vimentin, αSMA, and desmin confirmed smooth muscle differentiation within the stromal component. Given the biphasic architecture, morphological features, and aggressive biological behavior, several malignant biphasic renal tumors were considered in the differential diagnosis. These included papillary renal cell carcinoma with prominent spindle cell stroma, sarcomatoid renal cell carcinoma, and synovial sarcoma. Additional biphasic neoplasms, such as mesothelioma, epithelioid hemangiosarcoma, and urothelial carcinoma, were also included in the differential based on tumor architecture and anatomic location.

Correlation of histopathological and immunohistochemical findings enabled exclusion of these differential diagnoses. The presence of a well-defined mesenchymal component lined by an epithelial component ruled out papillary renal cell carcinoma with

spindle cell stroma. Renal cell carcinoma, including sarcomatoid variants, was excluded by the absence of CAIX expression. Epithelioid hemangiosarcoma was excluded by negative immunostaining for FVIII.

In malignant MEST in humans, malignant transformation can occur in either the epithelial or stromal component (20, 23–30), but most commonly malignant MEST has a malignant sarcomatous (stromal) component (25, 26, 29, 31, 32, 34), which is similar to our case. The malignant component in the herein described MEST resembled monophasic undifferentiated synovial sarcoma (23), synovial sarcoma (32), sarcoma with rhabdoid differentiation (24), rhabdomyosarcoma and chondrosarcoma (25), carcinosarcoma (36), undifferentiated sarcoma (26) and papillary renal cell carcinoma (28, 34). Sarcomas and carcinosarcomas typically lack expression of epithelial markers by immunohistochemistry. In contrast, the diffuse and strong immunolabeling for CK AE1/AE3 observed in the epithelial component is characteristic of MEST and differs from synovial sarcoma, in which cytokeratin expression is usually patchy, weak, and irregularly distributed among tumor cells (23, 25, 31, 32). PAX8, the tissue-specific transcription factors expressed primarily in the renal and Mullerian systems and also in Wolffian duct structures, has also been reported in MEST (20, 28, 41). Furthermore, coexpression of CD10 (20, 25, 36, 40), desmin (23, 36, 40), and αSMA (20, 23, 31, 40) in the stromal component provides additional support for the diagnosis of MEST.

In animals, MEST are even rarer than in humans. To the authors' knowledge, only three cases have been described in animals, specifically in a 14.5-year-old female ringtail lemur, a 22 month-old female beagle dog, and a 32-week-old female mouse, all of which had only benign pathomorphologic features (37–39). All three cases reported in animals were diagnosed in females (37–39) and were considered young (38) or middle-aged (37, 39).

Despite the gross and microscopic similarities of the tumor diagnosed in the tiger, the case differs from human MEST in many other features.

Most human MEST are diagnosed in women (29, 34), with a female to male ratio of 6:1 to 10:1, in perimenopause and at an average age of about 45 years (19, 42). Some studies describing long-term oral estrogen therapy and immunopositivity for estrogen and progesterone receptors suggest a possible role of hormonal stimulation in the pathogenesis of MEST (42–44). Only a few cases of MEST have been described in male patients (26, 27, 40, 45, 46). The age of men with MEST is variable; according to some data, patients were older, with an average age of 71 years (47), while other cases of MEST were diagnosed in forty-year-olds (45, 46) or even in 19-year-old men with no history of hormone therapy (27). In some cases, a history of hormone therapy for prostate cancer has been described, which may have an impact on tumor progression (26).

The tiger was an eight-year-old intact male, which given the species' average lifespan of 15 to.

20 years, corresponds to middle age (48). It was not under hormone therapy. Despite multiple attempts to optimize the protocol, immunolabeling of the tumor for estrogen and progesterone receptors was unsuccessful, thereby precluding assessment of their expression in the tumor. Due to the unsuccessful receptor immunolabeling, the involvement of hormonal mechanisms in the pathogenesis of the tumor cannot be confirmed, although this seems unlikely. The most common symptoms in human patients with MEST of the kidney are hematuria, flank pain, a palpable mass, or a urinary tract infection. Approximately 25% of cases of MEST are diagnosed incidentally (43, 44). In animals, the ringtail lemur with MEST showed reduced appetite and weight loss, and abdominal mass was palpated and visualized radiographically (37), while MEST in the dog and mouse were diagnosed incidentally at necropsy after the animals were euthanized for experimental studies (38, 39). The only clinical sign observed in the tiger prior to excisional biopsy and diagnosis of MEST of the kidney was hematuria that did not respond to antibiotic therapy.

The prognosis for MEST is generally good, while the prognosis for malignant MEST varies from case to case - some people are free of metastases or recurrence after nephrectomy, while some tumors have an aggressive clinical course with recurrence (27, 31, 32) or metastasize to the lymph nodes (22), liver (35) or lungs (33) or develop systemic metastases (36) and lead to death (26, 31, 32, 36). The shortest time from diagnosis to death was 5 months despite chemotherapy and radiotherapy (46). In our case, disease progression was also very rapid, with only 3 months from diagnosis to euthanasia due to widespread peritoneal seeding and metastases in the lungs and sternal lymph nodes. Complete resection with clean surgical margins is important for a good outcome in patients with malignant and benign MEST. Positive surgical margins, tumor disruption or tumor spillage during surgery may be associated with a poor outcome (32, 34, 46). Peritoneal seeding of benign MEST has been described after incomplete resection (48). Given the widespread peritoneal seeding and a large tumor in the abdominal wall at the laparotomy scar, we believe that the rapid progression of disease was probably the result of intraoperative tumor spillage and that the extreme size of the tumor probably contributed greatly to this outcome.

4 Conclusion

This is the first described case of malignant MEST of the kidney with peritoneal seeding and distant metastases in a Siberian tiger (*Panthera tigris altaica*) and the first such case ever reported in an animal. Although all previously reported MEST in animals have been benign and most MEST described in humans are also benign, the tumor described here showed microscopic features of malignant transformation with numerous and atypical mitoses, hemorrhages and necrosis on excisional biopsy. After only 3 months, the tiger was euthanized as its health had deteriorated considerably. This was the result of tumors at the site of the previously excised kidney tumor and laparotomy scar, peritoneal seeding, and metastases in the lungs and sternal lymph nodes, which microscopically consisted of the malignant component of the previously removed MEST of the kidney. This indicates that extreme caution is required when surgically removing these tumors, as intraoperative tumor spillage may lead to peritoneal seeding and distant metastases within a short period of time.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

Ethical approval was not required for the study involving animals in accordance with the local legislation and institutional requirements because the approval of the Ethics Committee/Welfare Authority was not required as all samples were taken intraoperatively or during necropsy. Written informed consent was obtained from the participant/patient(s) for the publication of this case report.

Author contributions

TŠ: Visualization, Data curation, Conceptualization, Writing – original draft, Writing – review & editing, Investigation. MG: Writing – review & editing, Investigation, Writing – original draft, Data curation, Visualization, Conceptualization. NK: Investigation, Writing – original draft, Visualization. MH: Investigation, Writing – review & editing, Data curation. MK: Investigation, Data curation, Writing – review & editing. PKr: Investigation, Data curation, Writing – review & editing. TD: Data curation, Investigation, Writing – review & editing. PKv: Investigation, Writing – review & editing, Supervision, Writing – original draft, Data curation.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Nyska A, Goldstein J, Klein B. Immunohistochemical study of pancreatic neuroendocrine tumor in *Panthera tigris tigris*. *J Wildl Dis*. (1996) 32:541–4. doi: 10.7589/0090-3558-32.3.541

2. Scudamore CL, Meredith AL. Sertoli cell tumour in an Amur tiger. *J Comp Pathol*. (2001) 124:79–82. doi: 10.1053/jcpa.2000.0422

3. Wiedner EB, Isaza R, Lindsay WA, Case AL, Decker J, Roberts J. Pericardial mesothelioma in a Bengal tiger (*Panthera tigris*). *J Zoo Wildl Med*. (2008) 39:121–3. doi: 10.1638/2007-0080.1

4. Coe SE, Garner MM, Kiupel M. Immunohistochemical characterization of mesothelioma in 6 large felids. *J Vet Diagn Invest*. (2021) 33:767–71. doi: 10.1177/10406387211015640

5. Steinmetz HW, Rütten M, Ruess-Melzer K, Ohlerth S, Lischer C, Oevermann A, et al. Clinical course of a malignant peripheral nerve sheath tumor in a Siberian tiger (*Panthera tigris altaica*). *J Vet Diagn Invest*. (2010) 22:970–5. doi: 10.1177/104063871002200621

6. Akin EY, Baumgartner WA, Lee JK, Beasley MJ. Meningioma in a Bengal tiger (*Panthera tigris tigris*). *J Zoo Wildl Med*. (2013) 44:761–4. doi: 10.1638/2012-0215R.1

7. Fecchio RS, Gomes M d S, Xavier JG, Kunze PE, Gioso MA. Maxillary calcifying epithelial odontogenic tumor in a Siberian tiger (*Panthera tigris altaica*). *J Vet Dent*. (2015) 32:120–1. doi: 10.1177/089875641503200206

8. Gombač M, Dolenšek T, Jaušovec D, Kvapil P, Švara T, Pogačnik M. Simultaneous occurrence of pancreatic adenocarcinoma and Brunner's gland adenoma in a Siberian tiger (*Panthera tigris altaica*). *J Comp Pathol*. (2015) 153:363–7. doi: 10.1016/j.jcpa.2015.08.008

9. de Oliveira AR, de Carvalho TF, Arenales A, Tinoco HP, Coelho CM, Costa M, et al. Mandibular squamous cell carcinoma in a captive Siberian tiger (*Panthera tigris altaica*). *Braz J Vet Pathol*. (2018) 11:97–101. doi: 10.24070/bjvp.1983-0246.v11i3p97-101

10. Kawata R, Tatsuhito I, Hori T, Machida Y, Ochiai K, Azakami D, et al. Leydig cell tumor in an Amur tiger (*Panthera tigris altaica*). *J Vet Med Sci*. (2019) 81:186–9. doi: 10.1292/jvms.18-0573

11. Eckstein C, Tinoco HP, Coelho CM, Lima PA, Rocha CEV, Santos RL. Cutaneous metastatic melanoma in a Siberian tiger (*Panthera tigris altaica*) – case report. *Arq Bras Med Vet Zootec*. (2020) 72:921–5. doi: 10.1590/1678-4162-10319

12. Fraser C, Keong Kok M, Shameha Abdul Razak I, Puspitasari Y, Salleh A. Diagnostic challenge in veterinary pathology: metastatic mammary tumor in a female tiger (*Panthera tigris*). *Vet Pathol*. (2024) 61:508–11. doi: 10.1177/03009858241226650

13. Owston MA, Ramsay EC, Rotstein DS. Neoplasia in felids at the Knoxville zoological gardens, 1979–2003. *J Zoo Wildl Med*. (2008) 39:608–13. doi: 10.1638/2008-068.1

14. Junginger J, Hansmann F, Herder V, Lehmbecker A, Peters M, Beyerbach M, et al. Pathology in captive wild felids at German zoological gardens. *PLoS One*. (2015) 10:e0130573. doi: 10.1371/journal.pone.0130573

15. Kloft HM, Ramsay EC, Sula MM. Neoplasia in captive *Panthera* species. *J Comp Pathol*. (2019) 166:35–44. doi: 10.1016/j.jcpa.2018.10.178

16. Mathieu A, Garner MM. A retrospective study of neoplasia in nondomestic felids in human care, with a comparative literature review. *J Zoo Wildl Med*. (2021) 52:413–26. doi: 10.1638/2020-0077

17. d'Aquino I, Piegari G, Casciaro SM, Prisco F, Rosato G, Silvestre P, et al. An overview of neoplasia in captive wild felids in southern Italy zoos. *Front Vet Sci*. (2022) 9:899481. doi: 10.3389/fvets.2022.899481

18. Michal M, Syrucek M. Benign mixed epithelial and stromal tumor of the kidney. *Pathol Res Pract*. (1998) 194:445–8. doi: 10.1016/S0344-0338(98)80038-1

19. Michal M. Benign mixed epithelial and stromal tumor of the kidney. *Pathol Res Pract*. (2000) 196:275–6. doi: 10.1016/S0344-0338(00)80078-3

20. Tinguria M, Chorneyko K. Mixed epithelial and stromal tumor: a rare renal neoplasm—case report with clinicopathologic features and review of the literature. *Case Rep Pathol*. (2023) 2023:1–11. doi: 10.1155/2023/3528377

21. Cruz Bendezú AM, Batter TH, Mufarrij P. Mixed epithelial and stromal tumor of the kidney extending to the proximal ureter in a 41-year-old female. *Urol Case Rep*. (2021) 38:1731. doi: 10.1016/j.eucr.2021.101731

22. Holkar PS, Jain T, Kavishwar V, Pandya JS. Metastasis in mixed epithelial stromal tumour of the kidney: a rare presentation. *BMJ Case Rep*. (2019) 12:293. doi: 10.1136/bcr-2019-229293

23. Svec A, Hes O, Michal M, Zachoval R. Malignant mixed epithelial and stromal tumor of the kidney. *Virchows Arch*. (2001) 439:700–2. doi: 10.1007/s004280100518

24. Sukov WR, Cheville JC, Lager DJ, Lewin JR, Sebo TJ, Lewin M. Malignant mixed epithelial and stromal tumor of the kidney with rhabdoid features: report of a case including immunohistochemical, molecular genetic studies and comparison to morphologically similar renal tumors. *Hum Pathol*. (2007) 38:1432–7. doi: 10.1016/j.humpath.2007.03.022

25. Jung SJ, Shen SS, Tran T, Jun SY, Truong L, Ayala AG, et al. Mixed epithelial and stromal tumor of kidney with malignant transformation: report of two cases and review of literature. *Hum Pathol*. (2008) 39:463–8. doi: 10.1016/j.humpath.2007.08.008

26. Suzuki T, Hiragata S, Hosaka K, Oyama T, Kuroda N, Hes O, et al. Malignant mixed epithelial and stromal tumor of the kidney: report of the first male case. *Int J Urol*. (2013) 20:448–50. doi: 10.1111/j.1442-2042.2012.03155.x

27. Zou L, Zhang X, Xiang H. Malignant mixed epithelial and stromal tumor of the kidney: the second male case and review of literature. *Int J Clin Exp Pathol*. (2014) 7:2658–63.

28. Mudaliar KM, Mehta V, Gupta GN, Picken MM. Expanding the morphologic spectrum of adult biphasic renal tumors – mixed epithelial and stromal tumor of the kidney with focal papillary renal cell carcinoma: case report and review of the literature. *Int J Surg Pathol*. (2014) 22:266–71. doi: 10.1177/1066896913488823

29. Vanecek T, Pivovarcikova K, Pitra T, Peckova K, Rotterova P, Daum O, et al. Mixed epithelial and stromal tumor of the kidney: mutation analysis of the DICER1 gene in 29 cases. *Appl Immunohistochem Mol Morphol*. (2017) 25:117–21. doi: 10.1097/PAI.0000000000000262

30. Bakavičius A, Barisienė M, Snicorius M, Valančienė D, Dasevičius D, Žalimas A, et al. Malignant mixed epithelial and stromal tumour of the kidney: a case report and a literature review. *Acta Med Litu*. (2018) 25:31–7. doi: 10.6001/actamedica.v25i1.3701

31. Nakagawa T, Kanai Y, Fujimoto H, Kitamura H, Furukawa H, Maeda S, et al. Malignant mixed epithelial and stromal tumours of the kidney: a report of the first two cases with a fatal clinical outcome. *Histopathology*. (2004) 44:302–4. doi: 10.1111/j.1365-2559.2004.01782.x

32. Yap YS, Coleman M, Olver I. Aggressive mixed epithelial-stromal tumour of the kidney treated with chemotherapy and radiotherapy. *Lancet Oncol*. (2004) 5:747–9. doi: 10.1016/S1470-2045(04)01651-1

33. Ozluk Y, Sari SO, Guzel NT, Firat P, Akbulut F, Kilicaslan I. Mixed epithelial and stromal tumor of the kidney with sarcomatous transformation metastatic to the lung: a case report. *Anal Quant Cytopathol Histpathol*. (2015) 37:199–205.

34. Arriola AGP, Taylor BL, Ma S, Malkowicz SB, Lal P. Malignant mixed epithelial and stromal tumor of the kidney with two simultaneous renal carcinomas in a male patient: case report and review of the literature. *Int J Surg Pathol*. (2018) 26:56–63. doi: 10.1177/1066896917720032

35. Levin NP, Damjanov I, Depillis VJ. Mesoblastic nephroma in an adult patient: recurrence 21 years after removal of the primary lesion. *Cancer*. (1982) 49:573–7. doi: 10.1002/1097-0142(19820201)49:3<>3.0.CO;2-#

36. Kuroda N, Sakaida N, Kinoshita H, Matsuda T, Hes O, Michal M, et al. Carcinosarcoma arising in mixed epithelial and stromal tumor of the kidney. *APMIS*. (2008) 116:1013–5. doi: 10.1111/j.1600-0463.2008.01063.x

37. Muller S, Oevermann A, Wenker C, Altermatt HJ, Robert N. A mixed epithelial and stromal tumor of the kidney in a ringtail lemur (*Lemur catta*). *Vet Pathol*. (2007) 44:243–6. doi: 10.1354/vp.44-2-243

38. Yamaoka M, Sato Y, Masumoto Y, Enomoto M, Mitsumori K. Spontaneous renal mixed epithelial and stromal tumor (MEST) in a beagle. *J Toxicol Pathol*. (2008) 21:189–92. doi: 10.1293/tox.21.189

39. Takabatake M, Takuwa Y, Takuwa N, Yasuno H, Matsumoto S, Shibutani M, et al. A case report of a renal mixed epithelial and stromal tumor in a heterozygous S1P2 receptor deficient mouse. *J Vet Med Sci*. (2008) 70:483–5. doi: 10.1292/jvms.70.483

40. Caliò A, Cheng L, Martignoni G, Zhang S, Brunelli M, Eble JN. Mixed epithelial and stromal tumours of the kidney with malignant transformation: a clinicopathological study of four cases. *Pathology*. (2022) 54:707–20. doi: 10.1016/j.pathol.2022.03.011

41. Picken MM, Bova D, Pins MR, Quek ML. Mixed epithelial and stromal tumor of the kidney with extension into inferior vena cava: case report and discussion of adult biphasic cystic renal lesions and the significance of vascular involvement. *Case Rep Pathol*. (2018) 2018:1–6. doi: 10.1155/2018/8234295

42. Mohanty SK, Parwani AV. Mixed epithelial and stromal tumors of the kidney: an overview. *Arch Pathol Lab Med*. (2009) 133:1483–6. doi: 10.5858/133.9.1483

43. Adsay NV, Eble JN, Srigley JR, Jones EC, Grignon DJ. Mixed epithelial and stromal tumor of the kidney. *Am J Surg Pathol*. (2000) 24:958–70. doi: 10.1097/00000478-200007000-00007

44. Lane BR, Campbell SC, Remer EM, Fergany AF, Williams SB, Novick AC, et al. Adult cystic nephroma and mixed epithelial and stromal tumor of the kidney: clinical, radiographic, and pathologic characteristics. *Urology*. (2008) 71:1142–8. doi: 10.1016/j.urology.2007.11.106

45. Jithesh M, Bhat S, Paul F, Jose A. An unusual presentation of mixed epithelial and stromal tumor of the kidney. *Kerala Med J*. (2016) 9:76–8.

46. Gokden N, Dawson K, Lindberg M. Malignant rhabdoid tumor arising in a mixed epithelial, stromal tumor of kidney: report of a male case, review of the literature. *Pathol Res Pract*. (2020) 216:3151. doi: 10.1016/j.prp.2020.153151

47. Williamson SR, Calio A. Tumours of the kidney. Mixed epithelial and stromal renal tumours In: SR Williamson, editor. WHO classification of Tumours editorial board: urinary and male genital tumours. Lyon: International Agency for Research on Cancer (2022)

48. Farias JA, Laryea J, Gokden N, Kamel MH. Peritoneal seeding following incomplete resection of mixed epithelial stromal tumor of the kidney: first case report. *Urol Ann*. (2016) 8:114–7. doi: 10.4103/0974-7796.171493