



OPEN ACCESS

EDITED BY

Gulmira Khassanova,
S. Seifullin Kazakh AgroTechnical Research
University, Kazakhstan

REVIEWED BY

Diaga Diouf,
Cheikh Anta Diop University, Senegal
Himanshu Sharma,
Amity University, Mohali, India

*CORRESPONDENCE

Hoang Dang Khoa Do
✉ dhdhoa@ntt.edu.vn

RECEIVED 13 May 2025

ACCEPTED 29 August 2025

PUBLISHED 12 September 2025

CITATION

Nguyen NN and Do HDK (2025)
Sequencing and characterizing the complete
chloroplast genome of *Ardisia silvestris* Pit.,
a potential medicinal plant in Asia.
Front. Plant Sci. 16:1627578.
doi: 10.3389/fpls.2025.1627578

COPYRIGHT

© 2025 Nguyen and Do. This is an open-
access article distributed under the terms of
the [Creative Commons Attribution License](#)
(CC BY). The use, distribution or reproduction
in other forums is permitted, provided the
original author(s) and the copyright owner(s)
are credited and that the original publication
in this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Sequencing and characterizing the complete chloroplast genome of *Ardisia silvestris* Pit., a potential medicinal plant in Asia

Nhat Nam Nguyen ¹ and Hoang Dang Khoa Do ^{2,3*}

¹School of Agriculture and Aquaculture, Tra Vinh University, Vinh Long, Vietnam, ²Functional Genomics Research Center, NTT Hi-Tech Institute, Nguyen Tat Thanh University, Ho Chi Minh City, Vietnam, ³Center for Hi-Tech Development, Saigon Hi-Tech Park, Nguyen Tat Thanh University, Ho Chi Minh City, Vietnam

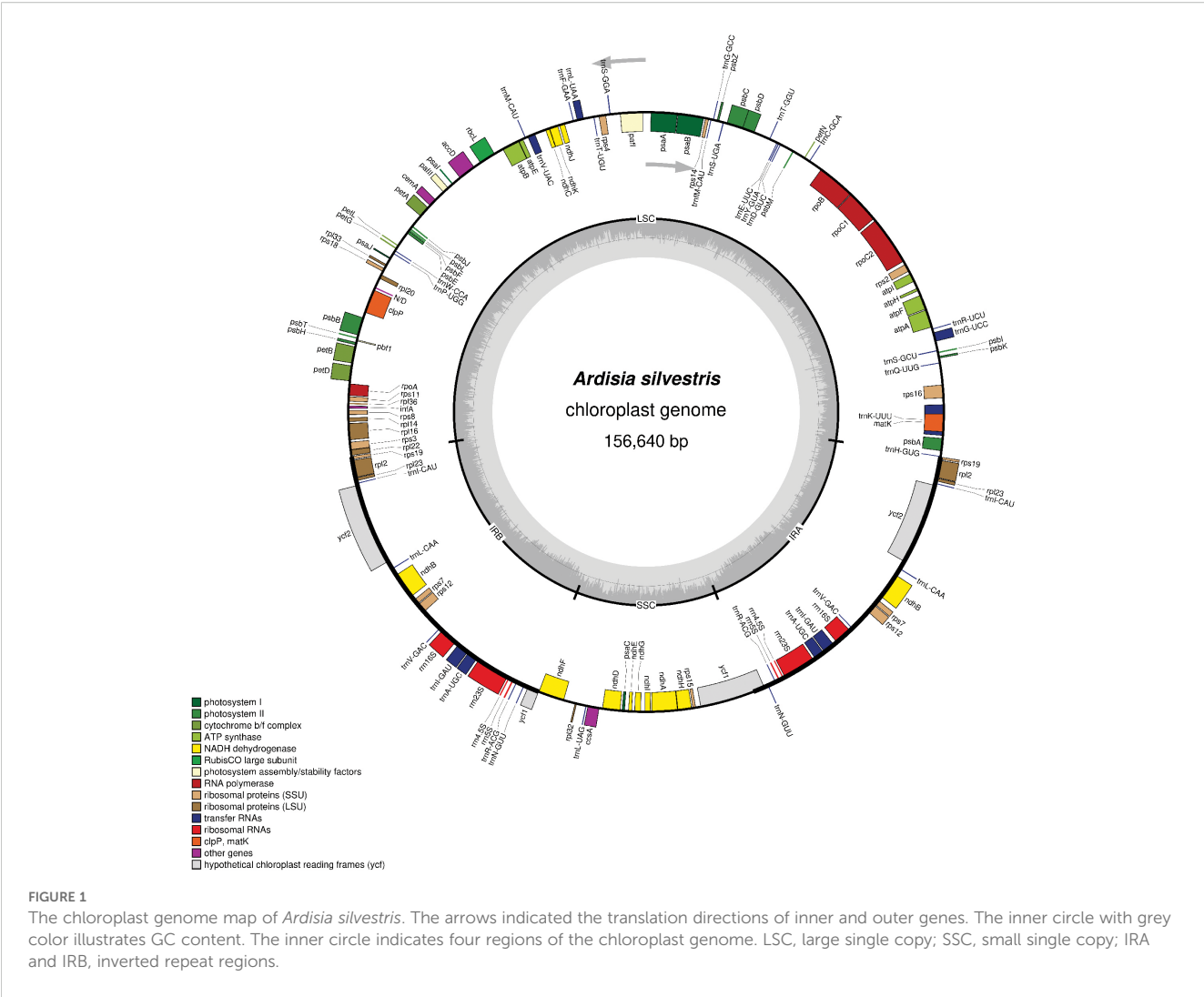
KEYWORDS

comparative genomics, myrsinoideae, plastid genome, plastome evolution, primulaceae

1 Introduction

Ardisia Sw. 1788 is one of 55 genera of Primulaceae and contains 739 accepted species that distribute in subtropical and tropical areas ([Plants of the World Online, 2025](#)). *Ardisia* species contains different phytochemical constituents such as coumarins, ardisiaquinones, and alkylphenols and was used as traditional medicine for fever, inflammation, and cancer ([Kobayashi and de Mejía, 2005](#); [de Mejía and Ramírez-Mares, 2011](#); [Liu et al., 2022](#); [Tian-Liang et al., 2024](#)). Specifically, a benzoquinonoid compound was extracted from *Ardisia crispa* and exhibited antimetastatic and antitumor features ([Kang et al., 2001](#)). The combination of *Ardisia gigantifolia* leaf extract and silver nanoparticles indicated an anti-cancer activity ([Le et al., 2023](#)). *Ardisia silvestris* is native to Vietnam and Hainan (China) and its ethanol extract possessed the characteristics of antiphotaging and skin-protective activities ([Huang et al., 2023](#); [Plant of the World Online, 2025](#)). Additionally, a previous study revealed a notable anti-inflammatory characteristic of *A. silvestris* ethyl acetate extract ([Thanh et al., 2025](#)). Also, the antioxidant and antibacterial properties of *A. silvestris* leaf extract ([Huynh, 2020](#)). These previous results demonstrated the medicinal values of *A. silvestris* and related species in *Ardisia* genus. However, genomic data, including nuclear, mitochondrial, and chloroplast genomes, of *A. silvestris* are limited and need further investigations.

Chloroplast genome is an essential component in autotrophic plants because it encodes genes responsible for performing photosynthesis ([Dobrogojski et al., 2020](#)). The chloroplast genome had a quadripartite structure including a large single copy, a small single copy, and two inverted repeat regions, which could be altered in both autotrophic and heterotrophic plants ([Daniell et al., 2016](#)). Additionally, the genomic information of chloroplast genomes reflected the evolutionary history, which was used to explore a billion years of plant evolution ([Gitzendanner et al., 2018](#)). Previously, chloroplast genomes of Primulaceae species have been reported ([Xu et al., 2020](#); [Xie et al., 2023](#); [Li et al., 2024](#)). The complete



chloroplast genomes of various *Ardisia* species such as *A. crispa*, *A. gigantifolia*, *A. crenata*, *A. villosa*, *A. mamillata*, *A. brunnescentis*, *A. pusilla*, *A. squamulosa*, *A. brevicaulis*, and *A. crenata* were also published (Xie et al., 2021; Ye et al., 2024; Yuan et al., 2024). In the current study, we report the complete chloroplast genome of *Ardisia silvestris*, collected in Vietnam, using the Illumina sequencing platform. The result of our study enriches the chloroplast genome data of *Ardisia* genus and provides initial chloroplast genomic data for further genomic studies examining phylogeny and molecular markers of *A. silvestris* and related taxa in Primulaceae.

2 Materials and methods

2.1 Plant sampling, DNA extraction, and next-generation sequencing

The healthy leaves of *Ardisia silvestris* were collected from living collection of medicinal plants at Tra Vinh University, Vinh Long

Province, Vietnam (9°55'25.0"N 106°20'52.4"E). Then, the leaves were stored at −80°C in a deep freezer for further experiments. The total genomic DNA was extracted from the frozen leaves of *A. silvestris* using DNeasy Plant Pro Kit (Qiagen, USA) following the manufacturer's instructions. The quality of DNA sample was checked using NanoDrop One Microvolume UV-Vis Spectrophotometer (Thermo Fisher Scientific, USA) and 1% agarose gel electrophoresis. The DNA sample selected for Nextseq550 sequencing (Illumina, USA) should have a concentration of 100 ng/μL and show a clear band on the agarose gel. The TruSeq DNA Nano kit (Illumina, USA) was used to prepare sequencing library to generate paired-end reads of 150 bp following the manufacturer's instructions.

2.2 Assembly and annotation of chloroplast genome

The raw reads were qualified and filtered using fastp v0.24.1 to remove the adapter sequences and eliminate the reads possessing a

TABLE 1 Gene composition of *Ardisia silvestris* chloroplast genome.

Groups of genes	Name of genes	Quantity
Ribosomal RNAs	<i>rrn4.5^a, rrn5^a, rrn16^a, rrn23^a</i>	8
Transfer RNAs	<i>trnA_UGC^{a,b}, trnC_GCA, trnD_GUC, trnE_UUC, trnF_GAA, trnG_UCC^b, trnG_GCC, trnH_GUG, trnI_GAU^{a,b}, trnK_UUU^b, trnL_CAA^a, trnL_UAA^b, trnL_UAG, trnI_M_CAU, trnI_CAU^a, trnM_CAU, trnN_GUU^a, trnP_UGG, trnQ_UUG, trnR_ACG^a, trnR_UCU, trnS_GCU, trnS_GGA, trnS_UGA, trnT_GGU, trnT_UGU, trnV_GAC^a, trnV_UAC^b, trnW_CCA, trnY_GUA</i>	37
Large units of ribosome	<i>rpl2^{a,b}, rpl14, rpl16^b, rpl20, rpl22, rpl23^a, rpl32, rpl33, rpl36</i>	11
Small units of ribosome	<i>rps2, rps3, rps4, rps7^a, rps8, rps11, rps12^a, rps14, rps15, rps16^b, rps18, rps19^a</i>	15
RNA polymerase	<i>rpoA, rpoB, rpoC1^b, rpoC2</i>	4
Translational initiation factor	<i>infA</i>	1
Subunit of photosystem I	<i>psaA, psaB, psaC, psaI, psaJ, pafI^c, pafII</i>	7
Subunit of photosystem II	<i>psbA, psbB, psbC, psbD, psbE, psbF, psbH, psbI, psbJ, psbK, psbL, psbM, psbT, psbZ</i>	15
Subunit of cytochrome	<i>petA, petB^b, petD^b, petG, petL, petN</i>	6
Subunit of ATP synthases	<i>atpA, atpB, atpE, atpF^b, atpH, atpI</i>	6
Large unit of Rubisco	<i>rbcL</i>	1
Subunit of NADH dehydrogenase	<i>ndhA^b, ndhB^{a,b}, ndhC, ndhD, ndhE, ndhF, ndhG, ndhH, ndhI, ndhJ, ndhK</i>	12
Maturase	<i>matK</i>	1
Envelope membrane protein	<i>cemA</i>	1
Subunit of acetyl-CoA	<i>accD</i>	1
C-type cytochrome synthesis gene	<i>ccsA</i>	1
ATP-dependent protease subunit P	<i>clpP1^c</i>	1
Hypothetical proteins and conserved reading frames	<i>ycf1^a, ycf2^a</i>	4

^aduplicated gene in IR region; ^bgenes containing single intron, ^cgenes containing two introns.

Qscore under 20, having length shorter than 100 bp, and containing more than five N bases (Chen et al., 2018). The remaining high-quality reads were then assembled to complete chloroplast genome using NOVOPlasty v4.3.5 with the reference sequence of *Ardisia fordii* (NCBI accession number NC_060707) and other default settings (Dierckxsens et al., 2016). Consequently, the newly completed chloroplast genome of *A. silvestris* was annotated using Geseq through online interface at <https://chlorobox.mpimp-golm.mpg.de/geseq.html> with default settings (Tillich et al., 2017). To verify the annotation of Geseq, the annotation of protein-coding region was rechecked the start and stop codon of each gene using Geneious Prime v2024.0.1 (<https://www.geneious.com/>) whereas the structural formation of tRNA regions were tested using tRNAscan-SE 2.0 available at <https://lowelab.ucsc.edu/tRNAscan-SE/index.html> with default settings (Chan and Lowe, 2019). Additionally, the quadripartite structure of chloroplast genome, including a large single copy, a small single copy, and two inverted repeat regions, was investigated using the “Find repeat” function with the setting of minimum repeat length of 10,000 bp of Geneious Prime v0.2024.1 to locate two inverted repeat regions that flanked the large single copy and the small single copy regions. The map of chloroplast genome was illustrated using OGDRAW v1.3.1 available at <https://chlorobox.mpimp-golm.mpg.de/OGDraw.html> with default settings for plastid sequences (Greiner et al., 2019). The complete chloroplast genome of *A. silvestris* was deposited to GenBank under accession number PV608499.

3 Results

The assembly process resulted in a quadripartite chloroplast genome of *A. silvestris* with a mean coverage of 1642x (Figure 1). This genome was 156,640 bp in length and had 37.3% GC content. Additionally, the complete chloroplast genome of *A. silvestris* consisted of a large single copy (LSC) region of 85,812 bp (35.2% GC content), a small single copy (SSC) region of 18,388 bp (30.4% GC content), and two inverted repeat (IR) regions of 26,220 bp (43.2% GC content) each. Further observation revealed that the junction between LSC and IR regions located within *rps19* coding region whereas that of SSC and IR regions was in the coding region of *ycf1*. The complete chloroplast genome of *A. silvestris* encoded 79 unique protein-coding genes, 30 unique transfer RNA genes, and four unique ribosomal RNA genes (Table 1). Among 113 unique coding genes, 19 regions were duplicated in IR region including *rps19*, *rpl2*, *rpl23*, *trnI_CAU*, *ycf2*, *trnL_CAA*, *ndhB*, *rps7*, *rps12*, *trnV_GAC*, *rrn16*, *trnI_GAU*, *trnA_UGC*, *rrn23*, *rrn4.5*, *rrn5*, *trnR_ACG*, *trnN_GUU*, and *ycf1*. Notably, *ycf1* and *rps19* exhibited incomplete duplication due to expansion of IR regions. Additionally, there were nine protein genes (including *rps16*, *atpF*, *rpoC1*, *petB*, *petD*, *rpl16*, *rpl2*, *ndhB*, and *ndhA*) and six tRNAs (including *trnK_UUU*, *trnI_GAU*, *trnA_UGC*, *trnG_UCC*, *trnL_UAA*, and *trnV_UAC*) contained one intron. Meanwhile, *pafI* and *clpP1* had two introns. The *rps12* gene was trans-spliced of which the exon 2 and exon 3 located in IR regions.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: <https://www.ncbi.nlm.nih.gov/genbank/>, PV608499 <https://www.ncbi.nlm.nih.gov/>, PRJNA1261444.

Author contributions

NN: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Project administration, Writing – original draft, Writing – review & editing. HD: Conceptualization, Data curation, Formal Analysis, Methodology, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

Acknowledgments

We acknowledge Nguyen Tat Thanh University, Ho Chi Minh City, Vietnam for supporting this study. We acknowledge the support of time and facilities from Tra Vinh University (TVU) for this study.

References

- Chan, P. P., and Lowe, T. M. (2019). “tRNAscan-SE: searching for tRNA genes in genomic sequences,” in *Gene prediction. Methods in molecular biology*. Ed. M. Kollmar (New York, NY: Humana), 1–14. doi: 10.1007/978-1-4939-9173-0_1
- Chen, S., Zhou, Y., Chen, Y., and Gu, J. (2018). fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34, i884–i890. doi: 10.1093/bioinformatics/bty560
- Daniell, H., Lin, C.-S., Yu, M., and Chang, W.-J. (2016). Chloroplast genomes: diversity, evolution, and applications in genetic engineering. *Genome Biol.* 17, 134. doi: 10.1186/s13059-016-1004-2
- de Mejia, E. G., and Ramirez-Mares, M. V. (2011). Ardisia: health-promoting properties and toxicity of phytochemicals and extracts. *Toxicol. Mech. Methods* 21, 667–674. doi: 10.3109/15376516.2011.601355
- Dierckxsens, N., Mardulyn, P., and Smits, G. (2016). NOVOPlasty: *de novo* assembly of organelle genomes from whole genome data. *Nucleic Acids Res.* 45, gkw955. doi: 10.1093/nar/gkw955
- Dobrogojski, J., Adamiec, M., and Luciński, R. (2020). The chloroplast genome: a review. *Acta Physiol. Plant* 42, 98. doi: 10.1007/s11738-020-03089-x
- Gitzendanner, M. A., Soltis, P. S., Wong, G. K.-S., Ruhfel, B. R., and Soltis, D. E. (2018). Plastid phylogenomic analysis of green plants: A billion years of evolutionary history. *Am. J. Bot.* 105, 291–301. doi: 10.1002/ajb2.1048
- Greiner, S., Lehwark, P., and Bock, R. (2019). OrganellarGenomeDRAW (OGDRAW) version 1.3.1: expanded toolkit for the graphical visualization of organellar genomes. *Nucleic Acids Res.* 47, W59–W64. doi: 10.1093/nar/gkz238
- Huang, L., You, L., Aziz, N., Yu, S. H., Lee, J. S., Choung, E. S., et al. (2023). Antiphotaging and skin-protective activities of ardisia silvestris ethanol extract in human keratinocytes. *Plants* 12, 1167. doi: 10.3390/plants12051167
- Huynh, B. V. (2020). Phytochemical analysis of Ardisia silvestris leaf extracts and their antioxidant and antibacterial activities. *J. Agric. Dev.* 19, 28–35. doi: 10.52997/jad.4.04.2020
- Kang, Y.-H., Kim, W. H., Park, M. K., and Han, B. H. (2001). Antimetastatic and antitumor effects of benzoquinonoid AC7–1 from Ardisia crispa. *Int. J. Cancer* 93, 736–740. doi: 10.1002/ijc.1384

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- Kobayashi, H., and de Mejia, E. (2005). The genus Ardisia: a novel source of health-promoting compounds and phytopharmaceuticals. *J. Ethnopharmacol.* 96, 347–354. doi: 10.1016/j.jep.2004.09.037
- Le, T. T. H., Ngo, T. H., Nguyen, T. H., Hoang, V. H., Nguyen, V. H., and Nguyen, P. H. (2023). Anti-cancer activity of green synthesized silver nanoparticles using Ardisia gigantifolia leaf extract against gastric cancer cells. *Biochem. Biophys. Res. Commun.* 661, 99–107. doi: 10.1016/j.bbrc.2023.04.037
- Li, J.-T., Ju, W.-B., Li, X., Zhu, Y., Cao, T.-Y., Zhou, Y.-S., et al. (2024). The complete chloroplast genome sequence of Primula medogensis (Primulaceae) and its phylogeny. *Mitochondrial DNA Part B* 9, 1404–1408. doi: 10.1080/23802359.2024.2415137
- Liu, B., Liu, R., Liu, Q., Ashby, C. R., Zhang, H., and Chen, Z. (2022). The ethnomedicinal and functional uses, phytochemical and pharmacology of compounds from Ardisia species: An updated review. *Med. Res. Rev.* 42, 1888–1929. doi: 10.1002/med.21894
- Plants of the World Online (2025). *Ardisia silvestris* Pit. Available online at: <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:587602-1> (Accessed March 3, 2025).
- Thanh, M. P., Dung, H. M., Tuan, L. A., Huong, N. T. T., Ngoc, T. T. H., Giang, L. T., et al. (2025). Anti-inflammatory effects of the extracts from Ardisia silvestris Pitard leaves in experiment. *Tạp chí Nghiên cứu Y học* 190, 188–195. doi: 10.52852/tcnycy.v190i5E16.3279
- Tian-Liang, Xie, Q., Shama, R., Yu, J., Xi-Gu-Ri-Gan, Q., Bao, Q., et al. (2024). Ethnobotanical study of Zhuang medicinal herbs of Ardisia: variety systematization, traditional uses, phytochemistry, pharmacology, clinical application, and toxicity. *J. Pharm. Pharmacol.* 76, 327–353. doi: 10.1093/jpp/rgad119
- Tillich, M., Lehwark, P., Pellizzer, T., Ulbricht-Jones, E. S., Fischer, A., Bock, R., et al. (2017). GeSeq – versatile and accurate annotation of organelle genomes. *Nucleic Acids Res.* 45, W6–W11. doi: 10.1093/nar/gkx391
- Xie, C., An, W., Liu, S., Huang, Y., Yang, Z., Lin, J., et al. (2021). Comparative genomic study on the complete plastomes of four official Ardisia species in China. *Sci. Rep.* 11, 22239. doi: 10.1038/s41598-021-01561-3

Xie, Y., Yang, G., Zhang, C., Zhang, X., and Jiang, X. (2023). Comparative analysis of chloroplast genomes of endangered heterostylous species *Primula wilsonii* and its closely related species. *Ecol. Evol.* 13, e9730. doi: 10.1002/ece3.9730

Xu, W., Xia, B., and Li, X. (2020). The complete chloroplast genome sequences of five pinnate-leaved *Primula* species and phylogenetic analyses. *Sci. Rep.* 10, 20782. doi: 10.1038/s41598-020-77661-3

Ye, J., Luo, Q., Lang, Y., Ding, N., Jian, Y., Wu, Z., et al. (2024). Analysis of chloroplast genome structure and phylogeny of the traditional medicinal of *Ardisia crispa* (Myrsinaceae). *Sci. Rep.* 14, 19045. doi: 10.1038/s41598-024-66563-3

Yuan, L., Ni, Y., Chen, H., Li, J., Lu, Q., Wang, L., et al. (2024). Comparative chloroplast genomes study of five officinal *Ardisia* Species: Unraveling interspecific diversity and evolutionary insights in *Ardisia*. *Gene* 912, 148349. doi: 10.1016/j.gene.2024.148349