

#### **OPEN ACCESS**

EDITED BY Jinyao Li, Xinjiang University, China

REVIEWED BY Miroslaw Janowski, University of Maryland, United States

\*CORRESPONDENCE
Chunbao Chen,

□ chen\_1992358@163.com
Zhinei Lin,
□ zhimlin82@163.com

RECEIVED 14 September 2025 REVISED 28 October 2025 ACCEPTED 11 November 2025 PUBLISHED 24 November 2025

#### CITATION

Du X, Chen C, Zeng Y and Lin Z (2025) Overcoming the blood-brain barrier: targeted delivery strategies for gliomas. Front. Pharmacol. 16:1705234. doi: 10.3389/fphar.2025.1705234

#### COPYRIGHT

© 2025 Du, Chen, Zeng and Lin. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Overcoming the blood-brain barrier: targeted delivery strategies for gliomas

Xue Du<sup>1</sup>, Chunbao Chen<sup>2</sup>\*, Yijun Zeng<sup>2</sup> and Zhinei Lin<sup>1</sup>\*

<sup>1</sup>Department of Oncology and Hematology, The 3RD Affiliated Hospital of Chengdu Medical College, Chengdu Pidu District People's Hospital, Chengdu, China, <sup>2</sup>Department of Neurosurgery, The 3RD Affiliated Hospital of Chengdu Medical College, Chengdu Pidu District People's Hospital, Chengdu, China

Neurological gliomas, as the most common and deadly primary brain tumors, face two major therapeutic obstacles: the blockade of the blood-brain barrier (BBB) and high tumor heterogeneity. In recent years, research on novel drug delivery systems has brought hope for glioma treatment. This article elaborates on the research progress of novel drug delivery systems in glioma treatment, including various nanocarriers, targeted delivery strategies, and gene therapy drug delivery systems. It analyzes their advantages and challenges, outlooks future development directions, and aims to provide a reference for optimizing drug delivery systems for glioma therapy.

KEYWORDS

neurological glioma, drug delivery systems, blood-brain barrier, blood-tumor barrier, nanocarriers, targeted therapy

## 1 Introduction

Neurological gliomas originate from glial cells and are the most common primary malignant tumors in the adult central nervous system (CNS). According to the 2021 World Health Organization (WHO) Classification of Tumors of the Central Nervous System, diffuse gliomas are classified into isocitrate dehydrogenase (IDH)-mutant astrocytomas (WHO Grades 2–4), IDH-mutant and 1p/19q codeleted oligodendrogliomas (WHO Grades 2 and 3), and IDH-wildtype glioblastomas (GBM, WHO Grade 4) (Louis et al., 2021). The clinical treatment regimen for brain gliomas involves comprehensive therapy including surgical resection, radiotherapy, and chemotherapy (Stupp et al., 2005). Due to the infiltrative growth and unclear boundaries of gliomas, complete resection is difficult to achieve. Simultaneously, the existence of the Blood-Brain Barrier (BBB) and the Blood-Tumor Barrier (BTB) hinders the entry of anti-tumor drugs into the brain to exert efficacy, leading to slow progress in glioma treatment (van Tellingen et al., 2015; Chen et al., 2023).

The BBB is a highly selective barrier between the blood and brain tissue, restricting the transmembrane transport of many drugs, making it difficult for approximately 98% of small molecule drugs and almost all large molecule drugs to enter the brain parenchyma (Pardridge, 2005). Furthermore, neurological gliomas can induce the formation of a blood-tumor barrier (BTB), further impeding drug delivery (van Tellingen et al., 2015). Therefore, developing efficient drug delivery systems to break through the limitations of the BBB and BTB and achieve effective drug enrichment at the tumor site is key to improving the therapeutic outcome of neurological gliomas.

# 2 Challenges in glioma treatment

# 2.1 Blood-brain barrier and blood-tumor barrier

The existence of the BBB is a major obstacle in glioma treatment. Its endothelial cells have tight junctions, lack fenestrations, possess highly expressed efflux transporters such as P-glycoprotein and breast cancer resistance protein, and contain abundant enzyme systems, all working together to prevent drugs from entering the brain (Obermeier et al., 2013; Kadry et al., 2020). Even if some drugs can cross the BBB, the amount reaching the tumor site is extremely low, often failing to achieve effective therapeutic concentrations. Although the BTB has higher permeability than the BBB in some aspects, it also has its own complexities. The abnormal structure and function of tumor vessels lead to heterogeneous permeability of the BTB, resulting in uneven distribution of drugs within the tumor tissue (Chen et al., 2023). Simultaneously, the BTB retains some of the barrier properties of the BBB, further increasing the difficulty of drug delivery (Arvanitis et al., 2020).

# 2.2 Tumor heterogeneity

Neurological gliomas are highly heterogeneous, exhibiting significant differences in molecular characteristics, biological behavior, and response to therapy among different patients, different regions of the same tumor, and different tumor cell subpopulations (Nicholson and Fine, 2021; Eschbacher et al., 2021). This heterogeneity makes it difficult for a single treatment regimen to be effective against all tumor cells, easily leading to tumor recurrence and drug resistance (Nicholson and Fine, 2021).

# 2.3 Limitations of Traditional therapeutic drugs

Traditional chemotherapeutic drugs like temozolomide, while capable of inhibiting tumor cell growth to some extent, lack targeting specificity. While acting on tumor cells, they also cause toxic side effects on normal tissue cells (Choi et al., 2018). Furthermore, long-term use of chemotherapeutic drugs can easily induce drug resistance in tumor cells, reducing treatment efficacy. Some novel therapeutic drugs, such as gene therapy drugs (Umemura et al., 2023) and immunotherapy drugs (Xu et al., 2020), although potentially highly effective and specific, face drug delivery challenges and struggle to reach tumor cells effectively to exert their effects (Song et al., 2024).

# 3 Research progress on novel drug delivery systems

#### 3.1 Nanocarriers

#### 3.1.1 Liposomes

Liposomes are vesicles composed of phospholipids and other lipid materials with bilayer or multilayer membrane structures. They

possess good biocompatibility, biodegradability, and low immunogenicity (Large et al., 2021). Liposomes can encapsulate both hydrophilic and hydrophobic drugs, increasing drug solubility and protecting drugs from degradation by enzymes and other components in body fluids (Almeida et al., 2020). Modifying the liposome surface, such as by attaching polyethylene glycol (PEG) to form PEGylated liposomes, can prolong their circulation time in the blood and reduce phagocytosis by the mononuclear phagocyte system (Tenchov et al., 2023). Further modification with targeting ligands, such as mannose (MAN) (Li et al., 2019) or transferrin (Tf) (Dos Santos Rodrigues et al., 2019), enables targeted recognition of specific receptors on tumor cells or the BBB (Eroğlu and İbrahim, 2020).

In delivering nucleic acid drugs, cationic liposomes can bind negatively charged siRNA through electrostatic interactions, achieving siRNA loading (Hu et al., 2024). However, cationic liposomes suffer from cytotoxicity and poor in vivo stability. The addition of PEG chains can improve their pharmacokinetic properties but may also limit the uptake of liposomes by tumor cells (Yin et al., 2022). Meng JL et al. developed a pH-responsive anti-polyethylene glycol (PEG) × anti-TfR bispecific antibody (pH-PEG engager TfR). This antibody can complex with PEGylated nanomedicine at physiological pH to trigger TfR-mediated transcytosis in brain microvascular endothelial cells, while rapidly dissociating from the PEGylated nanomedicine at acidic endosomes for efficient release of the nanomedicine to cross the BBB (Meng et al., 2025). The conditional release of PEGylated nanomedicine during BBB-related receptor-mediated transcytosis by pH-pH-PEG engager TfR is promising for enhanced brain drug delivery to treat CNS disorders. Although such smart delivery systems have demonstrated promising performance in preclinical studies, significant variations in the relative efficacy of different liposomal systems persist. The feasibility of large-scale production and batchto-batch consistency remain major obstacles to clinical translation.

#### 3.1.2 Polymeric nanoparticles

Polymeric nanoparticles are nanoscale particles prepared from synthetic or natural polymers (Sofini et al., 2024). Commonly used synthetic polymers include polylactic acid (PLA) (Tyler et al., 2016), poly ( $\gamma$ -glutamic acid) (PGA) (Zhang et al., 2024), and their copolymer poly (lactic-co-glycolic acid) (PLGA) (Danhier et al., 2012). Natural polymers include chitosan (Kluczka, 2023) and gelatin (Santoro et al., 2014). Polymeric nanoparticles have good stability, tunable size, and surface properties, enabling effective drug encapsulation and controlled release (Gu et al., 2024).

PLGA nanoparticles exhibit good biocompatibility and biodegradability. Surface modification with different targeting ligands can achieve targeted delivery to gliomas (Das et al., 2023). Chitosan is a natural cationic polysaccharide with good biocompatibility, bioadhesion, and degradability (Abourehab et al., 2022). Chitosan nanoparticles can bind negatively charged drugs or biomacromolecules through electrostatic interactions for loading and delivery (Path et al., 2023). Lakkadwala S et al. (Lakkadwala and Singh, 2019) demonstrated that liposomes surface-modified with transferrin (Tf) for receptor targeting and with cell-penetrating peptide PFVYLI (PFV) increased the translocation of doxorubicin (Dox) and erlotinib (Erlo) across the BBB into glioblastoma (U87) tumor cells. Using an *in vitro* brain

tumor model, the translocation of dual-functionalized liposomes across the BBB was significantly (p < 0.05) higher, delivering chemotherapeutic drugs to the glioblastoma tumor cells inside a PLGA-Chitosan scaffold and resulting in approximately 52% tumor cell death. Although these results are encouraging, multiple challenges arise in actual clinical translation. First, the preparation process for the dual-modified system is complex, making it difficult to ensure batch-to-batch consistency. Second, in vitro models cannot fully replicate the intricate in vivo BBB microenvironment. Finally, non-specific uptake of the cell-penetrating peptide may induce toxic reactions in normal brain tissue. These issues partially explain why such complex systems struggle to advance into clinical research phases.

#### 3.1.3 Protein nanocarriers

Protein nanocarriers possess good biocompatibility, low immunogenicity, and precise structure (Elzoghby et al., 2012). Human heavy-chain ferritin (HFn) is a well-studied protein nanocarrier. HFn has properties allowing specific tumor recognition and BBB crossing (Kim et al., 2024). To address the issue of lysosomal escape when ferritin delivers siRNA, researchers developed a class of internally cationic HFn variants (HFn + NPs). The top-performing candidate, HFn2, effectively delivered siRNA to glioma cells after traversing the BBB and achieved the highest silencing efficacy among HFn + NPs (Yuan et al., 2022). HFn can selectively deliver a large amount of cargo into tumors in vivo via transferrin receptor 1 (TfR1)-mediated tumor-cellspecific targeting followed by rapid internalization. Utilizing the intrinsic tumor-targeting property and unique nanocage structure of human HFn, a broad variety of cargo-loaded HFn formulations have been developed for biological analysis, imaging diagnosis, and medicine development (Song et al., 2021). Although HFn2 demonstrated the highest gene silencing efficacy in preclinical studies, the translation of this protein nanocarrier from laboratory to clinical settings faces unique challenges. These include high costs associated with large-scale production, storage stability issues, and potential immunogenicity risks. Even with human-derived proteins, individual variations may trigger immune responses. Furthermore, the TfR1 expression required for HFn function exhibits heterogeneity across patients and tumor types, potentially limiting its broad applicability.

## 3.1.4 Inorganic nanoparticles

Inorganic nanoparticles, such as gold nanoparticles, silver nanoparticles, silica nanoparticles, and magnetic nanoparticles, possess unique physicochemical properties and show potential application value in drug delivery (Kumar et al., 2023). Gold nanoparticles have good biocompatibility, high stability, and surface modifiability. They can be used for photothermal therapy via surface plasmon resonance effects and also serve as drug carriers for loading chemotherapeutic drugs or nucleic acid drugs (Zhang et al., 2023). Modifying gold nanoparticles with cell-penetrating peptides can increase their accumulation in the brain for glioma treatment (Griveau et al., 2022).

Magnetic nanoparticles can move directionally and accumulate at the tumor site under an external magnetic field, enabling targeted drug delivery (Gobbo et al., 2015). Simultaneously, magnetic nanoparticles can be used for magnetic resonance imaging (MRI), allowing real-time monitoring of the drug delivery process (Sun et al., 2008). Prepared magnetic nanoparticles surface-modified with targeting ligands can effectively aggregate at the glioma site under magnetic field guidance, improving drug therapeutic efficacy (Tong et al., 2025; Bartusik-Aebisher et al., 2025). However, the clinical translation of inorganic nanoparticles faces challenges due to the lack of long-term *in vivo* safety data, potential bioaccumulation risks, and quality control issues in large-scale production. While magnetic nanoparticles surface-modified with targeting ligands can guide targeting to gliomas under magnetic fields, the penetration depth of external magnetic fields in human tissues is limited, significantly reducing targeting efficiency for deep-seated brain tumors. Furthermore, inorganic nanoparticles typically exhibit poor degradability, potentially leading to long-term toxicity concerns.

# 3.1.5 Systematic challenges in the clinical translation of nanocarriers

Across all types of nanocarriers, the conversion rate from preclinical research to clinical application remains extremely low. This phenomenon stems from multiple factors. First, differences between animal models and human physiology cause many nanocarriers effective in animal studies to perform poorly in humans. Second, large-scale production of nanocarriers faces quality control challenges, as complex surface modification processes struggle to maintain batch consistency in industrial manufacturing. Finally, regulatory bodies impose stringent requirements on the safety and quality of nanomedicines, yet many nanocarrier systems lack sufficient safety data and clear studies on their *in vivo* metabolic pathways.

The coexisting reproducibility issues cannot be overlooked. Many studies have failed to adequately characterize the physicochemical parameters of nanocarriers, such as particle size distribution, surface charge, and drug loading capacity, making it difficult to compare results across different laboratories. The lack of standardized procedures for complex preparation processes and modification strategies, coupled with discrepancies between *in vitro* experimental conditions and *in vivo* environments, collectively contribute to the reproducibility crisis in nanocarrier research.

Future research should place greater emphasis on addressing these translational barriers. By simplifying carrier design, establishing standardized characterization methods, enhancing safety assessments, and conducting more reliable preclinical model studies, we can further advance the progress of nanocarriers toward clinical application.

# 3.2 Targeted delivery strategies

# 3.2.1 Passive targeting

Passive targeting primarily utilizes the enhanced permeability and retention (EPR) effect of tumor tissue (Torchilin, 2000). The gaps between tumor vascular endothelial cells are larger, allowing nanocarriers to enter the tumor tissue through these gaps. Concurrently, impaired lymphatic drainage in the tumor region allows nanocarriers to remain in the tumor tissue for an extended period, achieving passive drug accumulation at the tumor site (Sebak

et al., 2021). Factors such as the size, shape, surface charge, and hydrophobicity of nanocarriers influence their EPR effect (Zhao et al., 2019). Generally, nanocarriers in the size range of 10–200 nm can better utilize the EPR effect for tumor targeting (Xu et al., 2023). Some drug delivery systems based on liposomes and polymeric nanoparticles have successfully achieved passive targeted delivery to glioma through rational design of nanocarrier size and surface properties (Caro et al., 2021).

However, due to its uneven intensity across various tumor microenvironments, the existence and significance of the EPR effect in nanomedicine design and application have been questioned. The EPR effect exhibits inter-tumor heterogeneity, varying among different patients and different tumor sites. Nanocarriers may also accumulate to some extent in non-tumor tissues, leading to potential toxic side effects (Maeda, 2015). The real challenge now lies in leveraging the EPR effect to design and enhance the therapeutic efficacy of nanomedicine. When considering the EPR effect, maintaining efficacy is crucial, as drugs must sustain effective concentrations within tumor tissues for a sufficient duration to achieve satisfactory anticancer activity.

#### 3.2.2 Active targeting

Active targeting involves modifying the nanocarrier surface with specific targeting ligands, enabling them to bind to specific receptors on tumor cells or the BBB for precise drug delivery (Dutta et al., 2021; Anthony et al., 2021). Commonly used targeting ligands include peptides, antibodies, and aptamers. Peptide ligands have advantages such as small molecular weight, low immunogenicity, and ease of synthesis (Jiang et al., 2019). After modification on the nanocarrier surface, peptides can specifically bind to corresponding receptors, significantly increasing the enrichment of nanocarriers in the brain and at the tumor site (Gao et al., 2014). Antibodies have high specificity and affinity, but their large molecular weight may affect the pharmacokinetic properties of nanocarriers, and they pose immunogenicity issues (Arslan et al., 2021). However, preparing antibody fragments or humanized antibodies can mitigate these problems to some extent (Hillman et al., 2019). Aptamers are singlestranded nucleic acid molecules obtained through in vitro selection techniques that can specifically bind to targets. They offer high affinity, high specificity, and ease of synthesis and modification, showing potential application prospects in active targeted delivery for glioma (Esposito et al., 2018).

## 3.3 Gene therapy drug delivery systems

Gene therapy provides new strategies for glioma treatment, such as delivering small interfering RNA (siRNA) (Jin et al., 2025), short hairpin RNA (shRNA) (Lu et al., 2020), or plasmid DNA (Yao et al., 2015) to silence or regulate tumor-associated genes. However, gene therapy drugs are mostly large biomolecules, carry a negative charge, are susceptible to degradation by nucleases, and find it difficult to cross cell membranes and the BBB (Körbelin et al., 2024). Therefore, its therapeutic efficacy depends on efficient and safe delivery systems. In fact, the various nanocarriers discussed earlier, through targeted design, serve as the core tools for addressing the challenges of gene drug delivery.

Systems such as liposomes, polymeric nanoparticles, and protein nanocarriers are widely used for encapsulating and protecting gene drugs due to their inherent properties or functional modifications (Hassan et al., 2025). Cationic liposomes efficiently condense negatively charged siRNA or DNA through electrostatic interactions, forming lipid nanocomplexes that shield nucleic acids from nuclease degradation (Sun and Lu, 2023). Cationic polymers (e.g., poly (ethyleneimine), chitosan) and engineered cationic protein nanocages (e.g., HFn+ NPs) similarly load gene drugs via electrostatic interactions (Liu et al., 2025). Therefore, developing effective delivery systems for gene therapy drugs is crucial. Viral vectors are also commonly used gene delivery tools, such as adenoviruses (de la Nava et al., 2024), adeno-associated viruses (AAV) (Merkel et al., 2017), and lentiviruses (Nivajärvi et al., 2020). However, viral vectors pose potential risks of immune responses, insertional mutagenesis, and difficulties in preparation and purification. While non-viral vectors are safer, their transfection efficiency needs further improvement (Butt et al., 2022). In contrast, non-viral nanocarrier systems offer advantages such as high safety, large payload capacity, and ease of large-scale production. Although their transfection efficiency has historically been lower than that of viral vectors, their delivery efficiency is rapidly improving through the synergistic design of integrated targeting ligands and endosomal escape mechanisms.

# 3.4 Osmotic blood-brain barrier opening

Beyond the targeted modifications of the nanocarriers themselves, physical "door-opening" strategies can also enable trans-barrier delivery of chemotherapeutic drugs or nanoparticles. Among these, osmotic blood-brain barrier opening (OBBBO) induced by intra-arterial infusion of hypertonic solutions facilitates drug delivery to the brain, providing a method for achieving global delivery of therapeutic agents to brain tumors and their surrounding regions (Sato et al., 1998). Osmotic opening of the blood-brain barrier (BBB) most likely is mediated by modification of interendothelial tight junctions, subsequent to shrinkage of cerebrovascular endothelial cells, and not by stimulation of transendothelial vesicular transport or by channel formation (Rapoport and Robinson, 1986). Selective and superselective intra-arterial cerebral infusion (SIACI/SSIACI) delivers chemotherapy directly to tumor-supplying arteries. Combined with the opening of the permeable blood-brain barrier, this approach significantly increases local tumor drug concentrations (Ferreira et al., 2025). A recent study proposed a novel method for opening the BBB by combining 25% mannitol with 4% sodium chloride, significantly enhancing BBB permeability while demonstrating favorable safety profiles in mouse experiments (Qiao et al., 2025).

Although intra-arterial hypertonic mannitol infusion has established itself as a clinically viable technique for opening the blood-brain barrier, its non-selective opening mechanism may allow neurotoxic substances to enter the brain alongside therapeutic agents. Furthermore, the brief duration of the opening window limits its efficacy and scope of application (Bellavanc et al., 2008). However, by integrating this mature permeable open technology with meticulously designed nanomedicine delivery systems and

advanced imaging-guided techniques, it establishes a complete "open-delivery-monitoring" technological closed-loop system, demonstrating tremendous synergistic potential and innovative prospects.

# 3.5 Emerging physical/mechanical barrier opening strategy

Compared to biochemical approaches, emerging physical and mechanical strategies directly apply mechanical forces generated by external physical fields to the blood-brain barrier, achieving its opening. These methods offer unique advantages including high spatio-temporal precision, reversible opening effects, and generally not relying on specific cell receptors or pathways, thereby pioneering novel pathways for central nervous system drug delivery.

#### 3.5.1 Focused ultrasound technology

Microbubble-mediated focused ultrasound (Mb-FUS) is a promising non-invasive technique for BBBO, enhancing drug delivery and immunomodulation for brain disease treatments (Bae et al., 2025). The principle involves intravenous injection of micron-sized ultrasound microbubbles, followed by precise excitation of microbubble oscillations in specific brain regions using focused ultrasound. This mechanical oscillation induces multiple biological effects on vascular endothelial cells, including mechanically stretching tight junctions through stable cavitation effects and activating relevant signaling pathways, thereby reversibly increasing blood-brain barrier permeability (Ibsen et al., 2013; McMahon et al., 2019). Combined with transcranial MR-guided focused ultrasound (MRgFUS), this technology enables millimeterlevel precision in opening the blood-brain barrier (Meng et al., 2021) and has been widely applied in clinical research to enhance the intracerebral delivery of therapeutic antibodies for Alzheimer's disease (Bae et al., 2024) and Parkinson's disease (Meng et al., 2022). Its safety and feasibility have been validated through multiple human trials. Numerous preclinical studies have validated the efficacy of FUS-MB in animal GBM models (Sun et al., 2017; Englander et al., 2021). Meanwhile, several clinical trials have established the safety and feasibility of FUS therapy for glioblastoma, and have demonstrated that FUS can induce immunomodulatory effects (Chen et al., 2021). Furthermore, repeated FUS-mediated BBBO concurrently with radiotherapy is safe and feasible. FUS combined with radiotherapy may improve glioma treatment outcomes, but further studies are needed to optimize the magnitude of effect (Tazhibi et al., 2024; Fletcher et al., 2024).

#### 3.5.2 Convection-enhanced drug delivery

Convection-enhanced delivery (CED) is an invasive local drug administration technique. It employs a slowly and continuously applied fluid infusion pressure within tumor cavities or brain parenchyma via intracranially implanted catheters, thereby generating a "convective" field that drives therapeutic drugs to distribute extensively and uniformly within tissue interstices (Mehta et al., 2017). CED technology can bypass the blood-brain barrier to deliver high-concentration drugs directly to target areas, significantly increasing the

volume of distribution. It is particularly suitable for the localized treatment of macromolecular drugs and has undergone extensive clinical exploration in the treatment of diseases such as malignant glioma (Nordling-David et al., 2017; Zhan and Wang, 2018). Early trials lacked effective methods for real-time monitoring of drug distribution within the brain, making it impossible to promptly adjust infusion parameters to ensure complete coverage of the target area. Additionally, there was a lack of long-term efficacy assessment (D'Amico et al., 2021). With the maturation of real-time image guidance key technologies, currently conducted dose-escalation clinical trials utilize MR-guided stereotactic (CED) techniques (Souweidane et al., 2025; Narsinh et al., 2025). Real-time MRI enables surgeons to dynamically adjust infusion parameters based on the actual distribution of the drug within the patient's brain, achieving truly personalized drug delivery. This ensures that the tumor region in each patient is fully saturated with the medication.

#### 3.5.3 Laser interstitial thermal therapy

Laser interstitial thermal therapy (LITT) is a minimally invasive surgical technique commonly used for the ablation of brain tumors (Carpentier et al., 2012). At energy parameters below those causing irreversible tissue coagulation necrosis, the sublethal thermal energy generated by LITT can temporarily disrupt the blood-brain barrier structure in the irradiated region (Mo et al., 2021). This thermal effect may achieve opening of the blood-brain barrier by influencing tight junction proteins or the cytoskeleton, creating synergistic opportunities for subsequent drug delivery within the hyperthermia zone (Sabel et al., 2003).

#### 3.5.4 Photodynamic therapy

Photodynamic therapy (PDT) utilizes photosensitizers and laser irradiation at specific wavelengths to generate reactive oxygen species, thereby killing tumor cells. Research has confirmed that under specific low-dose photodynamic therapy conditions, the reactive oxygen species produced can trigger inflammatory responses in vascular endothelial cells and the reorganization of tight junctions, leading to a temporary opening of the blood-brain barrier without causing widespread cell death (Semyachk et al., 2018; Semyachkina-Glushkovskaya et al., 2023). This window of opportunity provides a valuable chance to enhance the brain accumulation of subsequent chemotherapy drugs, targeted therapies, and even nanomedicine delivery systems (Li et al., 2025). Synergistic strategy of PDT and nanodelivery systems, this approach employs PDT as the initial intervention. Following intravenous injection of photosensitizers and localized light irradiation to induce BBB opening, mainstream chemotherapy drugs or targeted nanomedicines are subsequently administered (Guo et al., 2025; Chen et al., 2024). Sequential-controlled combination therapy—this "open the door first, then deliver the cargo" approach—significantly enhances the accumulation of subsequent drugs at tumor sites, achieving synergistic therapeutic effects where 1 + 1>2. Of course, future work will also need to optimize light exposure parameters and photosensitizer dosage to strike the optimal balance between opening the blood-brain barrier and causing permanent vascular damage, as well as address the limited penetration of light into deep brain tissues.

# 4 Challenges and prospects

# 4.1 Challenges

Although novel drug delivery systems have made progress in glioma treatment research, they still face many challenges. Firstly, the large-scale preparation and quality control of nanocarriers are difficult; achieving efficient, low-cost, and reproducible nanocarrier production is an urgent problem to be solved (Dahiya et al., 2021; Alshawwa et al., 2022). Secondly, the in vivo safety and long-term biological effects of nanocarriers are not yet fully understood; nanomaterials may aggregate or undergo abnormal metabolism in the body, posing potential hazards (Xuan et al., 2020). Furthermore, although various targeting strategies have been proposed, achieving completely precise targeted delivery in practical applications remains challenging; drug distribution in non-target tissues may cause toxic side effects (Ezike et al., 2023). Moreover, due to the high heterogeneity of gliomas, a single drug delivery system and treatment strategy may not meet the needs of all patients (Nicholson and Fine, 2021; Song et al., 2024).

# 4.2 Future prospects

Future research on novel drug delivery systems for glioma treatment could focus on the following aspects. First, further optimize the design of nanocarriers, developing smarter and more efficient ones, such as responsive nanocarriers that can achieve precise drug release and targeted delivery based on changes in the tumor microenvironment (Zhao et al., 2021). Second, deeply research the interaction mechanisms between nanocarriers and biological systems, clarifying their in vivo metabolic pathways and long-term safety to provide a solid theoretical foundation for clinical application (Deb and Jain, 2024). Third, combine multimodal treatment strategies, integrating drug delivery systems with immunotherapy, photothermal therapy, and radiotherapy to exert synergistic therapeutic effects and improve glioma treatment outcomes (Huang et al., 2021; Molinaro et al., 2024). Fourth, utilize artificial intelligence and big data technologies to achieve personalized treatment for glioma patients, tailoring suitable drug delivery plans and treatment strategies based on the molecular characteristics of the patient's tumor and individual differences (Noury et al., 2025). Through continuous research and innovation, more effective drug delivery systems for glioma treatment are expected to be developed, improving patient prognosis.

# 5 Conclusion

The drug delivery challenges faced in glioma treatment severely constrain the improvement of therapeutic efficacy. Novel drug delivery systems, particularly research on various nanocarriers based on nanotechnology and targeted delivery strategies, provide new avenues to address this challenge. Nanocarriers such as liposomes, polymeric nanoparticles, protein nanocarriers, and inorganic nanoparticles demonstrate unique advantages in improving drug solubility and stability, breaking through the blood-brain barrier, and achieving tumor-targeted delivery. The application of passive

and active targeting strategies further enhances the precision of drug delivery. Research on gene therapy drug delivery systems also brings new hope for glioma treatment. However, this field still faces numerous challenges, requiring in-depth research on nanocarrier preparation, safety evaluation, improvement of targeting precision, and personalized therapy. It is believed that with continuous research advancements, novel drug delivery systems will bring revolutionary breakthroughs in glioma treatment, significantly improving patients' quality of life and prognosis.

## **Author contributions**

XD: Conceptualization, Writing – original draft, Writing – review and editing. CC: Supervision, Investigation, Writing – review and editing. YZ: Supervision, Writing – review and editing. ZL: Supervision, Writing – review and editing.

# **Funding**

The authors declare that no financial support was received for the research and/or publication of this article.

# Acknowledgements

We thank the researchers who developed the CiteSpace software and the R software.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Generative AI statement

The authors declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

#### References

Sofini, H., Balasubramanian, D., Girigoswami, A., and Girigoswami, K. (2024). Biomedical applications of natural and synthetic polymer based nanocomposites. *J. Biomater. Sci. Polym. Ed.* 35 (2), 269–294. Epub 2024 Jan 25. PMID: 37962432. doi:10.1080/09205063.2023.2283910

Abourehab, M. A. S., Pramanik, S., Abdelgawad, M. A., Abualsoud, B. M., Kadi, A., Ansari, M. J., et al. (2022). Recent advances of chitosan formulations in biomedical applications. *Int. J. Mol. Sci.* 23 (18), 10975. PMID: 36142887; PMCID: PMC9504745. doi:10.3390/ijms231810975

Almeida, B., Nag, O. K., Rogers, K. E., and Delehanty, J. B. (2020). Recent progress in bioconjugation strategies for liposome-mediated drug delivery. *Molecules* 25 (23), 5672. doi:10.3390/molecules25235672

Alshawwa, S. Z., Kassem, A. A., Farid, R. M., Mostafa, S. K., and Labib, G. S. (2022). Nanocarrier drug delivery systems: characterization, limitations, future perspectives and implementation of artificial intelligence. *Pharmaceutics* 14 (4), 883. PMID: 35456717; PMCID: PMC9026217. doi:10.3390/pharmaceutics14040883

Anthony, D. P., Hegde, M., Shetty, S. S., Rafic, T., Mutalik, S., and Rao, B. S. S. (2021). Targeting receptor-ligand chemistry for drug delivery across blood-brain barrier in brain diseases. *Life Sci.* 274, 119326. Epub 2021 Mar 9. PMID: 33711385. doi:10.1016/j. lfs.2021.119326

Arslan, F. B., Ozturk Atar, K., and Calis, S. (2021). Antibody-mediated drug delivery. *Int. J. Pharm.* 596, 120268. Epub 2021 Jan 21. PMID: 33486037. doi:10.1016/j.ijpharm. 2021.120268

Arvanitis, C. D., Ferraro, G. B., and Jain, R. K. (2020). The blood-brain barrier and blood-tumour barrier in brain tumours and metastases. *Nat. Rev. Cancer* 20 (1), 26–41. doi:10.1038/s41568-019-0205-x

Bae, S., Liu, K., Pouliopoulos, A. N., Ji, R., Jiménez-Gambín, S., Yousefian, O., et al. (2024). Transcranial blood-brain barrier opening in Alzheimer's disease patients using a portable focused ultrasound system with real-time 2-D cavitation mapping. *Theranostics* 14 (11), 4519–4535. PMID: 39113808; PMCID: PMC11303073. doi:10.7150/thno.94206

Bae, S., Lee, S. A., Kim, S., Tsitsos, F., Liu, Y., and Konofagou, E. E. (2025). Ultrasound flow imaging for assessing cerebrovascular changes following focused-ultrasound blood-brain barrier opening. *Theranostics* 15 (18), 10028–10044. PMID: 41041070; PMCID: PMC12486155. doi:10.7150/thno.98098

Bartusik-Aebisher, D., Rogóż, K., and Aebisher, D. (2025). Nanoparticles for glioblastoma treatment. *Pharmaceutics* 17 (6), 688. PMID: 40574001; PMCID: PMC12196212. doi:10.3390/pharmaceutics17060688

Bellavance, M. A., Blanchette, M., and Fortin, D. (2008). Recent advances in blood-brain barrier disruption as a CNS delivery strategy. *AAPS J.* 10 (1), 166–177. Epub 2008 Mar 18. PMID: 18446517; PMCID: PMC2751463. doi:10.1208/s12248-008-9018-7

Butt, M. H., Zaman, M., Ahmad, A., Khan, R., Mallhi, T. H., Hasan, M. M., et al. (2022). Appraisal for the potential of viral and nonviral vectors in gene therapy: a review. *Genes (Basel)* 13 (8), 1370. PMID: 36011281; PMCID: PMC9407213. doi:10. 3390/genes13081370

Caro, C., Avasthi, A., Paez-Muñoz, J. M., Pernia Leal, M., and García-Martín, M. L. (2021). Passive targeting of high-grade gliomas *via* the EPR effect: a closed path for metallic nanoparticles? *Biomater. Sci.* 9 (23), 7984–7995. PMID: 34710207. doi:10.1039/d1bm01398j

Carpentier, A., Chauvet, D., Reina, V., Beccaria, K., Leclerq, D., McNichols, R. J., et al. (2012). MR-guided laser-induced thermal therapy (LITT) for recurrent glioblastomas. Lasers Surg. Med. 44 (5), 361–368. Epub 2012 Apr 5. PMID: 22488658. doi:10.1002/lsm. 22025

Chen, K. T., Chai, W. Y., Lin, Y. J., Lin, C. J., Chen, P. Y., Tsai, H. C., et al. (2021). Neuronavigation-guided focused ultrasound for transcranial blood-brain barrier opening and immunostimulation in brain tumors. *Sci. Adv.* 7 (6), eabd0772. PMID: 33547073; PMCID: PMC7864566. doi:10.1126/sciadv.abd0772

Chen, X., Momin, A., Wanggou, S., Wang, X., Min, H. K., Dou, W., et al. (2023). Mechanosensitive brain tumor cells construct blood-tumor barrier to mask chemosensitivity. *Neuron* 111 (1), 30–48.e14. doi:10.1016/j.neuron.2022.10.007

Chen, Y., Ma, Y., Shi, K., Chen, H., Han, X., Wei, C., et al. (2024). Self-disassembling and oxygen-generating porphyrin-lipoprotein nanoparticle for targeted glioblastoma resection and enhanced photodynamic therapy. *Adv. Mater* 36 (15), e2307454. Epub 2024 Feb 7. PMID: 38299428. doi:10.1002/adma.202307454

Choi, S., Yu, Y., Grimmer, M. R., Wahl, M., Chang, S. M., and Costello, J. F. (2018). Temozolomide-associated hypermutation in gliomas. *Neuro Oncol.* 20 (10), 1300–1309. doi:10.1093/neuonc/noy016

D'Amico, R. S., Aghi, M. K., Vogelbaum, M. A., and Bruce, J. N. (2021). Convection-enhanced drug delivery for glioblastoma: a review. *J. Neurooncol* 151 (3), 415–427. Epub 2021 Feb 21. PMID: 33611708; PMCID: PMC8034832. doi:10.1007/s11060-020-03408-9

Dahiya, S., Dahiya, R., and Hernández, E. (2021). Nanocarriers for anticancer drug targeting: recent trends and challenges. *Crit. Rev. Ther. Drug Carr. Syst.* 38 (6), 49–103. PMID: 34587429. doi:10.1615/CritRevTherDrugCarrierSyst.2021035650

Danhier, F., Ansorena, E., Silva, J. M., Coco, R., Le Breton, A., and Préat, V. (2012). PLGA-based nanoparticles: an overview of biomedical applications. *J. Control Release* 161 (2), 505–522. Epub 2012 Feb 4. PMID: 22353619. doi:10.1016/j.jconrel.2012.01.043

Das, D., Narayanan, D., Ramachandran, R., Gowd, G. S., Manohar, M., Arumugam, T., et al. (2023). Intracranial nanomedicine-gel with deep brain-penetration for glioblastoma therapy. *J. Control Release* 355, 474–488. Epub 2023 Feb 14. PMID: 36739909. doi:10.1016/j.jconrel.2023.01.085

Deb, V. K., and Jain, U. (2024).  $\rm Ti_3C_2$  (MXene), an advanced carrier system: role in photothermal, photoacoustic, enhanced drugs delivery and biological activity in cancer therapy. *Drug Deliv. Transl. Res.* 14 (11), 3009–3031. Epub 2024 May 7. PMID: 38713400. doi:10.1007/s13346-024-01572-3

Dutta, B., Barick, K. C., and Hassan, P. A. (2021). Recent advances in active targeting of nanomaterials for anticancer drug delivery. *Adv. Colloid Interface Sci.* 296, 102509. Epub 2021 Aug 18. PMID: 34455211. doi:10.1016/j.cis.2021.102509

Elzoghby, A. O., Samy, W. M., and Elgindy, N. A. (2012). Protein-based nanocarriers as promising drug and gene delivery systems. *J. Control Release* 161 (1), 38–49. Epub 2012 Apr 28. PMID: 22564368. doi:10.1016/j.jconrel.2012.04.036

Englander, Z. K., Wei, H. J., Pouliopoulos, A. N., Bendau, E., Upadhyayula, P., Jan, C. I., et al. (2021). Focused ultrasound mediated blood-brain barrier opening is safe and feasible in a murine pontine glioma model. *Sci. Rep.* 11 (1), 6521. PMID: 33753753; PMCID: PMC7985134. doi:10.1038/s41598-021-85180-y

Eroğlu, İ., and İbrahim, M. (2020). Liposome-ligand conjugates: a review on the current state of art. *J. Drug Target* 28 (3), 225–244. doi:10.1080/1061186X.2019.1648479

Eschbacher, K. L., Ida, C. M., Johnson, D. R., Alvi, M. A., Jenkins, S. M., Ruff, M. W., et al. (2021). Diffuse gliomas of the brainstem and cerebellum in adults show molecular heterogeneity. *Am. J. Surg. Pathol.* 45 (8), 1082–1090. doi:10.1097/PAS. 0000000000001690

Esposito, C. L., Nuzzo, S., Catuogno, S., Romano, S., de Nigris, F., and de Franciscis, V. (2018). STAT3 gene silencing by Aptamer-siRNA chimera as selective therapeutic for glioblastoma. *Mol. Ther. Nucleic Acids* 10, 398–411. Epub 2017 Dec 30. PMID: 29499951; PMCID: PMC5862137. doi:10.1016/j.omtn.2017.12.021

Ezike, T. C., Okpala, U. S., Onoja, U. L., Nwike, C. P., Ezeako, E. C., Okpara, O. J., et al. (2023). Advances in drug delivery systems, challenges and future directions. *Heliyon* 9 (6), e17488. PMID: 37416680; PMCID: PMC10320272. doi:10.1016/j.heliyon.2023. e17488

Ferreira, C., Ferreira, M. Y., Cardoso, L. J. C., Scarramal, J. P. L., Nogueira, A., Wong, T., et al. (2025). Safety and efficacy of selective and superselective intra-arterial cerebral infusion with blood-brain barrier disruption for glioma: a systematic review and meta-analysis. *J. Neurointerv Surg.* doi:10.1136/jnis-2025-024057

Fletcher, S. P., Chisholm, A., Lavelle, M., Guthier, R., Zhang, Y., Power, C., et al. (2024). A study combining microbubble-mediated focused ultrasound and radiation therapy in the healthy rat brain and a F98 glioma model. *Sci. Rep.* 14 (1), 4831. PMID: 38413663; PMCID: PMC10899261. doi:10.1038/s41598-024-55442-6

Gao, H., Xiong, Y., Zhang, S., Yang, Z., Cao, S., and Jiang, X. (2014). RGD and interleukin-13 peptide functionalized nanoparticles for enhanced glioblastoma cells and neovasculature dual targeting delivery and elevated tumor penetration. *Mol. Pharm.* 11 (3), 1042–1052. Epub 2014 Feb 19. PMID: 24521297. doi:10.1021/mp400751g

Gobbo, O. L., Sjaastad, K., Radomski, M. W., Volkov, Y., and Prina-Mello, A. (2015). Magnetic nanoparticles in cancer theranostics. *Theranostics* 5 (11), 1249–1263. PMID: 26379790; PMCID: PMC4568452. doi:10.7150/thno.11544

Griveau, A., Arib, C., Spadavecchia, J., and Eyer, J. (2022). Biological activity of gold nanoparticles combined with the NFL-TBS.40-63 peptide, or with other cell penetrating peptides, on rat glioblastoma cells. *Int. J. Pharm.* 4, 100129. PMID: 36164551; PMCID: PMC9508353. doi:10.1016/j.ijpx.2022.100129

Guo, H., and Mi, P. (2024). Polymer-drug and polymer-protein conjugated nanocarriers: design, drug delivery, imaging, therapy, and clinical applications. *Wiley Interdiscip. Rev. Nanomed Nanobiotechnol* 16 (4), e1988. PMID: 39109479. doi:10.1002/wnan.1988

Guo, J., Wang, M. F., Yuan, S. J., Li, K., Zhang, Q., Lei, H. M., et al. (2025). Photocontrolled co-delivery of verteporfin and acriflavine via platelets achieves potentiated glioblastoma-targeted photodynamic therapy. *J. Nanobiotechnology* 23 (1), 371. PMID: 40405165; PMCID: PMC12096713. doi:10.1186/s12951-025-03395-x

Hassan, A. A. A., Ramadan, E., Kristó, K., Regdon, G., Jr, and Sovány, T. (2025). Lipid-Polymer hybrid nanoparticles as a smart drug delivery system for peptide/protein delivery. *Pharmaceutics* 17 (6), 797. PMID: 40574109; PMCID: PMC12197309. doi:10. 3390/pharmaceutics17060797

Hillman, Y., Lustiger, D., and Wine, Y. (2019). Antibody-based nanotechnology. *Nanotechnology* 30 (28), 282001. Epub 2019 Mar 25. PMID: 30909177. doi:10.1088/1361-6528/ab12f4

Hu, D., Zou, J., Nie, S., Wang, Y., and Wang, S. (2024). *In vitro* study of a siRNA delivery liposome constructed with an ionizable cationic lipid. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* 49 (10), 1591–1600. doi:10.11817/j.issn.1672-7347.2024.230247

Huang, L., Asghar, S., Zhu, T., Ye, P., Hu, Z., Chen, Z., et al. (2021). Advances in chlorin-based photodynamic therapy with nanoparticle delivery system for cancer

treatment. Expert Opin. Drug Deliv. 18 (10), 1473–1500. Epub 2021 Jul 15. PMID: 34253129. doi:10.1080/17425247.2021.1950685

- Ibsen, S., Schutt, C. E., and Esener, S. (2013). Microbubble-mediated ultrasound therapy: a review of its potential in cancer treatment. *Drug Des. Devel Ther.* 7, 375–388. PMID: 23667309; PMCID: PMC3650568. doi:10.2147/DDDT.S31564
- Jiang, Z., Guan, J., Qian, J., and Zhan, C. (2019). Peptide ligand-mediated targeted drug delivery of nanomedicines. *Biomater. Sci.* 7 (2), 461–471. PMID: 30656305. doi:10. 1039/c8bm01340c
- Jin, Y., Zhang, B., Li, J., Guo, Z., Zhang, C., Chen, X., et al. (2025). Bioengineered protein nanocarrier facilitating siRNA escape from lysosomes for targeted RNAi therapy in glioblastoma. *Sci. Adv.* 11 (8), eadr9266. Epub 2025 Feb 19. PMID: 39970222; PMCID: PMC11838010. doi:10.1126/sciadv.adr9266
- Kadry, H., Noorani, B., and Cucullo, L. (2020). A blood-brain barrier overview on structure, function, impairment, and biomarkers of integrity. *Fluids Barriers CNS* 17 (1), 69. doi:10.1186/s12987-020-00230-3
- Kim, M., Yoon, H. J., Lee, C., Lee, M., Park, R. W., Lee, B., et al. (2024). Immune checkpoint-blocking nanocages cross the blood-brain barrier and impede brain tumor growth. *ACS Biomater. Sci. Eng.* 10 (1), 575–587. Epub 2023 Dec 27. PMID: 38150627; PMCID: PMC10777349. doi:10.1021/acsbiomaterials.3c01200
- Kluczka, J. (2023). Chitosan: structural and chemical modification, properties, and application. *Int. J. Mol. Sci.* 25 (1), 554. PMID: 38203726; PMCID: PMC10779193. doi:10.3390/ijms25010554
- Körbelin, J., Arrulo, A., and Schwaninger, M. (2024). Gene therapy targeting the blood-brain barrier. *Vitam. Horm.* 126, 191–217. Epub 2024 Apr 12. PMID: 39029973. doi:10.1016/bs.vh.2024.03.001
- Kumar, L., Verma, S., Utreja, P., and Kumar, D. (2023). Overview of inorganic nanoparticles: an expanding horizon in tumor therapeutics. *Recent Pat. Anticancer Drug Discov.* 18 (3), 343–363. PMID: 36200151. doi:10.2174/1574892817666221005094423
- Lakkadwala, S., and Singh, J. (2019). Co-delivery of doxorubicin and erlotinib through liposomal nanoparticles for glioblastoma tumor regression using an *in vitro* brain tumor model. *Colloids Surf. B Biointerfaces* 173, 27–35. Epub 2018 Sep 21. PMID: 30261346; PMCID: PMC6296250. doi:10.1016/j.colsurfb.2018.09.047
- Large, D. E., Abdelmessih, R. G., Fink, E. A., and Auguste, D. T. (2021). Liposome composition in drug delivery design, synthesis, characterization, and clinical application. *Adv. Drug Deliv. Rev.* 176, 113851. doi:10.1016/j.addr.2021.113851
- Li, C., Lai, C., Qiu, Q., Luo, X., Hu, L., Zheng, H., et al. (2019). Dual-ligand modification of PEGylated liposomes used for targeted doxorubicin delivery to enhance anticancer efficacy. *AAPS PharmSciTech* 20 (5), 188. doi:10.1208/s12249-019-1385-0
- Li, J., Xu, X., Zhao, Y., Wu, X., Chen, S., Xu, J., et al. (2025). Microglial membrane-coated biomimetic nanoplatform for enhanced blood-brain barrier penetration and targeted photodynamic therapy in orthotopic glioblastoma. *Adv. Healthc. Mater* 4, e02808. Epub ahead of print. PMID: 40904226. o. doi:10.1002/adhm.202502808
- Liu, H., Su, R., Qi, W., and Wang, Y. (2025). Cationic polymers for gene delivery: properties and functional optimization. *Chembiochem* 26 (10), e202500029. Epub 2025 Apr 4. PMID: 40127206. doi:10.1002/cbic.202500029
- Louis, D. N., Perry, A., Wesseling, P., Brat, D. J., Cree, I. A., Figarella-Branger, D., et al. (2021). The 2021 WHO classification of tumors of the central nervous system: a summary. *Neuro Oncol.* 23 (8), 1231–1251. doi:10.1093/neuonc/noab106
- Lu, Y. J., Lan, Y. H., Chuang, C. C., Lu, W. T., Chan, L. Y., Hsu, P. W., et al. (2020). Injectable thermo-sensitive chitosan hydrogel containing CPT-11-Loaded EGFR-targeted graphene oxide and SLP2 shRNA for localized drug/gene delivery in glioblastoma therapy. *Int. J. Mol. Sci.* 21 (19), 7111. PMID: 32993166; PMCID: PMC7583917. doi:10.3390/ijms21197111
- Maeda, H. (2015). Toward a full understanding of the EPR effect in primary and metastatic tumors as well as issues related to its heterogeneity. *Adv. Drug Deliv. Rev.* 91, 3–6. Epub 2015 Jan 9. PMID: 25579058. doi:10.1016/j.addr.2015.01.002
- McMahon, D., Poon, C., and Hynynen, K. (2019). Evaluating the safety profile of focused ultrasound and microbubble-mediated treatments to increase blood-brain barrier permeability. *Expert Opin. Drug Deliv.* 16 (2), 129–142. Epub 2019 Jan 29. PMID: 30628455; PMCID: PMC6576291. doi:10.1080/17425247.2019.1567490
- Mehta, A. M., Sonabend, A. M., and Bruce, J. N. (2017). Convection-enhanced delivery. *Neurotherapeutics* 14 (2), 358–371. PMID: 28299724; PMCID: PMC5398992. doi:10.1007/s13311-017-0520-4
- Meng, Y., Jones, R. M., Davidson, B., Huang, Y., Pople, C. B., Surendrakumar, S., et al. (2021). Technical principles and clinical workflow of transcranial MR-Guided focused ultrasound. *Stereotact. Funct. Neurosurg.* 99 (4), 329–342. Epub 2020 Dec 10. PMID: 33302282. doi:10.1159/000512111
- Meng, Y., Pople, C. B., Huang, Y., Jones, R. M., Ottoy, J., Goubran, M., et al. (2022). Putaminal recombinant glucocerebrosidase delivery with magnetic resonance-guided focused ultrasound in Parkinson's disease: a phase I study. *Mov. Disord.* 37 (10), 2134–2139. Epub 2022 Sep 11. PMID: 36089809. doi:10.1002/mds.29190
- Meng, J. L., Dong, Z. X., Chen, Y. R., Lin, M. H., Liu, Y. C., Roffler, S. R., et al. (2025). pH-Responsive polyethylene glycol engagers for enhanced brain delivery of PEGylated

- nanomedicine to treat glioblastoma. ACS Nano 19 (1), 307–321. Epub 2025 Jan 3. PMID: 39749925; PMCID: PMC11752499. doi:10.1021/acsnano.4c05906
- Merkel, S. F., Andrews, A. M., Lutton, E. M., Mu, D., Hudry, E., Hyman, B. T., et al. (2017). Trafficking of adeno-associated virus vectors across a model of the blood-brain barrier; a comparative study of transcytosis and transduction using primary human brain endothelial cells. *J. Neurochem.* 140 (2), 216–230. Epub 2016 Dec 15. PMID: 27718541; PMCID: PMC5298820. doi:10.1111/jnc.13861
- Mo, F., Pellerino, A., Soffietti, R., and Rudà, R. (2021). Blood-brain barrier in brain tumors: biology and clinical relevance. *Int. J. Mol. Sci.* 22 (23), 12654. PMID: 34884457; PMCID: PMC8657947. doi:10.3390/ijms222312654
- Molinaro, M., Skrodzki, D., and Pan, D. (2024). Chemoradiotherapy and nanomedicine: drug mechanisms and delivery systems. Wiley Interdiscip. Rev. Nanomed Nanobiotechnol 16 (4), e1984. PMID: 39109509. doi:10.1002/wnan.1984
- Narsinh, K. H., Kumar, K., Bankiewicz, K., Martin, A. J., Berger, M., Clarke, J., et al. (2025). A phase I study of convection-enhanced delivery (CED) of liposomal-irinotecan using real-time magnetic resonance imaging in patients with recurrent high-grade glioma. *J. Neurooncol* 172 (1), 219–227. Epub 2025 Jan 6. PMID: 39760796; PMCID: PMC11832582. doi:10.1007/s11060-024-04904-y
- de la Nava, D., Ausejo-Mauleon, I., Laspidea, V., Gonzalez-Huarriz, M., Lacalle, A., Casares, N., et al. (2024). The oncolytic adenovirus Delta-24-RGD in combination with ONC201 induces a potent antitumor response in pediatric high-grade and diffuse midline glioma models. *Neuro Oncol.* 26 (8), 1509–1525. PMID: 38554031; PMCID: PMC11300018. doi:10.1093/neuonc/noae066
- Nicholson, J. G., and Fine, H. A. (2021). Diffuse glioma heterogeneity and its therapeutic implications. *Cancer Discov.* 11 (3), 575–590. doi:10.1158/2159-8290. CD-20-1474
- Nivajärvi, R., Olsson, V., Hyppönen, V., Bowen, S., Leinonen, H. M., Lesch, H. P., et al. (2020). Detection of lentiviral suicide gene therapy in C6 rat glioma using hyperpolarised [1-<sup>13</sup> C]pyruvate. *NMR Biomed.* 33 (4), e4250. Epub 2020 Jan 7. PMID: 31909530. doi:10.1002/nbm.4250
- Nordling-David, M. M., Yaffe, R., Guez, D., Meirow, H., Last, D., Grad, E., et al. (2017). Liposomal temozolomide drug delivery using convection enhanced delivery. J. Control Release 261, 138–146. Epub 2017 Jun 27. PMID: 28666727. doi:10.1016/j. iconrel.2017.06.028
- Noury, H., Rahdar, A., Romanholo Ferreira, L. F., and Jamalpoor, Z. (2025). Aldriven innovations in smart multifunctional nanocarriers for drug and gene delivery: a mini-review. *Crit. Rev. Oncol. Hematol.* 210, 104701. Epub 2025 Mar 13. PMID: 40086770. doi:10.1016/j.critrevonc.2025.104701
- Obermeier, B., Daneman, R., and Ransohoff, R. M. (2013). Development, maintenance and disruption of the blood-brain barrier. *Nat. Med.* 19 (12), 1584–1596. doi:10.1038/nm.3407
- Pardridge, W. M. (2005). The blood-brain barrier: bottleneck in brain drug development.  $NeuroRx\ 2$  (1), 3–14. doi:10.1602/neurorx.2.1.3
- Pathak, R., Bhatt, S., Punetha, V. D., and Punetha, M. (2023). Chitosan nanoparticles and based composites as a biocompatible vehicle for drug delivery: a review. *Int. J. Biol. Macromol.* 253 (Pt 7), 127369. Epub 2023 Oct 13. PMID: 37839608. doi:10.1016/j.ijbiomac.2023.127369
- Qiao, G., Chu, C., Gulisashvili, D., Sharma, S., Kalkowski, L., Fadon-Padilla, L., et al. (2025). A safe MRI- and PET-guided method for increasing osmotic blood-brain barrier permeability. *Radiology* 316 (3), e243396. PMID: 40891972; PMCID: PMC12501621. doi:10.1148/radiol.243396
- Rapoport, S. I., and Robinson, P. J. (1986). Tight-junctional modification as the basis of osmotic opening of the blood-brain barrier. *Ann. N. Y. Acad. Sci.* 481, 250–267. doi:10.1111/j.1749-6632.1986.tb27155.x
- Sabel, M., Rommel, F., Kondakci, M., Gorol, M., Willers, R., and Bilzer, T. (2003). Locoregional opening of the rodent blood-brain barrier for paclitaxel using Nd:YAG laser-induced thermo therapy: a new concept of adjuvant glioma therapy? *Lasers Surg. Med.* 33 (2), 75–80. PMID: 12913878. doi:10.1002/lsm.10181
- Santoro, M., Tatara, A. M., and Mikos, A. G. (2014). Gelatin carriers for drug and cell delivery in tissue engineering. *J. Control Release* 190, 210–218. Epub 2014 Apr 16. PMID: 24746627; PMCID: PMC4142078. doi:10.1016/j.jconrel.2014.04.014
- Dos Santos Rodrigues, B., Kanekiyo, T., and Singh, J. (2019). ApoE-2 brain-targeted gene therapy through transferrin and penetratin tagged liposomal nanoparticles. *Pharm. Res.* 36 (11), 161. doi:10.1007/s11095-019-2691-7
- Sato, S., Kawase, T., Harada, S., Takayama, H., and Suga, S. (1998). Effect of hyperosmotic solutions on human brain tumour vasculature. *Acta Neurochir.* (Wien) 140 (11), 1135–1142. PMID: 9870058. doi:10.1007/s007010050227
- Sebak, A. A., El-Shenawy, B. M., El-Safy, S., and El-Shazly, M. (2021). From passive targeting to personalized nanomedicine: multidimensional insights on nanoparticles' interaction with the tumor microenvironment. *Curr. Pharm. Biotechnol.* 22 (11), 1444–1465. doi:10.2174/1389201021666201211103856
- Semyachkina-Glushkovskaya, O., Chehonin, V., Borisova, E., Fedosov, I., Namykin, A., Abdurashitov, A., et al. (2018). Photodynamic opening of the blood-brain barrier and pathways of brain clearing. *J. Biophot.* 11 (8), e201700287. Epub 2018 Mar 5. PMID: 29380947. doi:10.1002/jbio.201700287

Semyachkina-Glushkovskaya, O., Bragin, D., Bragina, O., Socolovski, S., Shirokov, A., Fedosov, I., et al. (2023). Low-level laser treatment induces the blood-brain barrier opening and the brain drainage system activation: delivery of liposomes into mouse glioblastoma. *Pharmaceutics* 15 (2), 567. PMID: 36839889; PMCID: PMC9966329. doi:10.3390/pharmaceutics15020567

Song, N., Zhang, J., Zhai, J., Hong, J., Yuan, C., and Liang, M. (2021). Ferritin: a multifunctional nanoplatform for biological detection, imaging diagnosis, and drug delivery. *Acc. Chem. Res.* 54 (17), 3313–3325. Epub 2021 Aug 20. PMID: 34415728. doi:10.1021/acs.accounts.1c00267

Song, B., Wang, X., Qin, L., Hussain, S., and Liang, W. (2024). Brain gliomas: diagnostic and therapeutic issues and the prospects of drug-targeted nano-delivery technology. *Pharmacol. Res.* 206, 107308. doi:10.1016/j.phrs.2024.107308

Souweidane, M. M., Bander, E. D., Zanzonico, P., Reiner, A. S., Manino, N., Haque, S., et al. (2025). Phase 1 dose-escalation trial using convection-enhanced delivery of radio-immunotheranostic 124I-Omburtamab for diffuse intrinsic pontine glioma. *Neuro Oncol.* 27 (8), 2117–2126. doi:10.1093/neuonc/noaf039

Stupp, R., Mason, W. P., van den Bent, M. J., Weller, M., Fisher, B., Taphoorn, M. J., et al. (2005). Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N. Engl. J. Med.* 352 (10), 987–996. doi:10.1056/NEJMoa043330

Sun, D., and Lu, Z. R. (2023). Structure and function of cationic and ionizable lipids for nucleic acid delivery. *Pharm. Res.* 40 (1), 27–46. doi:10.1007/s11095-022-03460-2

Sun, C., Lee, J. S. H., and Zhang, M. (2008). Magnetic nanoparticles in MR imaging and drug delivery. *Adv. Drug Deliv. Rev.* 60 (11), 1252–1265. Epub 2008 Apr 10. PMID: 18558452; PMCID: PMC2702670. doi:10.1016/j.addr.2008.03.018

Sun, T., Zhang, Y., Power, C., Alexander, P. M., Sutton, J. T., Aryal, M., et al. (2017). Closed-loop control of targeted ultrasound drug delivery across the blood-brain/tumor barriers in a rat glioma model. *Proc. Natl. Acad. Sci. U. S. A.* 114 (48), E10281–E10290. Epub 2017 Nov 13. PMID: 29133392; PMCID: PMC5715774. doi:10.1073/pnas. 1713328114

Tazhibi, M., McQuillan, N., Wei, H. J., Gallitto, M., Bendau, E., Webster, C. A., et al. (2024). Focused ultrasound-mediated blood-brain barrier opening is safe and feasible with moderately hypofractionated radiotherapy for brainstem diffuse midline glioma. *J. Transl. Med.* 22 (1), 320. PMID: 38555449; PMCID: PMC10981822. doi:10.1186/s12967-024-05096-9

van Tellingen, O., Yetkin-Arik, B., de Gooijer, M. C., Wesseling, P., Wurdinger, T., and de Vries, H. E. (2015). Overcoming the blood-brain tumor barrier for effective glioblastoma treatment. *Drug Resist Updat* 19, 1–12. doi:10.1016/j.drup. 2015.02.002

Tenchov, R., Sasso, J. M., and Zhou, Q. A. (2023). PEGylated lipid nanoparticle formulations: immunological safety and efficiency perspective. *Bioconjug Chem.* 34 (6), 941–960. doi:10.1021/acs.bioconjchem.3c00174

Tong, H., Ma, Z., Yu, J., Li, D., Zhu, Q., Shi, H., et al. (2025). Optimizing peptide-conjugated lipid nanoparticles for efficient siRNA delivery across the blood-brain barrier and treatment of glioblastoma multiforme. *ACS Chem. Biol.* 20 (4), 942–952. Epub 2025 Mar 13. PMID: 40080657. doi:10.1021/acschembio.5c00039

Torchilin, V. P. (2000). Drug targeting. Eur. J. Pharm. Sci. 11 (Suppl. 2), S81–S91. PMID: 11033430. doi:10.1016/s0928-0987(00)00166-4

Tyler, B., Gullotti, D., Mangraviti, A., Utsuki, T., and Brem, H. (2016). Polylactic acid (PLA) controlled delivery carriers for biomedical applications. *Adv. Drug Deliv. Rev.* 107, 163–175. Epub 2016 Jul 15. PMID: 27426411. doi:10.1016/j.addr.2016.06.018

Umemura, Y., Orringer, D., Junck, L., Varela, M. L., West, M. E. J., Faisal, S. M., et al. (2023). Combined cytotoxic and immune-stimulatory gene therapy for primary adult high-grade glioma: a phase 1, first-in-human trial. *Lancet Oncol.* 24 (9), 1042–1052. doi:10.1016/S1470-2045(23)00347-9

Xu, S., Tang, L., Li, X., Fan, F., and Liu, Z. (2020). Immunotherapy for glioma: current management and future application. *Cancer Lett.* 476, 1-12. doi:10.1016/j.canlet.2020.02.002

Xu, M., Qi, Y., Liu, G., Song, Y., Jiang, X., and Du, B. (2023). Size-dependent *in vivo* transport of nanoparticles: implications for delivery, targeting, and clearance. *ACS Nano* 17 (21), 20825–20849. Epub 2023 Nov 3. PMID: 37921488. doi:10.1021/acsnano.3c05853

Xuan, L., Ju, Z., Skonieczna, M., Zhou, P. K., and Huang, R. (2020). Nanoparticles-induced potential toxicity on human health: applications, toxicity mechanisms, and evaluation models. *MedComm* 4 (4), e327. PMID: 37457660; PMCID: PMC10349198. doi:10.1002/mco2.327

Yao, H., Wang, K., Wang, Y., Wang, S., Li, J., Lou, J., et al. (2015). Enhanced blood-brain barrier penetration and glioma therapy mediated by a new peptide modified gene delivery system. *Biomaterials* 37, 345–352. Epub 2014 Oct 25. PMID: 25453963. doi:10. 1016/j.biomaterials.2014.10.034

Yin, L., Pang, Y., Shan, L., and Gu, J. (2022). The *in vivo* pharmacokinetics of block copolymers containing polyethylene glycol used in nanocarrier drug delivery systems. *Drug Metab. Dispos.* 50 (6), 827–836. Epub 2022 Jan 22. PMID: 35066464. doi:10.1124/dmd.121.000568

Yuan, Z., Wang, B., Teng, Y., Ho, W., Hu, B., Boakye-Yiadom, K. O., et al. (2022). Rational design of engineered H-ferritin nanoparticles with improved siRNA delivery efficacy across an *in vitro* model of the mouse BBB. *Nanoscale* 14 (17), 6449–6464. PMID: 35416195. doi:10.1039/dlnr07880a

Zhan, W., and Wang, C. H. (2018). Convection enhanced delivery of chemotherapeutic drugs into brain tumour. *J. Control Release* 271, 74–87. Epub 2017 Dec 20. PMID: 29274437. doi:10.1016/j.jconrel.2017.12.020

Zhang, R., Kiessling, F., Lammers, T., and Pallares, R. M. (2023). Clinical translation of gold nanoparticles. *Drug Deliv. Transl. Res.* 13 (2), 378–385. Epub 2022 Aug 31. PMID: 36045273; PMCID: PMC9432795. doi:10.1007/s13346-022-01232-4

Zhang, H., Gao, X., Sun, Q., Dong, X., Zhu, Z., and Yang, C. (2024). Incorporation of poly(y-glutamic acid) in lipid nanoparticles for enhanced mRNA delivery efficiency in vitro and in vivo. Acta Biomater. 177, 361–376. Epub 2024 Feb 10. PMID: 38342193. doi:10.1016/j.actbio.2024.02.004

Zhao, Z., Ukidve, A., Krishnan, V., and Mitragotri, S. (2019). Effect of physicochemical and surface properties on *in vivo* fate of drug nanocarriers. *Adv. Drug Deliv. Rev.* 143, 3–21. Epub 2019 Jan 11. PMID: 30639257. doi:10.1016/j.addr. 2019.01.002

Zhao, X., Bai, J., and Yang, W. (2021). Stimuli-responsive nanocarriers for therapeutic applications in cancer. *Cancer Biol. Med.* 18 (2), 319–335. PMID: 33764711; PMCID: PMC8185873. doi:10.20892/j.issn.2095-3941.2020.0496