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Harnessing nature's pharmacy: investigating natural compounds as novel therapeutics for ulcerative colitis

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Backgrounds: Ulcerative colitis (UC) is a form of chronic inflammatory bowel disease, and UC diagnosis rates continue to rise throughout the globe. The research and development of new drugs for the treatment of UC are urgent, and natural compounds are an important source. However, there is a lack of systematic summarization of natural compounds and their mechanisms for the treatment of UC.

Methods: We reviewed the literature in the databases below from their inception until July 2023: Web of Science, PubMed, China National Knowledge Infrastructure, and Wanfang Data, to obtain information on the relationship between natural compounds and UC.

Results: The results showed that 279 natural compounds treat UC through four main mechanisms, including regulating gut microbiota and metabolites (Mechanism I), protecting the intestinal mucosal barrier (Mechanism II), regulating intestinal mucosal immune response (Mechanism III), as well as regulating other mechanisms (Mechanism IV) such as cellular autophagy modulation and ferroptosis inhibition. Of these, Mechanism III is regulated by all natural compounds. The 279 natural compounds, including 62 terpenoids, 57 alkaloids, 52 flavonoids, 26 phenols, 19 phenylpropanoids, 9 steroids, 9 saponins, 8 quinonoids, 6 vitamins, and 31 others, can effectively ameliorate UC. Of these, terpenoids, alkaloids, and flavonoids have the greatest potential for treating UC. It is noteworthy to highlight that a total of 54 natural compounds exhibit their therapeutic effects by modulating Mechanisms I, II, and III.

Conclusion: This review serves as a comprehensive resource for the pharmaceutical industry, researchers, and clinicians seeking novel therapeutic approaches to combat UC. Harnessing the therapeutic potential of these natural compounds may significantly contribute to the improvement of the quality of life of patients with UC and promotion of disease-modifying therapies in the future.

KEYWORDS

ulcerative colitis, natural compounds, gut microbiota, intestinal mucosal barrier, intestinal immune responses

1 Introduction

Ulcerative colitis (UC) is an idiopathic, chronic, inflammatory bowel disease (IBD) characterized by continuous inflammation starting from the rectum (Hoivik et al., 2013; Conrad et al., 2014; Nanki et al., 2019). World Health Organization has classified UC as a clinically intractable disease. Its global prevalence and incidence have been increasing with time; currently, its incidence and prevalence are 8–10 cases/100,000 subjects and 150–200 cases/100,000 subjects, respectively (da Silva et al., 2014; Ungaro et al., 2017). The annual UC treatment costs (direct and indirect) are estimated to be approximately US\$8.1–14.9 billion and €12.5–29.1 billion in the United States and Europe, respectively (Cohen et al., 2010).

UC is primarily treated with medicines, including aminosalicylates, immunomodulators, steroids, and biologics. However, due to potential adverse reactions and reduced efficiency of standard therapies, a comprehensive search for the identification of novel and natural medicines has been initiated to replace or complement present treatment options (Pastorelli et al., 2009; Wan et al., 2014). Many researchers are now turning to natural resources to seek effective compounds that can be used against UC (Cao et al., 2019).

Currently, there are some reviews on natural compounds and UC, such as summarizing some natural compounds or a class of compounds. These studies are significant for finding drugs for UC, but there is still a lack of systematic summaries. Therefore, this study

reviews the current progress made in the intervention of natural compounds in UC, and provides a complete overview of natural compounds and their mechanisms of action. More importantly, we hope that such a systematic summary will lead to important natural compounds and mechanisms of action for the treatment of UC. This review serves as a comprehensive resource for the pharmaceutical industry, researchers, and clinicians seeking novel therapeutic approaches to combat UC. Harnessing the therapeutic potential of these natural compounds may significantly contribute to the improvement of the quality of life of patients with UC and promotion of disease-modifying therapies in the future.

2 The etiology of UC

The most accepted hypothesis states that UC pathogenesis comprises complex communications between, external, immunological, and gut microbial factors in a genetically susceptible host (Figure 1) (Abraham et al., 2017; Glassner et al., 2020).

2.1 External factors

The diet structure of modern people is constantly changing. People are gradually consuming less dietary fiber, whereas

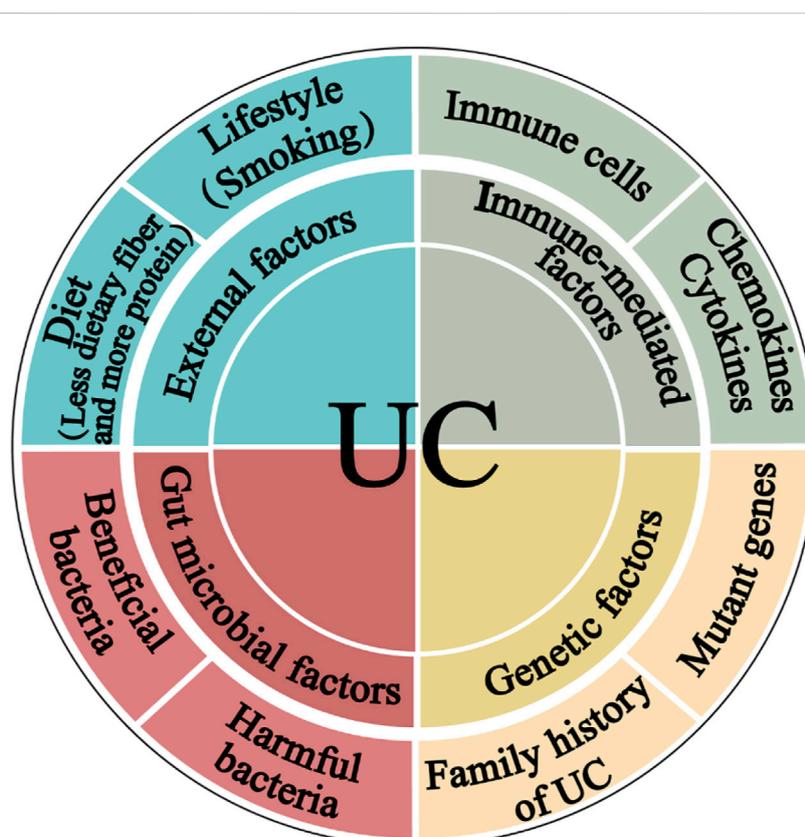


FIGURE 1

The pathogenesis of ulcerative colitis. The four main components linked to epithelial barrier abnormalities that drive the ulcerative colitis pathogenic mechanism are external factors, immune dysregulation, gut microbiota, and genetic inheritance.

increasing meat, egg, and milk product intake, is the main reason for the increase in the prevalence of UC. Furthermore, although smoking cigarettes is a critical Crohn's disease (CD) risk factor, quitting it has been linked to UC. According to a meta-analysis, smoking is more protective against UC than not smoking (Mahid et al., 2006). UC individuals who smoked had a milder disease course than non-smokers. UC is harsher for those who stop smoking. It may be mediated by carbon monoxide that can suppress interleukin-10 (IL-10) through a heme oxygenase (HO)-1-dependent pathway in UC mice (Sheikh et al., 2011).

2.2 Immune-mediated factors

The immune response is intricately associated with the pathophysiology of UC. The buildup of innate lymphoid cells (ILC), natural killer (NK) cells, macrophages, dendritic cells, neutrophils, and abnormal T and B cells inside the intestinal mucosa, along with the production of chemokines and cytokines that may trigger an inflammatory response. This inflammatory process can lead to the disruption of the intestinal mucosa and ultimately result in the development of UC (Liu Y. et al., 2022).

2.3 Gut microbial factors

The gut microbiota directly impacts the maintenance of homeostasis in the intestinal pro-inflammatory and anti-inflammatory responses. Germ-free conditions prevent the development of colitis in genetically susceptible mice (Veltkamp et al., 2001). Moreover, the introduction of proinflammatory bacteria or microbiota from patients with UC into healthy mice can induce inflammation (Ohkusa et al., 2003), while colonization of mice with intestinal microbiota from donors with IBD exacerbates colitis by modulating immune responses (Britton et al., 2019).

2.4 Genetic factors

Genetic factors have also been linked with UC. 12% of UC patients have a family history of IBD (Childers et al., 2014). Genome-wide association studies have identified 200 risk loci for IBD to date, with most genes contributing to both UC and CD phenotypes (Jostins et al., 2012; Liu et al., 2015). Examples of loci associated with increased UC susceptibility include human leukocyte antigen and genes associated with barrier function, such as HNF4A and CDH1 (Consortium et al., 2009). In addition, with increasing knowledge about UC pathogenesis, natural compounds have become a research hotspot because of their more efficient application prospects for preventing and mitigating UC occurrence and development.

3 The mechanism of natural compounds in intervention UC

We reviewed the scientific papers in the databases below from their inception to July 2023 to identify the studies relevant to the

mechanism and activity of natural compounds against UC: PubMed, Web of Science, Wanfang Data, and the China National Knowledge Infrastructure. The present study provides a comprehensive summary of 279 natural compounds demonstrated to treat UC through various mechanisms primarily. These mechanisms include regulating gut microbiota and metabolites (Mechanism I), protecting the intestinal mucosal barrier (Mechanism II), regulating intestinal mucosal immune response (Mechanism III), as well as the other mechanisms (Mechanism IV) such as cellular autophagy modulation and ferroptosis inhibition (as depicted in Figure 2; Supplementary Table S1). It is noteworthy to highlight that Mechanism III is regulated by all natural compounds; Mechanisms II and III can be modulated by at least half of the compounds. Research on these mechanisms may give information on the etiology of UC.

3.1 Regulating gut microbiota and metabolites

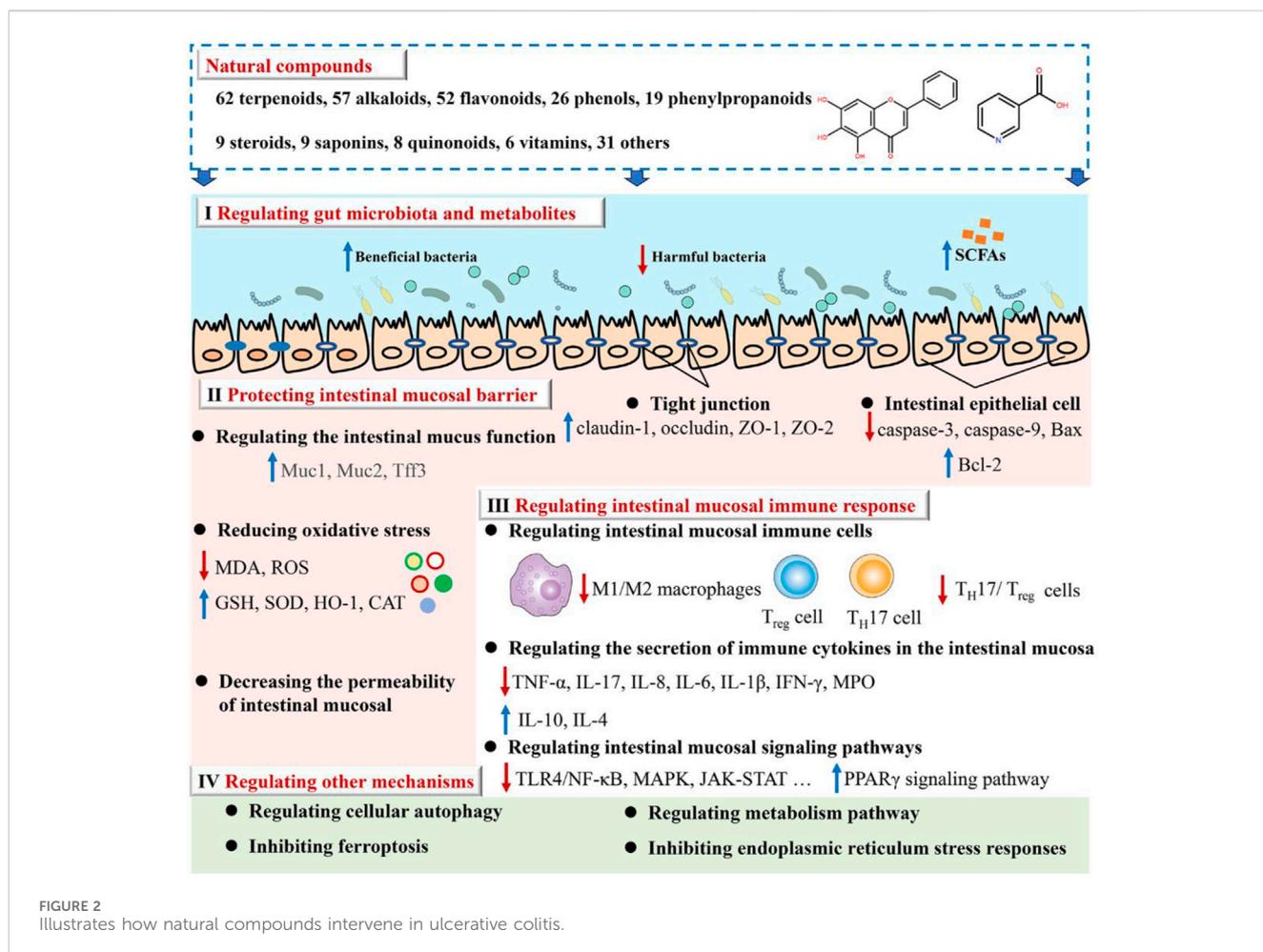
The available evidence indicates that UC is an increased immune response in the mucosal lining, which is triggered by an imbalance in particular gut bacteria. This condition is defined by an abnormal composition of the microbiota and the presence of bacterial products. According to the data shown in Supplementary Table S1, there has been extensive research conducted on natural compounds to investigate their prebiotic qualities. These compounds have been found to have an impact on the makeup of the microbiota and its metabolites, as well as the prevention of colonization by intestinal pathogens and the reduction of the risk of recurrence of ulcerative colitis, as illustrated in Figure 3.

3.1.1 Enhancing the abundance and diversity of gastrointestinal microbes

Patients with UC exhibit a diminished diversity of gut microbes, and an imbalance in the composition of the microbiome continues throughout the progression of the disease (Fang et al., 2021). Microbial diversity and community richness of species are reflected by Shannon and Simpson's indexes and abundance-based coverage estimator (ACE) and Chao1 indexes, respectively (Wang et al., 2022). Research shows that atractylenolide I (Qu et al., 2022), kaempferol (Qu et al., 2021), and 6-shogaol (Wei et al., 2022) indicated a substantial elevation of the Shannon, Simpson, and Chao1 indexes in the UC mice intestinal flora. Ginkgolide C (Xu et al., 2022), hydroxytyrosol (Miao, 2022), jatrorrhizine (Zhang et al., 2022), luteolin (Li et al., 2021), and sauchinone (Wu et al., 2023) enhanced the diversity and abundance of UC intestinal flora by increasing Chao1 and Shannon indexes. Whereas apigenin (Fu et al., 2022), berberine (Wei et al., 2022), docosapentaenoic acid (Dong et al., 2022a), ginsenoside Rg1 (Cheng et al., 2022; Long et al., 2022), and 2,3,5,4'-tetrahydroxystilbene-2-O- β -D-glucoside (He et al., 2021) elevated the abundance and diversity by Chao1 and ACE indexes upregulation.

3.1.2 Regulating the composition of gut microbiota

The composition of gut microbiota in individuals with UC or animal models has exhibited considerable heterogeneity across different studies. In general, when comparing the microbiota of



individuals or animals in good health, it is observed that there is a reduction in the abundance of beneficial bacteria and an increase in the prevalence of harmful bacteria. Seven alkaloids [aspartate (Hu et al., 2022), berberine (Li et al., 2022a), cepharanthine (Wang et al., 2022), evodiamine (Wang et al., 2020), jatrorrhizine (Zhang et al., 2022), matrine (Yao et al., 2021), and sanguinarine (Li et al., 2022b)], 15 flavonoids [acacetin (Ren et al., 2021), apigenin (Fu et al., 2022), astragaln (Peng et al., 2020), corylin (Wang et al., 2023), galangin (Xuan et al., 2020), icariin (Zhang et al., 2021a), kaempferol (Qu et al., 2021), luteolin (Li et al., 2021), licoflavone B (Zhang et al., 2022b), licochalcone A (Zhang et al., 2021), α -mangostin (Gutierrez-Orozco et al., 2014), naringin (Cao et al., 2021), pinocembrin (Yue et al., 2020), phloretin (Wu et al., 2019), and vitexin (Zhang et al., 2022c)], 6 phenols [hydroxytyrosol (Miao, 2022), methyl gallate (Zhou et al., 2022), paeonol (Zheng et al., 2022), prim-O-Glucosylcimifugin (Yin et al., 2022), salidroside (Liu et al., 2023), and 6-shogaol (Wei et al., 2022)], 2 quinonoids [juglone (Hua et al., 2021), rhein (Dong et al., 2022)], 8 terpenoids [atractylenolide-1 (Qu et al., 2022), aromatic-turmerone (Li et al., 2022), bilobalide (Zhang et al., 2021c), β -caryophyllene (Yeom et al., 2022), ginkgolide C (Xu et al., 2022), β -patchoulene (Liu et al., 2020), triptolide (Wu et al., 2020) and terpinen-4-ol (Zhang et al., 2017a)], 2 phenylpropanoids [caffeic acid (Zhang et al., 2016), chlorogenic acid (Niu et al., 2022)] could stimulate the propagation of beneficial bacteria and reduce some pathogenic bacteria. For instance, the administration of

berberine in mice with DSS-induced UC has been found to induce a range of protective effects (Zhang et al., 2017; Jiang et al., 2021; Zheng et al., 2021; Li et al., 2022a). These effects include the mitigation of colon inflammation and oxidative stress, restoration of the epithelial barrier's functionality, and improvement of the gut microenvironment. Specifically, berberine supplementation has been observed to increase the abundance of Bacillibacteria, *Bacteroides fragilis*, Eubacterium, *Lactobacillales*, and *Lactobacillus/Lactococcus*. Conversely, it has been found to decrease the levels of *Akkermansia muciniphila*, *Bacteroides*, *Desulfovibrio*, Enterobacteriaceae, Segmented filamentous bacteria, Verrucomicrobiae, and Verrucomicrobiales.

3.1.3 Regulating the gut microbiota-metabolites

The metabolites of gut microbiota, including tryptophan, bile acids, and short-chain fatty acids (SCFAs), affect UC development. Most of the current research has focused primarily on the effects of SCFAs. The research indicates reduced SCFA-producing bacteria, including *Clostridium clusters IV and XIVb*, *Faecalibacterium*, *Leuconostocaceae*, *Odoribacter*, and *Roseburia* in UC patients (Kostic et al., 2014). Moreover, recently many natural compounds, for instance, apigenin (Fu et al., 2022), baicalin (Zhu et al., 2020), evodiamine (Shen et al., 2019), galangin (Xuan et al., 2020), ginsenoside Rg1 (Long et al., 2022), hydroxytyrosol (Miao, 2022), octacosanol (Miao et al., 2022), pinocembrin (Hu et al., 2019),

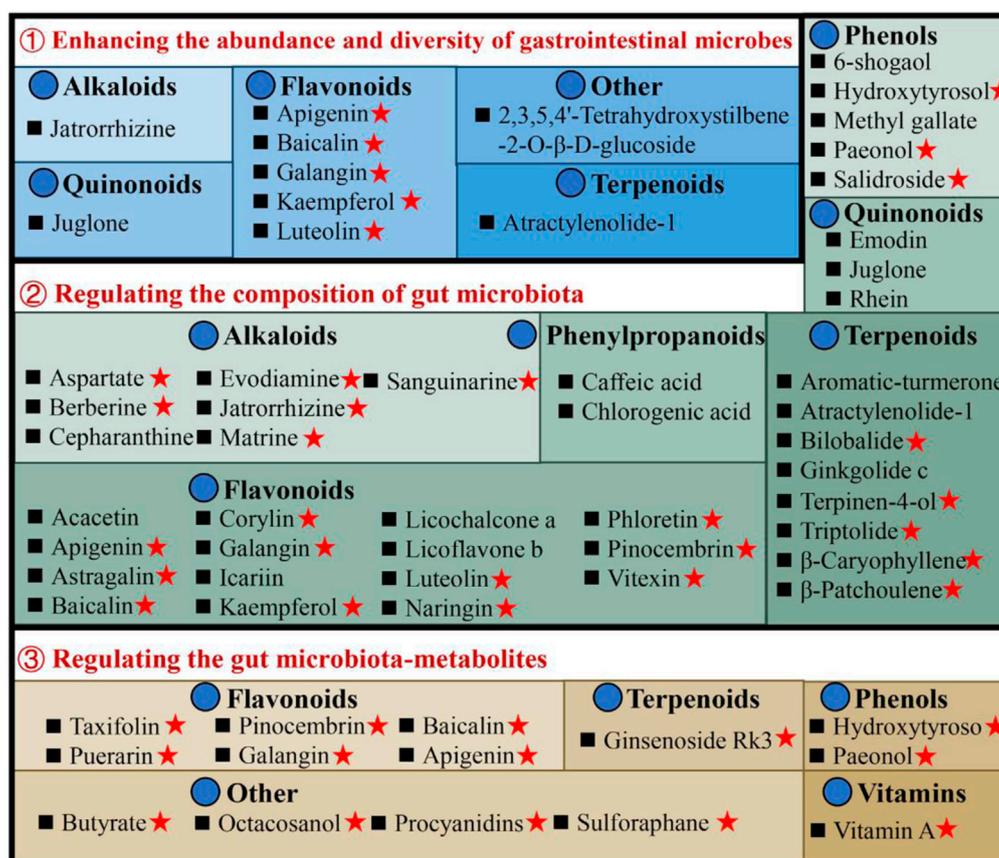


FIGURE 3

Illustrates the utilization of natural compounds in the management of ulcerative colitis through the modulation of gut microbiota and metabolites. Red pentagrams indicate compounds involved in Mechanism I, II, and III. The natural compounds involved in this paper are shown in Supplementary Table S1.

paeonol (Zheng et al., 2022), procyanidins (Huang et al., 2022), sulforaphane (Zhang et al., 2020), and vitamin A (Pang et al., 2021), could increase SCFA-producing bacteria in UC models. For example, taxifolin can ameliorate DSS-induced colitis by altering gut microbiota to increase the production of SCFAs (Li et al., 2022). Furthermore, SCFAs function by stimulating G-protein-coupled receptors (GPCR) and suppressing histone deacetylases (Kostic et al., 2014). It is reported that taxifolin can increase the level of GPR41 and GPR43 in the colon, and increase the level of the content of SCFAs, thereby reducing DSS-induced intestinal inflammatory reaction and protecting the intestinal mucosa (Li et al., 2022).

The present study has specifically examined the impact of pharmaceutical substances on the composition and diversity of the gastrointestinal microbiota. However, the gut microbiota exerts a significant influence on the chemical alteration, pharmacological action, and metabolic mechanisms of natural compounds (Zhao et al., 2022a). Certain gut microorganisms possess the ability to break down and convert organic substances, resulting in the production of metabolites and functional chemicals that exhibit physiological actions that are not naturally generated by the host organism (Koppel et al., 2017). There is currently a significant amount of research being dedicated to comprehending the distinct ways in which microorganisms alter natural products and the consequent effects of these metabolites on the health of the

host organism (Luca et al., 2020). This is a matter that warrants further investigation in our research.

3.2 Protecting intestinal mucosal barrier

The intestinal mucosal barrier damage is a crucial UC characteristic (Ungaro et al., 2017). Complete healing of intestinal mucosa is the most desired goal in UC treatment (Du et al., 2020). As shown in Supplementary Table S1, natural compounds can improve the barrier function of the UC mucosa through multiple perspectives, these include upregulation of the expression of tight junction protein, reduction in the intestinal mucosal permeability, regulation of the intestinal mucus function, reduction of oxidative stress, and protection of the intestinal epithelial cells (Figure 4).

3.2.1 Upregulation of the tight junction protein expression

The tight junctions (TJs) are present between epithelial cells in the junctions' apical region and comprise multiple proteins, such as claudins, junctional adhesion molecules, occludin, and tricellulin (Pan et al., 2023). TJs are an intestinal mucosal mechanical barrier required for the maintenance of intestinal epithelium integrity and

① Up-regulation of the tight junction protein expression								
Alkaloids ■ Berberine★ ■ Berberine hydrochloride★ ■ Berberrubine ■ Coptisine ■ Dihydroberberine ■ Ergothioneine ■ Evodiamine★ ■ Heterophyllin B★ ■ Jatrorrhizine ■ Matrine★ ■ Nigakinone ■ Nigeglanine ■ Palmatine★ ■ Piperine		Terpenoids ■ Andrographolide★ ■ Astragaloside IV ■ Atractylenolide III ■ Betulinic acid hydroxamate ■ Bilobalide★ ■ Bryodulcosigenin ■ Carnosol ■ Celastrol★ ■ Dihydroartemisinin ■ Epoxymicheliolide ■ Geniposide ■ Ginkgolide C★ ■ Ginsenoside Rk3★ ■ Loganin ■ Mogrol ■ Morroniside ■ Patchouli alcohol ■ Plumericin ■ Sauchinone★ ■ Terpinen-4-ol★ ■ Triptolide★ ■ β-Patchoulene★		Phenylpropanoids ■ Acteoside ■ Honokiol ■ Phillygenin ■ Rosmarinic acid ■ Schisandrin B ■ Sinapic acid ■ Wedelolactone				
Flavonoids ■ Alpinetin ■ Apigenin★ ■ Astragalin★ ■ Baicalin★ ■ Desmethybellidifolin★ ■ Eriodictyol ■ Eupatilin ■ Hesperidin ■ Hyperoside ■ Isovitexin ■ Kaempferol★ ■ Licochalcone A★ ■ Luteolin★ ■ Myricetin ■ Naringin★ ■ Phloretin★ ■ Pinocembrin★ ■ Puerarin★ ■ Tiliroside ■ Vitexin★ ■ Wogonoside		Other ■ 2,3,5,4'-Tetrahydroxystilbene ■ -2-O-β-D-glucoside ■ 4-Geranyloxy-2,6-dihydroxybenzophenone ■ 6,7-Dihydroxy-2,4-dimethoxyphenanthrene ■ Arbutin ■ Octacosanol ■ Phytic acid ■ Sodium houttuyfonate★ ■ Sulforaphane★		Quinonoids ■ Aloin A ■ Barbaloin ■ Diacetylrhein	Saponins ■ Dioscin ■ Ginsenoside compound K★			
Alkaloids ■ Berberine★		Phenols ■ Cannabidivarin★	Quinonoids ■ Aloin A	Saponins ■ Platycodin D ■ Ginsenoside compound K★	Flavonoids ■ Luteolin★ ■ Vitexin★			
② Decreased permeability of intestinal mucosal								
Alkaloids ■ Berberine★ ■ Berberrubine ■ Dihydroberberine ■ Heterophyllin B★ ■ Matrine★ ■ Palmatine★				Flavonoids ■ Astragalin★ ■ Catechin-7-O-β-D-glucopyranoside ■ Hesperidin ■ Hyperoside ■ Kaempferol★ ■ Phloretin★ ■ Puerarin★ ■ Vitexin★	Terpenoids ■ Betulinic acid hydroxamate ■ Carnosol ■ Celastrol★ ■ Dehydrocostus Lactone ■ Loganin ■ Morroniside ■ Patchouli alcohol	Phenylpropanoids ■ Honokiol ■ Rosmarinic acid	Phenols ■ Epigallocatechin gallate ■ Zingerone★	Steroids ■ Ginsenoside Rg3★
Other ■ Alanyl-glutamine		Saponins ■ Dioscin ■ Ginsenoside compound K★ ■ Saikosaponin D★		Quinonoids ■ Aloin A	Vitamins ■ Vitamin A★			
Alkaloids ■ 8-Oxypalmatine ■ Alliin ■ Berberine★ ■ Betaine ■ Boldine ■ Cavidine ■ Glutamate ■ Glutamine ■ Indirubin ■ Isatin ■ Matrine★ ■ Melittin ■ Palmitoylethanolamide ■ Piperine ■ Sanguinarine★ ■ Sinigrin				Flavonoids ■ Alpinetin ■ Apigenin★ ■ Baicalin★ ■ Cardamonin ■ Casticin ■ Diosmin★ ■ Eriodictyol ■ Eupatilin ■ Flavocoxid ■ Galangin★ ■ Hesperidin ■ Hesperidin methyl chalcone ■ Myricetin ■ Naringin★ ■ Phloretin★ ■ Puerarin★ ■ Quercetin★	Terpenoids ■ Asperuloside ■ Astragaloside IV ■ Atractylenolide III ■ Bruceine D ■ Carnosic acid ■ Epoxymicheliolide ■ Geniposide ■ Ginsenoside Rd ■ Linalool ■ Lycopene ■ Perillyl alcohol ■ Sericic acid ■ Stevioside ■ Triptolide★ ■ Ursolic acid ■ Zeaxanthin ■ β-Caryophyllene★	Phenylpropanoids ■ Acteoside ■ Arctigenin ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Ferulic Acid ■ Imperatorin ■ Phillygenin ■ Rosmarinic acid ■ Sesamin ■ Sinapic acid		
Other ■ 1,25-Dihydroxyvitamin D3 ■ 3,4-Oxo-isopropylidene-shikimic acid ■ Albiflorin ■ Apocynin ■ D-Pinitol ■ Erianin		Phenols ■ 10-Gingerol ■ 6-Gingerol ■ 6-Paradol ■ 8-Gingerol ■ Curcumin★ ■ Epicatechin ■ Zingerone★	Phenylpropanoids ■ Epigallocatechin gallate ■ Hydroxytyrosol★ ■ Polydatin ■ Resveratrol ■ Thymol	Phenylpropanoids ■ Acteoside ■ Arctigenin ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Ferulic Acid ■ Imperatorin ■ Phillygenin ■ Rosmarinic acid ■ Sesamin ■ Sinapic acid				
③ Regulation of the intestinal mucus function								
Alkaloids ■ Berberine★ ■ Berberrubine ■ Dihydroberberine ■ Heterophyllin B★ ■ Matrine★ ■ Palmatine★		Flavonoids ■ Astragalin★ ■ Catechin-7-O-β-D-glucopyranoside ■ Hesperidin ■ Hyperoside ■ Kaempferol★ ■ Phloretin★ ■ Puerarin★ ■ Vitexin★	Terpenoids ■ Betulinic acid hydroxamate ■ Carnosol ■ Celastrol★ ■ Dehydrocostus Lactone ■ Loganin ■ Morroniside ■ Patchouli alcohol	Phenylpropanoids ■ Honokiol ■ Rosmarinic acid	Phenols ■ Epigallocatechin gallate ■ Zingerone★			
Other ■ Alanyl-glutamine		Saponins ■ Dioscin ■ Ginsenoside compound K★ ■ Saikosaponin D★		Quinonoids ■ Aloin A	Vitamins ■ Vitamin A★ ■ Vitamin E			
④ Oxidative stress reduction								
Alkaloids ■ 8-Oxypalmatine ■ Alliin ■ Berberine★ ■ Betaine ■ Boldine ■ Cavidine ■ Glutamate ■ Glutamine ■ Indirubin ■ Isatin ■ Matrine★ ■ Melittin ■ Palmitoylethanolamide ■ Piperine ■ Sanguinarine★ ■ Sinigrin		Flavonoids ■ Alpinetin ■ Apigenin★ ■ Baicalin★ ■ Cardamonin ■ Casticin ■ Diosmin★ ■ Eriodictyol ■ Eupatilin ■ Flavocoxid ■ Galangin★ ■ Hesperidin ■ Hesperidin methyl chalcone ■ Myricetin ■ Naringin★ ■ Phloretin★ ■ Puerarin★ ■ Quercetin★	Terpenoids ■ Asperuloside ■ Astragaloside IV ■ Atractylenolide III ■ Bruceine D ■ Carnosic acid ■ Epoxymicheliolide ■ Geniposide ■ Ginsenoside Rd ■ Linalool ■ Lycopene ■ Perillyl alcohol ■ Sericic acid ■ Stevioside ■ Triptolide★ ■ Ursolic acid ■ Zeaxanthin ■ β-Caryophyllene★	Phenylpropanoids ■ Acteoside ■ Arctigenin ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Ferulic Acid ■ Imperatorin ■ Phillygenin ■ Rosmarinic acid ■ Sesamin ■ Sinapic acid				
Other ■ 1,25-Dihydroxyvitamin D3 ■ 3,4-Oxo-isopropylidene-shikimic acid ■ Albiflorin ■ Apocynin ■ D-Pinitol ■ Erianin		Phenols ■ 10-Gingerol ■ 6-Gingerol ■ 6-Paradol ■ 8-Gingerol ■ Curcumin★ ■ Epicatechin ■ Zingerone★	Phenylpropanoids ■ Epigallocatechin gallate ■ Hydroxytyrosol★ ■ Polydatin ■ Resveratrol ■ Thymol	Phenylpropanoids ■ Acteoside ■ Arctigenin ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Ferulic Acid ■ Imperatorin ■ Phillygenin ■ Rosmarinic acid ■ Sesamin ■ Sinapic acid				
⑤ Protecting the intestinal epithelial cells								
Alkaloids ■ Berberine★ ■ Berberrubine ■ Betaine ■ Coptisine ■ Cyclosporine ■ Evodiamine★ ■ Fumigaclavine C ■ Glutamate ■ Glutamine ■ Indirubin ■ Isatin ■ Neferine ■ Nigakinone ■ Nigeglanine ■ Palmatine★ ■ Piperine ■ Sanguinarine★ ■ Serine ■ Sinomenine hydrochloride		Flavonoids ■ Alpinetin ■ Apigenin★ ■ Baicalin★ ■ Diosmin★ ■ Eriodictyol ■ Flavocoxid ■ Genistein ■ Hesperidin ■ Hesperidin methyl chalcone ■ Luteolin★ ■ Naringin★ ■ Phloretin ■ Wogonin	Steroids ■ Diosgenin ■ Ginsenoside Rg3★ ■ Taraxasterol ■ Taurine ■ Taurohydoxycholic acid	Phenylpropanoids ■ Acteoside ■ Chlorogenic Acid★ ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Schisandrin B ■ Sinapic acid	Other ■ 6,7-Dihydroxy-2,4-dimethoxyphenanthrene ■ Alanyl-glutamine ■ Arbutin ■ Garlicin ■ Nervonic acid			
Terpenoids ■ Artemisinin ■ Artesunate ■ Betulin ■ Brusatol ■ Bryodulcosigenin ■ Carnosic acid ■ Celastrol★ ■ Cycloastragenol ■ Geniposide ■ Ginsenoside Rd ■ Ginsenoside Rk3★ ■ Paeoniflorin★ ■ Patchouli alcohol ■ Picroside II ■ Plumericin ■ Terpinen-4-ol★ ■ Zeaxanthin		Phenols ■ Polydatin ■ Hydroxytyrosol★ ■ Gallic acid ■ Salidroside★ ■ Gallotannin corilagin	Quinonoids ■ Aloin A ■ Diacetylrhein	Vitamins ■ Vitamin D3				

FIGURE 4 Natural compounds against ulcerative colitis via intestinal mucosal barrier protection. Red pentagrams indicate compounds involved in Mechanism I, II, and III. The natural compounds involved in this paper are shown in Supplementary Table S1.

intestinal mucosal permeability by modulating ions and molecules' entrance into the paracellular channels (Suzuki, 2013). The destruction or reduction of TJ proteins can disrupt the gastrointestinal mucosal barrier, causing UC and other intestinal disorders. However, some natural compounds, such as 8 alkaloids [berberine hydrochloride (Zhu et al., 2019a), berberrubine (Yu et al., 2018), coptisine (Wang et al., 2021a), dihydroberberine (Li et al., 2021), evodiamine (Shen et al., 2019), matrine (Yan et al., 2020), nigakinone (Liu et al., 2023), and piperine (Guo et al., 2020)], 8 flavonoids [apigenin (Fu et al., 2022), kaempferol (Qu et al., 2021), licochalcone A (Zhang et al., 2021), licoflavone B (Zhang et al., 2022b), phloretin (Zhang Z. et al., 2019), pinocembrin (Hu et al., 2019), puerarin (Wu et al., 2020), and wogonoside (Huang et al., 2020)], 2 phenylpropanoids [honokiol (Wang and Wang, 2022), sinapic acid (Qian et al., 2020)], 5 terpenoids [carnosol (Xu et al., 2022b), ginsenoside Rk3 (Tian et al., 2020), patchouli alcohol (Wu et al., 2020), plumericin (Rapa et al., 2021), and sauchinone (Wu et al., 2023)], arbutin (Zhang et al., 2021), sodium houttuynifonate (Cheng et al., 2023), can promote TJ proteins expression in UC animals, such as claudin-1, occludin, and zona occludens 1 (ZO-1), thus efficiently prevent the paracellular permeability disruption. Additionally, berberrubine (Yu et al., 2018), coptisine (Wang et al., 2021a), dihydroberberine (Li et al., 2021), patchouli alcohol (Wu et al., 2020), and palmatine (Zhang et al., 2018) promote ZO-2 protein levels in UC animals. Multiple researches are investigating UC alleviation by alkaloids and flavonoids, which upregulate TJ proteins.

3.2.2 Decreased permeability of intestinal mucosal

The permeability of the intestinal mucosa controls the transport of molecular substances across the epithelium of the intestinal mucosa by the process of simple diffusion. Increased mucosal permeability (Nakarai et al., 2012) has been reported in UC patients (Wang et al., 2022), allowing the entrance of intestinal pathogens as well as their toxic metabolites in the liver, lymph, peripheral tissues, and blood, causing enhanced oxidative stress and inflammation. Intestinal permeability allows accurate, direct, and quantitative evaluation of the colonic epithelial barrier (Huang et al., 2016). Generally, FITC-dextran (fluorescein isothiocyanate dextran) permeability is utilized for the elucidation of epithelium integrity. It has recently been revealed that after taking FITC-dextran orally, the serum of DSS mice had markedly increased FITC-dextran levels (Zhang et al., 2022c). Interestingly, berberine (Zheng et al., 2021), cannabidiol (Pagano et al., 2019), dioscin (Cai et al., 2021), ginsenoside compound K (Wang et al., 2022), luteolin (Xie et al., 2022), platycodin D (Guo et al., 2021), vitexin (Zhang et al., 2022c), and wogonoside (Huang et al., 2020), can decrease serum FITC-dextran level in UC animals.

3.2.3 Regulation of the intestinal mucus function

The structure of the intestinal mucus is composed of the glycoprotein network containing host-specific glycan that prevents the interaction of bacteria and epithelium, inhibits infection, and modulates the balance between exogenous stimulation and immune function (Johansson et al., 2014). Intestinal mucosal layer dysfunction compromises intestinal epithelium integrity and enhances pathogenic susceptibility. The main intestinal mucosal component is mucin (Johansson et al.,

2011). During active UC, there are decreased goblet cells in the colon epithelium, the protective mucus layer thickness reduces, and the mucus levels in mucin, glycosylation, and phosphatidylcholine alters. Alterations in the levels of colon proteins, such as trefoil factor 3 (Tff3), mucin 1 (Muc1), and Muc2, increase susceptibility to chronic inflammation, indicating mucins' importance in intestinal barrier repair. The research suggests that 7 flavonoids [astragalin (Peng et al., 2020), catechin-7-O- β -D-glucopyranoside (Kook et al., 2015), hyperoside (Cheng et al., 2021), kaempferol (Park et al., 2012), puerarin (Wu et al., 2020), phloretin (Wu et al., 2019), and vitexin (Zhang et al., 2022c)], 8 alkaloids [berberrubine (Yu et al., 2018), berberine (Dong et al., 2022c), dihydroberberine (Li et al., 2021), evodiamine (Wang et al., 2020), heterophyllin B (Chen et al., 2022), matrine (Yan et al., 2020), palmatine (Zhang et al., 2018), and tryptophan (Islam et al., 2017)], 8 terpenoids [betulinic acid hydroxamate (Prados et al., 2021), carnosol (Xu et al., 2022b), celastrol (Li et al., 2022), dehydrocostus lactone (Zhou et al., 2020), ginsenoside Rg3 (Liu et al., 2023), loganin (Yuan et al., 2020), morroniside (Yuan et al., 2020), and patchouli alcohol (Wu et al., 2020)], 2 phenols [epigallocatechin gallate (Diwan and Sharma, 2022), zingerone (Zhang et al., 2022)], aloin A (Jiang et al., 2022), alanyl-glutamine (Hou et al., 2013), rosmarinic acid (Formiga et al., 2020), saikosaponin D (Li et al., 2020) and vitamin A (Pang et al., 2021) can effectively enhance the colon tissue expression of mucus-linked mucins and Tff3 in the UC mice to improve the function of colonic barrier.

3.2.4 Oxidative stress reduction

Increased oxidative stress causes colonic mucosal barrier activity loss and a marked reduction in TJ proteins, thus enhancing the risk for the development of UC. In the intestine, inflammation and oxidative stress together disrupt the mucosal redox balance and promotes apoptosis of intestinal epithelial cell (IEC) (Seo et al., 2014). It has been indicated that bruceine D (Dou et al., 2018) and casticin (Ma et al., 2018) reduce malondialdehyde (MDA) and reactive oxygen species (ROS) and enhances glutathione (GSH) and superoxide dismutase to alleviate the damage caused by oxidative stress damage in colon tissues and UC symptoms in animals. Stevioside can alleviate colonic epithelium oxidative damage by UC, including ROS reduction and intestinal mucosal GSH consumption and elevating the enzyme activity of catalase (CAT), GSH (Mostafa et al., 2020), and heme oxygenase-1 (HO-1). Furthermore, there are many compounds, including atractylenolide III (Han et al., 2022), astragaloside IV (Zhong et al., 2022), asperuloside (Chen et al., 2021), alpinetin (Tan and Zheng, 2018), acteoside (Guo et al., 2022), brusatol (Zhou et al., 2018), bruceine D (Dou et al., 2018), betaine (Chen et al., 2022), berberine (Zhang et al., 2017), baicalin (Yao et al., 2016), carnosic acid (Yang et al., 2017), D-pinitol (Lin et al., 2021), epoxy-micheliolide (He et al., 2022), geniposide (Yang et al., 2020), galangin (Sangaraju et al., 2019), 6-gingerol (Ajayi et al., 2018), hydroxytyrosol (Elmaksoud et al., 2021), isatin (Socca et al., 2014), imperatorin (Luo and Luo, 2021), lycopene (Tekeli et al., 2018; Li et al., 2021; Yin et al., 2023), naringenin (Al-Rejaie et al., 2013), 8-oxypalmatine (Cheng et al., 2022), and 3,4-Oxo-isopropylidene-shikimic acid (Xing et al., 2012), puerarin (Jeon et al., 2020), sesamin (Bai et al., 2019), syringic acid (Fang et al., 2019), stevioside (Alavala et al., 2019; Mostafa et al., 2020), sinigrin (Kotipalli et al., 2023), sinapic acid (Qian et al., 2020),

tyrosol (Guvenc et al., 2019), vitamin C (Yan et al., 2015), wogonin (Zhou et al., 2022), et al. that decrease colonic epithelium oxidative damage by UC.

3.2.5 Protecting the intestinal epithelial cells

The IECs have rapid renewal capability (Krndija et al., 2019), ensuring normal digestion and barrier activity, and are based on non-inflammatory apoptosis. At UC onset, IEC travels to the damaged area to maintain the intestinal barrier's integrity (Maria-Ferreira et al., 2018). However, the excessive apoptosis and uncontrolled IEC inflammation are primarily responsible for impaired intestinal mucosal barrier activity in UC. Caspase is the most critical protease associated with apoptosis; Bax and Bcl-2 are essential apoptosis modulatory genes. Some natural compounds, such as 5 alkaloids [berberine (Jia et al., 2020), coptisine (Wang et al., 2021a), indirubin (Gao et al., 2018), isatin (Gao et al., 2018), and palmatine (Zhang et al., 2018)], 3 flavonoids [baicalin (Shen et al., 2019b), hesperidin (Shafik et al., 2019), and wogonin (Zhou et al., 2022)], 2 phenols [hydroxytyrosol (Elmaksoud et al., 2021), polydatin (Lv et al., 2018)], 4 phenylpropanoids [acteoside (Guo et al., 2022), chlorogenic acid (Gao et al., 2019), deoxyschizandrin (Zhang et al., 2016; Yu and Qian, 2021), and sinapic acid (Shahid et al., 2022)], 3 steroids [taraxasterol (Che et al., 2019), taurine (Giris et al., 2008), and taurohyodeoxycholic acid (Laukens et al., 2014)], 4 terpenoids [cycloastragenol (Bagalagel et al., 2022), plumericin (Rapa et al., 2021), paeoniflorin (Gu et al., 2017), and patchouli alcohol (Qu et al., 2017)] and arbutin (Zhang et al., 2021), diacetylrhein (Zohny et al., 2022), glutamate (Li et al., 2014), and nervonic acid (Yuan et al., 2023), have indicated UC improvement, decreasing the expression of Bax, caspase-3, and caspase-9, whereas increasing Bcl-2 in epithelial cells. Meanwhile, anemoside B4 (Zhang et al., 2021), bryodulcosigenin (Li et al., 2022), and berberrubine (Yu et al., 2018) could decrease the ratio of Bax/Bcl-2 and caspase-3, while artesunate (Yin et al., 2020) increasing the ratio of Bcl-2/Bax and decreasing caspase-3. Diosgenin (Tang et al., 2020) can protect against colonic apoptosis by downregulating the Bax/Caspase-1 pathway. In addition, cyclosporine protects against epithelial apoptosis linked with increased tumor growth factor- β -related signaling (Satoh et al., 2009).

3.3 Regulating intestinal mucosal immune response

The intestinal mucosal immunological disorder is the essential factor for UC pathogenesis, characterized by innate immune system alterations, adaptive immune system activation, increased pro-inflammatory mediators, and anti-inflammatory signals inhibition, causing chronic intestinal inflammation. Currently, 283 natural compounds have been indicated to improve mucosal immune response in UC, primarily by regulating cytokine, inflammatory signaling pathways, and immune cells as shown in Figure 5.

3.3.1 Regulating intestinal mucosal immune cells

Various cell types, including antigen-presenting (dendritic cells and macrophages) and effector and regulatory T cells, are critically linked with UC pathogenesis, as they promote or inhibit

inflammation. Macrophages are essential for intestinal homeostasis and the pathology of IBD. Generally, persistent M1 macrophage activation causes excessive stimulation of pro-inflammatory cytokines to release, causing an imbalance of colonic homeostasis and barrier disruption (Bain et al., 2013). Whereas M2 macrophages stimulate anti-inflammatory cytokines to alleviate UC progression (Formentini et al., 2017). It has been reported that didymin (Lv et al., 2021), 1,25-dihydroxyvitamin D3 (Cao et al., 2020), ginsenoside Rg1 (Long et al., 2022), loganin (Liu et al., 2020), methyl gallate (Zhou et al., 2022), platycodin D (Guo et al., 2021), triptolide (Tang et al., 2020), and tiliroside (Zhuang et al., 2021) suppresses M1 macrophages activation and promote M2 macrophages, thereby alleviating UC. In addition, baicalin (Zou et al., 2015), cinnamtannin D1 (Yang et al., 2022), curcumin (Wang et al., 2022), cinnamaldehyde (Qu et al., 2021), epigallocatechin gallate (Xu et al., 2015), ginsenoside compound K (Wang et al., 2022), hyperoside (Cheng et al., 2021), paeoniflorin (Zheng et al., 2020), and salidroside (Liu et al., 2023) can ameliorate chronic intestinal inflammation in UC, and its mechanism that promote intestinal mucosal immune imbalance, thereby regulating Th17/Treg balance.

Interestingly, these compounds are mainly phenols. The endoscopic findings consistently indicate that supplementation with phenols has demonstrated benefits in individuals with IBD. However, to acquire a more comprehensive understanding of the influence of phenols, it is necessary to conduct long-term trials that incorporate both clinical and mechanistic investigations (Hagan et al., 2021).

3.3.2 Regulating the secretion of immune cytokines in the intestinal mucosa

In UC patients' intestines, increased pro-inflammatory cytokines cause persistent mucosal inflammation and are directly linked with UC pathogenesis. The mucosal immune system is the primary factor affecting intestinal injury and inflammation, and with cytokines, it modulates inflammation (Ardizzone and Bianchi Porro, 2005). Therefore, cytokines are a logical UC target that can be modulated by specific inhibitors. The literature has indicated that chemokine ligand 5 (CCL5), cyclooxygenase (COX-2), IL-1 β , IL-6, IL-8, IL-17, interferon- γ (IFN- γ), inducible nitric oxide synthase (iNOS), myeloperoxidase (MPO), nitric oxide (NO), and tumor necrosis factor alpha (TNF- α) expression enhanced in UC animal models, while anti-inflammatory cytokines secretion such as Arg-1, IL-10, and IL-4 decreased. Most natural compounds compiled here can regulate these cytokines to treat UC. Taurohyodeoxycholic acid (TA), a natural 6 α -hydroxylated bile acid with hydrophilic properties, is the main component of traditional Chinese medicine (TCM) *Pulvis Fellis Suis*. TA can modulate multiple cytokines in UC, including CXC motif chemokine ligand 2 (Ccl2), IL-1 β , IL-4, IL-6, IL-10, IL-17A, IL-21, IL-22, IFN- γ , MPO, and TNF- α for maintaining the immune balance of the body (He et al., 2011; Laukens et al., 2014; Lv et al., 2023).

3.3.3 Regulating intestinal mucosal signaling pathways

The pathogenesis of UC is associated with multiple complex inflammatory signaling pathways. Natural compounds directly or indirectly interact with the immune system, stimulating different

① Intestinal mucosal immune cells regulation					
Alkaloids ■ Berberine★ ■ Indirubin ■ Norisoboldine Phenylpropanoids ■ Cinnamaldehyde ■ Rosmarinic acid	Flavonoids ■ Alpinetin ■ Baicalin★ ■ Curcumin★ ■ Didymin ■ Hyperoside ■ Myricetin ■ Tiliroside	Phenols ■ Cinnamtannin D1 ■ Curcumin★ ■ Epigallocatechin gallate ■ Methyl gallate ■ Salidroside★	Terpenoids ■ Astragaloside IV ■ Carnosol ■ Celastrol★ ■ Loganin ■ Paconiflorin★ ■ Triptolide★	Steroids ■ Ginsenoside Rg1 ■ Taurohydroxycholeic acid Other ■ 1,25-Dihydroxyvitamin D3	Saponins ■ Ginsenoside compound K ■ Platycodin D
② Regulation of the secretion of immune cytokines in the intestinal mucosa					
Alkaloids ■ 8-Oxypalmitate ■ Alliin ■ Aspartate★ ■ Berberine★ ■ Berberine★ hydrochloride ■ Citrulline ■ Ergothioneine	■ Berberrubine ■ Betaine ■ Boldine ■ Camptothecin ■ Caulerpin ■ Citrulline ■ Coptisine ■ Corynoline ■ Cyclosporine ■ Demethyleneberberine ■ Dihydroberberine ■ Ergothioneine	■ Evodiamine★ ■ Fumigaclavine C ■ Glutamate ■ ...	Flavonoids ■ Acacetin ■ Alpinetin ■ Amentoflavone ■ Apigenin★ ■ Astragaloside IV ■ Baicalin★ ■ Casticin	■ Catechin-7-O-β-D-glucopyranoside ■ Daidzein ■ Didymin ■ Eriocitrin ■ Eriodictyol ■ Eupatilin ■ Galangin★	■ Genistein ■ Hesperidin ■ Hesperidin methyl chalcone ■ Hydroxyafflor yellow A ■ Hyperoside ■ Isobavachalcone ■ ...
Other ■ 1,25-Dihydroxyvitamin D3 ■ 2,3,5,4'-Tetrahydroxystilbene-2-O-β-D-glucoside★ ■ 3,3'-Diselenodipropionic acid ■ 3,4-Oxo-isopropylidene-shikimic acid ■ 4-Geranyloxy-2,6-Dihydroxybenzophenone ■ 6,7-Dihydroxy-2,4-Dimethoxyphenanthrene ■ Alanyl-glutamine	■ Albiiflorin ■ Anemonin ■ Apocynin ■ Arbutin ■ Butyrate★ ■ Docosapentaenoic acid ■ D-Pinitol	■ Erianin ■ Liriodendrin ■ Nerolidol ■ Nervonic acid ■ Procyanidins★ ■ Sodium houttuynifonate ■ ...	Phenols ■ 6-Gingerol ■ 6-Shogaol ■ Cannabidiol★ ■ Cinnamtannin D1 ■ Curcumin★ ■ Epigallocatechin gallate ■ Ethyl rosmarinic acid ■ Gallic acid ■ Gallotannin corilagin ■ Hydroxytyrosol★	■ Methyl gallate ■ Paconol★ ■ Polydatin ■ Prim-O-Glucosyleimifugin★ ■ Procyanidin A1 ■ Resveratrol ■ Salidroside★ ■ Syringic acid ■ Thymol ■ Zingerone★	Phenylpropanoids ■ Aescoside ■ Aesculin ■ Bergenin ■ Caffeic acid ■ Caffeic acid phenethyl ester ■ Chlorogenic Acid★ ■ Cinnamaldehyde ■ Deoxyschizandrin ■ Ferulic Acid ■ Honokiol ■ Phillygenin ■ Rosmarinic acid ■ Schisandrin B ■ Sesamin ■ Sinapic acid ■ Wedelolactone
Steroids ■ Convallatoxin ■ Diosgenin ■ Ginsenoside Rg1 ■ Ginsenoside Rg3 ■ Physalin B	Vitamin ■ Vitamin A★ ■ Vitamin D3 ■ Vitamin E ■ β-Carotene★	Saponins ■ Anemoside B4★ ■ Astragaloside II ■ Dioscin ■ Ginsenoside compound K★	Quinonoids ■ Barbaloin ■ Diacetylrhein ■ Emodin ■ Juglone ■ Shikonin ■ Rhein		
③ Intestinal mucosal signaling pathways regulation					
(1) Inhibition of the TLR4/NF-κB signaling					
Alkaloids ■ Coptisine ■ Demethyleneberberine ■ Eodiamine ■ Homoharringtonine ■ Indirubin	Flavonoids ■ Amentoflavone ■ Astragaloside IV ■ Daidzein ■ Fisetin ■ Galangin★ ■ Icarin ■ Isovitexin ■ Luteolin★ ■ Wogonoside	■ Naringin★ ■ Phloretin★ ■ Puerarin★ ■ Taxifolin ■ Wogonoside	Terpenoids ■ Epoxymicheliolide ■ Andrographolide★ ■ Artesunate ■ Bilobalide★ ■ Carnosic acid ■ Dihydroartemisinin	■ Geniposide ■ Ginsenoside rd ■ Loganin ■ Lycopene ■ Paconiflorin★ ■ Perillyl alcohol ■ Ursolic acid ■ Sauchinone★ ■ Sericic acid ■ Squalene ■ Stevioside ■ Terpinen-4-ol★ ■ Ursolic acid ■ B-patchoulene	
(2) Inhibition of the MAPK signaling pathway					
Alkaloids ■ Berberine★ ■ Camptothecin ■ Indirubin ■ Isatin	Flavonoids ■ Baicalin★ ■ Eupatilin ■ Fisetin ■ Isovitexin ■ Wogonoside	Phenylpropanoids ■ Aretigenin ■ Chlorogenic acid ■ Phillygenin ■ Rosmarinic acid ■ Wedelolactone	Saponins ■ Anemoside B4★ ■ Dioscin	Terpenoids ■ Bilobalide★ ■ Geniposide ■ Ginkgolide C★ ■ Squalene ■ Stevioside	
(3) Inhibition of the JAK-STAT signaling pathway		(4) Inhibition of the PI3K/Akt/mTOR signaling pathway			
Alkaloids ■ Berberine★	Terpenoids ■ Dihydroartemisinin	Alkaloids ■ Phellodendrine	Flavonoids ■ Quercetin★	Phenols ■ Resveratrol	Quinonoids ■ Emodin ■ Rhein
(5) Inhibition of the ROCK-MLC signaling pathway		(6) Inhibition of the NLRP3 inflammasome			
Alkaloids ■ Berberine★	Flavonoids ■ Wogonoside	Quinonoids ■ Barbaloin	Terpenoids ■ Geniposide ■ β-Patchoulene★	Vitamins ■ Vitamin D3	Other ■ 1,25-Dihydroxyvitamin D3 ■ 3,3'-Diselenodipropionic acid
(7) Activation of the PPARγ signaling pathway		(8) Regulation of the Wnt/β-catenin signaling pathway			
Alkaloids ■ Demethyleneberberine ■ Tetramethylpyrazine	Flavonoids ■ Baicalin★ ■ Hyperoside	Quinonoids ■ Emodin	Steroids ■ Convallatoxin	Alkaloids ■ Berberine★ ■ Betaine ■ Evodiamine★ ■ Heterophyllin B★ ■ Nigakinone★	Phenylpropanoids ■ Bergenin ■ Chlorogenic Acid★ ■ Cinnamaldehyde ■ Ferulic Acid ■ Schisandrin B ■ Sinapic acid ■ Wedelolactone
Terpenoids ■ Artemisinin ■ Geniposide ■ Glycyrrhizin ■ Stevioside	Other ■ D-Pinitol ■ Dioscin	Phenols ■ Salidroside★ ■ Zingerone★	Phenols ■ Gallic acid ■ Hydroxytyrosol★ ■ Salidroside★ ■ Genistein ■ Naringin★ ■ Phloretin★	Terpenoids ■ Bryodulcosigenin ■ Geniposide	■ 1,25-Dihydroxyvitamin D3 ■ Ginsenoside Rg1 ■ Ginsenoside Rk3★ ■ Mogrol ■ Terpinen-4-ol★

FIGURE 5 Natural compounds against ulcerative colitis via regulation of intestinal mucosal immune response. Red pentagrams indicate compounds involved in Mechanism I, II, and III. The natural compounds involved in this paper are shown in Supplementary Table S1.

molecular and cellular pathways and producing anti-inflammatory effects. Therefore, UC prevention and therapy by natural molecules regulate one or more complicated signaling pathways.

3.3.3.1 Inhibition of the TLR4/NF- κ B signaling pathway

NF- κ B is a transcription factor that stimulates inflammatory cytokines's genetic transcription and is linked with multiple inflammatory diseases. Physiologically, inactive NF- κ B interacts with cytoplasmic inhibitor protein I κ B (I κ B). During inflammation, I κ B undergoes phosphorylation and degradation, dissociating NF- κ B and translocating it from the cytoplasm to the nucleus, thereby activating downstream gene transcription, such as pro-inflammatory cytokines and iNOS. Therefore, NF- κ B inhibition is an efficient strategy to prevent UC and inflammatory cytokines release in UC patients. It is reported that 14 alkaloids [corynoline (Zhang et al., 2023), cavidine (Niu et al., 2015), caulerpin (Lucena et al., 2018), coptisine (Wang et al., 2021a), demethyleneberberine (Zhao et al., 2022b), evodiamine (Shen et al., 2019), homoharringtonine (Liu et al., 2022), isatin (Gao et al., 2018), indirubin (Gao et al., 2018), leonurine (Zheng et al., 2021), melittin (Ahmedy et al., 2020), nigeplanine (Gao et al., 2019), platycodin D (Guo et al., 2021), and sinomenine (Zhou et al., 2023)], 16 flavonoids [astragaloside (Peng et al., 2020), and amentoflavone (Sakthivel and Guruvayoorappan, 2013), baicalin (Feng et al., 2014), daidzein (Shen et al., 2019c), eupatilin (Zhou et al., 2018), euptailin (Zhou et al., 2018), fisetin (Sahu et al., 2016), galangin (Sangaraju et al., 2019), icariin (Zhang et al., 2021a), licochalcone A (Liu et al., 2018), luteolin (Li et al., 2021), α -mangostin (You et al., 2017), naringin (Cao et al., 2018), puerarin (Jeon et al., 2020), phloretin (Zhang et al., 2019), and taxifolin (Li W. et al., 2022)], 6 phenols [epicatechin (Zhang et al., 2016), gallic acid (Zhu et al., 2019b), polydatin (Yao et al., 2011), prim-*o*-glucosylcimifugin (Yin et al., 2022), thymol (Chamanara et al., 2019), and zingerone (Zhang et al., 2022)], 7 phenylpropanoids [arctigenin (Wu et al., 2014), aesculin (Tiana et al., 2019), bergenin (Wang et al., 2018), caffeic acid (Zhang et al., 2016), chlorogenic acid (Zeng et al., 2020), phillygenin (Xue et al., 2023), and wedelolactone (Wei et al., 2017)], 25 terpenoids [astragaloside IV (Wu and Chen, 2019), astragaloside (Peng et al., 2020), asperuloside (Chen et al., 2021), artesunate (Chen et al., 2019), bilobalide (Zhang et al., 2021c), brusatol (Zhou et al., 2018), carnosic acid (Yang et al., 2017), β -carotene (Zhu et al., 2021), diosgenin (Tang et al., 2020), dihydroartemisinin (Li N. et al., 2019), epoxymicheliolide (He et al., 2022), geniposide (Yang et al., 2020), ginkgolide C (Xu D. et al., 2022), ginsenoside Rd (Qu et al., 2022), patchouli alcohol (Wu et al., 2020), parthenolide (Zhao et al., 2012), picroside II (Yao et al., 2022), paeoniflorin (Gu et al., 2017), rographolide (Zhang et al., 2020), stevioside (Alavala et al., 2019), squalene (Sanchez-Fidalgo et al., 2015), sericic acid (Lifei et al., 2023), sauchinone (Wu et al., 2023), terpinen-4-ol (Zhang et al., 2017a), and ursolic acid (Liu et al., 2016)], and 2 steroids [convallatoxin (Li et al., 2019), physalin B (Zhang et al., 2020)], 2 aponins [dioscin (Cai et al., 2021), saikosaponin A (Zhou et al., 2019)] and albiflorin (Wang et al., 2023), erianin (Dou et al., 2020), 4-geranyloxy-2,6-dihydroxybenzophenone (Wang et al., 2023), liriiodendrin (Zhang et al., 2017c), nervonic acid (Yuan et al., 2023), and vitamin C (Kondo et al., 2019) have been indicated to improve UC related systemic symptoms by suppressing NF- κ B inflammatory signaling pathway.

Furthermore, the Toll-like receptor 4 (TLR4) is an essential signaling pathway associated with colon inflammation (Rashidian et al., 2020). As an innate immune receptor, TLR4 is activated during inflammation after gut pathogen-associated molecular patterns (PAMPs) recognition, conformation alterations, and dimerization. Activated TLR4 is then recruited at aptamer, activating NF- κ B. Much research suggests that colitis is linked with excessive activation of the TLR4/NF- κ B signaling pathway (Rashidian et al., 2016; Liu et al., 2017). It is reported that the baicalin (Cui et al., 2014; Feng et al., 2014), berberine (Zhang et al., 2011), cinnamaldehyde (Tan et al., 2023), deoxyschizandrin (Zhang et al., 2016), eriodictyol (Hu et al., 2021), emodin (Xu et al., 2021), honokiol (Wang et al., 2022), hyaconitin (Zhang et al., 2011), hydroxysafflor yellow A (Feng et al., 2022), honokiol (Wang et al., 2022), methyl gallate (Zhou et al., 2022), naringenin (Dou et al., 2013), perillyl alcohol (Puppala et al., 2022), vitexin (Duan et al., 2020), and vitexin (Duan et al., 2020) have been linked with the inhibition of the TLR4/NF- κ B signaling pathway and inflammatory cytokines, thereby exerting anti-UC effect.

3.3.3.2 Inhibition of the MAPK signaling pathway

MAPKs family comprises evolutionarily conserved serine/threonine protein kinases, which regulate cellular pathways, such as inflammation-related genes. Among these, stimulation of extracellular-signal-regulated kinases 1/2 (ERK-1/2), c-Jun N-terminal kinase (JNK), and p38 kinase (p38) promotes cell apoptosis and aggravates intestinal inflammation. Much research has been published indicating that natural compounds, 7 alkaloids [camptothecin (Wang et al., 2021b), isatin (Gao et al., 2018), indirubin (Gao et al., 2018), melittin (Ahmedy et al., 2020), nigeplanine (Gao et al., 2019), sinigrin (Kotipalli et al., 2023), and tetramethylpyrazine (He et al., 2012)], 5 flavonoids [baicalin (Liang et al., 2019), eupatilin (Zhou et al., 2018), licochalcone B (Zhang et al., 2022b), licochalcone A (Zhang et al., 2021), and naringin (Cao et al., 2018)], 8 terpenoids [bilobalide (Zhang et al., 2021c), ginkgolide C (Xu et al., 2022), geniposide (Lu et al., 2021), pedunculolide (Liu et al., 2020), squalene (Sanchez-Fidalgo et al., 2015), stevioside (Alavala et al., 2019), squalene (Sanchez-Fidalgo et al., 2015), and ursolic acid (Sheng et al., 2021)], 4 phenylpropanoids [arctigenin (Wu et al., 2014), chlorogenic acid (Gao et al., 2019), phillygenin (Xue et al., 2023), and wedelolactone (Wei et al., 2017)], albiflorin (Wang et al., 2023), atractylodin (Qu et al., 2021), β -carotene (Zhu et al., 2021), dioscin (Cai et al., 2021), α -mangostin (You et al., 2017), nervonic acid (Yuan et al., 2023), and prim-*O*-Glucosylcimifugin (Yin et al., 2022) have suppressive effect on MAPK pathway, reducing the expression and inflammatory mediators release.

3.3.3.3 Inhibition of the JAK-STAT signaling pathway

The janus kinase/signal transducer and activator of transcription (JAK/STAT) is a common signaling pathway for transducing signals from various cytokines, which widely regulate cell growth, differentiation, inflammation, apoptosis, and other mechanisms. berberine (Zhang et al., 2017), dihydroartemisinin (Jiang et al., 2021), and erianin (Dou et al., 2020) can ameliorate UC's intestinal mucosal inflammation by downregulating phosphorylated Janus kinase 2 (p-JAK2), JAK2, phosphorylated signal transducer and activator of transcription 3 (p-STAT3), and

signal transducer and activator of transcription 3 (STAT3) expression.

3.3.3.4 Inhibition of the PI₃K/Akt/mTOR signaling pathway

Studies have shown that the PI₃K/AKT signaling pathway plays an important role in the occurrence of UC (Dong et al., 2022). The inflammatory response can be alleviated by blocking this signal transduction pathway, thus presenting a promising target for treating UC. Interestingly, dihydroartemisinin (Jiang et al., 2021), glutamine (Yan et al., 2020), ihydroartemisinin (Li et al., 2019), luteolin (Vukelic et al., 2020), platycodin D (Guo et al., 2021), quercetin (Zhang et al., 2023), and rhein (Dong et al., 2022) attenuate DSS-induced colitis *via* PI₃K/AKT signaling pathway inhibition. Additionally, mTOR, a downstream target of PI₃K/AKT, primarily modulates cell growth and metabolism, promoting anabolism, including ribosome biogenesis and synthesizing nucleotides, proteins, fatty acids, and lipids, and inhibiting catabolism. P-mTOR upregulation in the colon tissues of DSS-induced UC rats causes autophagy dysfunction. However, alpinetin (Miao et al., 2019), dioscin (Li et al., 2022), friedelin (Shi et al., 2021), phellodendrine (Su et al., 2021), and rhein (Dong et al., 2022) reverse this effect, return mTOR to normal levels, and inhibit inflammatory cascade, thereby improving intestinal inflammation.

3.3.3.5 Inhibition of the ROCK-MLC signaling pathway

The Ras homologous protein A-Rho kinase (RhoA-ROCK) signaling pathway modulates TJ synthesis, polymerization, and epithelial cell gap permeability, typically linked with the ROCK-MLC pathway (Liu et al., 2020). ROCKs is a serine-threonine kinase family member, including Rho-associated kinase 1 (ROCK1) and ROCK2. ROCK1 directly modulates myosin light chain 2 (MLC2) activation and myosin contraction for TJ depolymerization, accompanied by increased intercellular permeability (Fu et al., 2019). According to a study, UC animals have increased ROCK1 and MLC2 phosphorylation in the colon; however, barbaloin (Gai et al., 2019), geniposide (Xu et al., 2017), and β -patchoulene (Liu et al., 2020) substantially downregulate them to improve the colonic barrier.

3.3.3.6 Inhibition of the NLRP³ inflammasome

Recently, it was observed that single nucleotide polymorphisms (SNPs) in genes encoding the NOD-like receptor protein 3 (NLRP3) are associated with IBD susceptibility. NLRP3 belongs to the NOD-like receptor (NLR) family (Kanneganti et al., 2007; Abraham et al., 2017). It is a cytosolic platform protein that assembles inflammasome, a protein complex involved in proteolytic maturation and release of IL-1 and IL-18 pro-inflammatory cytokines (Martinon et al., 2002; Villani et al., 2009). NLRP3 inflammasome regulates various inflammatory and autoimmune disorders (Cao et al., 2020). There were 11 alkaloids [betaine (Chen et al., 2022), berberine (Li et al., 2020), demethyleneberberine (Zhao et al., 2022b), evodiamine (Shen et al., 2019), heterophyllin B (Chen et al., 2022), nigeplanine (Gao et al., 2019), nigakinone (Liu et al., 2023), 8-Oxypalmatine (Cheng et al., 2022), palmatine (Mai et al., 2019), sinomenine hydrochloride (Zhou et al., 2021), and sanguinarine (Li et al., 2022b)], 7 phenylpropanoids [bergenin (Lopes de Oliveira et al., 2019), cinnamaldehyde (Tan et al., 2023), chlorogenic acid (Zeng

et al., 2020), ferulic acid (Yu S. et al., 2023), sinapic acid (Qian et al., 2020), schisandrin B (Zhang et al., 2021), and wedelolactone (Wei et al., 2017)], 9 terpenoids [brusatol (Zhou et al., 2018), bryodulcosigenin (Li et al., 2022), ginsenoside Rk3 (Tian et al., 2020), ginsenoside Rg3 (Liu et al., 2023), ginsenoside Rd (Liu C. et al., 2018), geniposide (Pu et al., 2020), mogrol (Liang et al., 2021), picroside II (Yao et al., 2022), and terpinen-4-ol (Zhang et al., 2017a)], 2 flavonoids [naringin (Cao et al., 2018), phloretin (Zhang et al., 2019)], 3 phenols [gallic acid (Yu et al., 2023), hydroxytyrosol (Miao, 2022), and salidroside (Liu et al., 2019)], diacetylrhein (Zohny et al., 2022), dioscin (Cai et al., 2021), 1,25-dihydroxyvitamin D3 (Cao et al., 2020), 3,3'-diselenodipropionic acid (Zheng et al., 2023), physalin B (Zhang et al., 2020), and vitamin D3 (Gao et al., 2023) have been linked with the alleviation of UC *via* NLRP3 inhibition.

3.3.3.7 Activation of the PPAR γ signaling pathway

PPAR γ is a transcriptional factor expressed mainly in colonic epithelial cells, and UC, its expressions are reduced (Su et al., 1999; Aoyagi et al., 2010). PPAR γ activation decreases UC-mediated NF- κ B pathway stimulation and inflammatory cytokines (IL-6, IL-1 β , and TNF- α) expression (Aoyagi et al., 2010). Furthermore, targeted PPAR γ expression alteration enhances mice's susceptibility towards DSS-induced colitis (Shah et al., 2007; Aoyagi et al., 2010). However, natural compounds, artemisinin (Jia et al., 2022), baicalin (Xu et al., 2021), bergenin (Wang et al., 2018), aesculin (Tiana et al., 2019), convallatoxin (Li et al., 2019), dioscin (Wu et al., 2021), d-pinitol (Lin et al., 2021), demethyleneberberine (Zhao et al., 2022b), emodin (Luo et al., 2022), glycyrrhizin (Sethuraman et al., 2015), geniposide (Zhang et al., 2017d), honokiol (Wang et al., 2022), hyperoside (Cheng et al., 2021), luteolin (Li et al., 2021), naringin (Cao et al., 2018), phloretin (Zhang et al., 2019), salidroside (Liu et al., 2019), stevioside (Mostafa et al., 2020), tetramethylpyrazine (He et al., 2012), and zingerone (Zhang et al., 2022), can upregulate PPAR γ expression to alleviate UC.

3.3.3.8 Regulation of the Wnt/ β -catenin signaling pathway

It has been observed that the Wnt signaling pathway substantially affects epithelial cell proliferation to repair mechanical barriers (Kuhnert et al., 2004). Wnt modulates β -catenin expression and is involved in the pathological and physiological mechanisms of injury (Whyte et al., 2012). Multiple research indicates that berberine (Dong et al., 2022c) and 6-gingerol (Ajayi et al., 2018) alleviate UC by maintaining intestinal mucosal barrier function and structure and function, regulating the homeostasis of intestinal mucosal immunity *via* the Wnt/ β -catenin pathway.

3.4 Regulating other mechanisms

3.4.1 Regulating cellular autophagy

One of the cellular self-protection mechanisms is autophagy, which is a self-protective mechanism that maintains homeostasis. It is an evolutionarily conserved mechanism that starts with the generation of a double-membrane autophagosome with cytoplasmic contents (Xie et al., 2020). It is essential for maintaining intestinal homeostasis, modulation of gut ecology,

appropriate intestinal immune responses, and microbial protection. It has been suggested that autophagy can substantially suppress cells' inflammatory reactions (Lin et al., 2019; Larabi et al., 2020). Notably, natural compounds [i.e., alpinetin (Miao et al., 2019), berberine (Xu et al., 2022c), curcumin (Zhang et al., 2019), dioscin (Li et al., 2022), friedelin (Shi et al., 2021), galangin (Xuan et al., 2020), luteolin (Vukelic et al., 2020), palmatine (Mai et al., 2019), procyanidin A1 (Zhang et al., 2022), resveratrol (Pan et al., 2020)], and salidroside (Liu et al., 2023) can improve autophagy and reduce inflammation in the intestinal disorders. Berberine alleviates DSS-induced UC and suppresses the expression and release of lysozyme by stimulating autophagy via adenosine 5'-monophosphate (AMP)-triggered protein kinase (AMP-activated protein kinase) (AMPK)/mammalian target of rapamycin (mechanistic target of rapamycin kinase) (MTOR)/unlike autophagy activating kinase 1 ULK1 (unc-51 like autophagy activating kinase 1) pathway (Foerster et al., 2022).

3.4.2 Inhibiting ferroptosis

In 2012, ferroptosis was formally stated as an iron-dependent, non-apoptotic cell death manifested by the accumulation of lipid peroxidation products and the depletion of membrane polyunsaturated fatty acid (Dixon et al., 2012). It is characterized by lipid peroxidation, iron accumulation, and increased ROS generation. Iron sagging includes iron deposition, increased lipid peroxidation, reduced GSH, inactivation of glutathione peroxidase 4 (GPX4), and enhanced lipoxygenase (LOX), all of which are linked with UC pathogenesis (Huang et al., 2022). These findings validate that ferroptosis inhibition might be a novel target for treating UC (Wang et al., 2020; Chen et al., 2021; Dong et al., 2021; Tang et al., 2021). β -Caryophyllene is widely found in various plant essential oils, and its flavor and fragrance resembles bicyclic sesquiterpene (Jha et al., 2021). A study revealed that β -caryophyllene acts as an inhibitor of ferroptosis that represses lipid peroxidation and inflammation, thereby alleviating UC (Wu et al., 2022).

3.4.3 Regulating metabolism pathway

The literature suggests that metabolic reprogramming can regulate the activation of macrophages. The metabolic signals furnish energy and polarize macrophages. M1 macrophages substantially depend on glycolytic metabolism, whereas M2 primarily depends on oxidative phosphorylation (Saha et al., 2017). Glucose is converted to pyruvate and lactic acid glycolysis via a series of cytoplasmic enzymes. Pyruvate dehydrogenase kinase 1 (PDK1) knockdown is a key modulator enzyme of glucose metabolism, reducing M1 but enhancing M2 macrophage activation (Tan et al., 2015). Glycolysis inhibitor 2-deoxy-D-glucose (2-DG) reduces M1 macrophage activation and pro-inflammatory cytokines secretion (Wang et al., 2018). It has been revealed that tiliroside alleviates UC by restoring the M1/M2 macrophage balance via the HIF-1 α /glycolysis pathway (Zhuang et al., 2021).

3.4.4 Inhibiting endoplasmic reticulum stress responses

The endoplasmic reticulum (ER) is an essential cellular organelle with multiple functions to store free calcium and

synthesize, mature, and transport various lipids, proteins, sterols, etc. Because of multiple cellular factors, proteins are unable to fold correctly, resulting in the accumulation of newly synthesized unfolded proteins in cells, thereby promoting ER stress (Song et al., 2021). Much research indicates that ER stress is associated with UC progression. Highly secretory cells, such as intestinal paneth and goblet cells, are specifically impressionable to ER stress (Kaser et al., 2010). Inhibition of ER stress responses is thus an important therapeutic rationale for UC. Limonin might be utilized for this purpose as it blocks the PERK-ATF4-CHOP pathway of ER stress (Song et al., 2021). Furthermore, berberine (Shen et al., 2020) and artesunate (Yin et al., 2021) reduce ER stress-related marker proteins (glucose-regulated protein, GRP78, C/EBP-homologous protein, CHOP) to treat UC.

4 Analysis of important natural compounds

Since pharmacotherapy based on a single target has been insufficient for drug development in complex diseases, the emerging multi-target approach is a promising strategy for the search of new drug candidates. Therefore, we analyzed the relationship between the 279 natural compounds and mechanisms covered in this review. The 279 natural compounds, including 62 terpenoids, 57 alkaloids, 52 flavonoids, 26 phenols, 19 phenylpropanoids, 9 steroids, 9 saponins, 8 quinonoids, 6 vitamins, and 31 others, can effectively ameliorate UC. Of these, terpenoids, alkaloids, and flavonoids have the greatest potential for treating UC. It is noteworthy to highlight that a total of 54 compounds are linked to Mechanism I, II, and III; 151 compounds are associated with Mechanism I and II; 18 compounds are associated with Mechanism II and III; 4 compounds are associated with Mechanism I; 50 compounds are associated with Mechanism II; 2 compounds are related to Mechanism III (Figure 6).

Furthermore, we conducted a comprehensive search of the Pubchem and Drugbank databases to obtain pertinent data regarding the clinical studies associated with the aforementioned natural compounds. Consequently, a total of 6 compounds (andrographolide, berberine, berberine hydrochloride, butyrate, curcumin, and diosmin) for the therapeutic management of UC were identified to be either in the clinical stage of development or already available on the market (Table 1). Interestingly, the vast majority of these compounds can alleviate UC by Mechanism I, II, and III. This indicates that we should pay more attention to the compounds with multiple mechanisms in the follow-up UC drug research (Figure 7).

Many synthetic drugs are currently in use to treat UC such as 5-aminosalicylic acid (5-ASA) (Hossen et al., 2020). Therapeutic mechanisms of 5-ASA for UC include inhibition of cyclooxygenases and lipoxygenase, activation of peroxisome proliferator activated receptor γ , inhibition of T-cell proliferation and activation, reduction of chemotaxis, adhesion and phagocytosis, inhibition of nuclear factor- $\kappa\beta$ (Hauso et al., 2015). Overall, 5-ASA appears to exert its therapeutic effect by topical action on the affected areas of inflammation. This is the

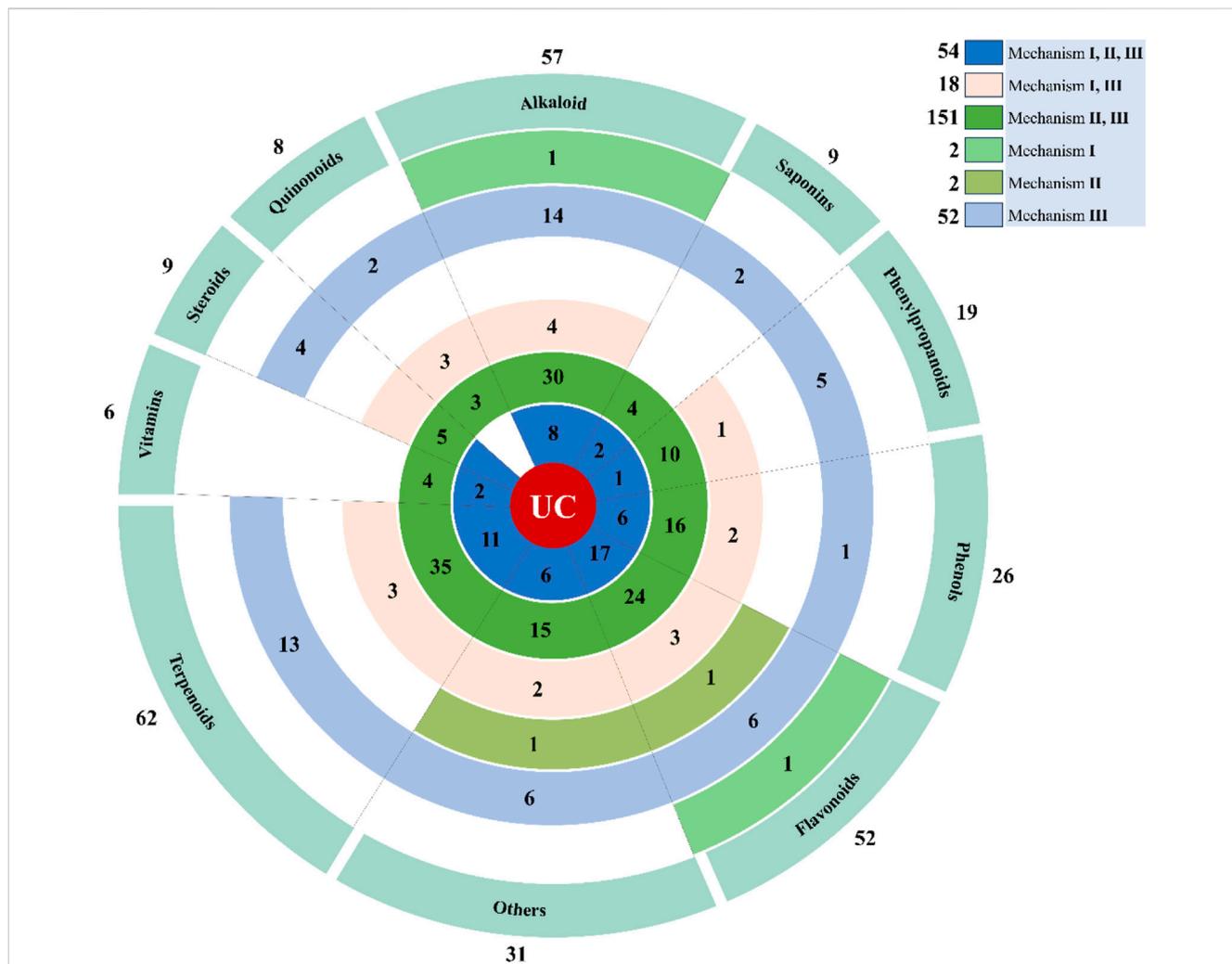


FIGURE 6 Displays the total amount of natural compounds associated with various mechanisms. The natural compounds involved in this paper are shown in Supplementary Table S1.

TABLE 1 Natural compounds in clinical trials for the treatment of ulcerative colitis.

Natural compounds	Clinical trial ID/ Approval number	Study title	Phase	Drug name
Diosmin	NCT05626166	The efficacy and safety of diosmin in patients with ulcerative colitis	Phase III	-
Curcumin	NCT01320436	Curcumin + aminosalicic acid (5ASA) versus 5ASA alone in the treatment of mild to moderate ulcerative colitis	Phase III	-
Butyrate	NCT05218850	The use of butyrate therapy in pediatric ulcerative colitis	Phase I	-
Berberine	NCT02962245	Efficacy of treatment with berberine to maintain remission in ulcerative colitis	Phase IV	-
Berberine hydrochloride	H61022181	-	Approved	Berberine Hydrochloride Tablets
Andrographolide	NCT00659802	Phase II study of HMPL-004 in patients with ulcerative colitis	Phase II	-
	NCT01882764	HMPL-004 maintenance treatment in subjects with mild to moderate ulcerative colitis	Phase III	-
	NCT01805791	A phase III trial of HMPL-004 in patients with mild to moderate active ulcerative colitis	Phase III	-

Note: Drugbank and Pubchem databases were used to obtain natural compounds used in clinical studies.

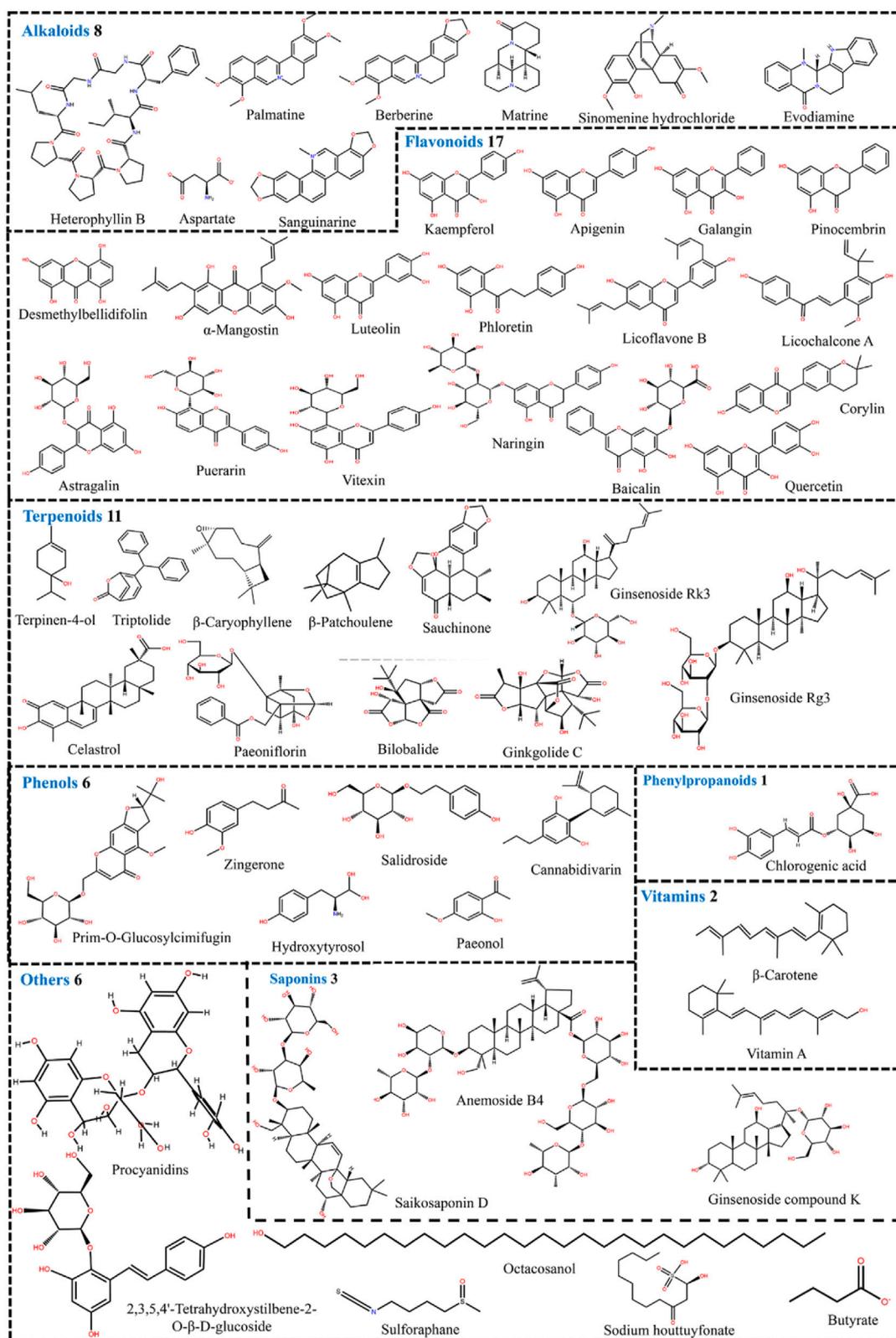


FIGURE 7 54 natural compounds that can treat ulcerative colitis by regulating multiple mechanisms (Mechanism I, II, and III).

same as one of the mechanisms (Mechanism III) by which natural compounds treat UC. However, the mechanism of natural compounds against UC is more complex compared to synthetic drugs. Furthermore, 5-ASA have some drawbacks as long-term use results in side effects including nausea, vomiting, fatigue, diarrhea, abdominal pain, pulmonary fibrosis, etc (Rogler, 2010). For centuries, herbal treatments have shown their potential to ameliorate countless diseases and disorders with no or fewer side effects. In conclusion, natural compounds have a richer mechanism for treating UC than synthetic drugs, and natural compounds are more abundantly available and have fewer side effects.

5 Concluding remarks and future directions

This review provides a comprehensive overview of the protective effects exhibited by natural substances against UC, while also delving into their probable mechanisms of action in mitigating colitis. Results indicated that 279 natural compounds (62 terpenoids, 57 alkaloids, 52 flavonoids, 26 phenols, 19 phenylpropanoids, 9 steroids, 9 saponins, 8 quinonoids, 6 vitamins, and 31 others) can act on various mechanisms to improve UC, such as regulating gut microbiota and metabolites (Mechanisms I), protecting the intestinal mucosal barrier (Mechanisms II), regulating intestinal mucosal immune response (Mechanisms III), as well as the other mechanisms (cellular autophagy modulation and ferroptosis inhibition). More importantly, (1) 54 natural compounds exhibit their therapeutic effects by modulating Mechanisms I, II, and III, which can be used to develop multitargeted drugs for UC; Terpenoids, alkaloids, and flavonoids have the greatest potential for treating UC. (2) Mechanism III is regulated by all natural compounds; Mechanisms II and III can be modulated by at least half of the compounds, which may give information on the etiology of UC. In conclusion, this review serves as a comprehensive resource for the pharmaceutical industry, researchers, and clinicians seeking novel therapeutic approaches to combat UC. Harnessing the therapeutic potential of these natural compounds may significantly contribute to the improvement of the quality of life of patients with UC and promotion of disease-modifying therapies in the future.

This review fails to resolve some issues and requires further research and refined methodology to provide evidence for the natural compound's therapeutic efficacy. The limitations include: (1) Disadvantages including reduced water insolubility, oral bioavailability, rapid metabolism, and increased degradation limit the clinical use of various natural compounds. However, different drug delivery strategies can resolve these issues. (2) Clinical trials are required to assess natural compounds' safety and efficacy profiles, the elucidation criteria of which are not uniform for UC, and the treatment mechanism is not thoroughly studied. Research requires standardization and rationalization to improve UC's therapeutic effect and promote new drug development. (3) Co-treatment of natural compounds and other drugs should be studied for improved treatment. Furthermore, applying targeted preparations would benefit the targeted delivery of natural compounds with an increased curative effect and potential.

Author contributions

YH: Methodology, Software, Writing–original draft, Writing–review and editing. QW: Software, Writing–review and editing. SL: Investigation, Resources, Software, Writing–review and editing. XL: Data curation, Software, Writing–review and editing. SY: Data curation, Software, Writing–review and editing. RZ: Data curation, Software, Writing–review and editing. CF: Conceptualization, Supervision, Writing–original draft, Writing–review and editing. ZZ: Conceptualization, Software, Supervision, Writing–original draft, Writing–review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fphar.2024.1394124/full#supplementary-material>

References

- Abraham, B. P., Ahmed, T., and Ali, T. (2017). Inflammatory bowel disease: pathophysiology and current therapeutic approaches. *Handb. Exp. Pharmacol.* 239, 115–146. doi:10.1007/164_2016_122
- Ahmedy, O. A., Ibrahim, S. M., Salem, H. H., and Kandil, E. A. (2020). Antiulcerogenic effect of melittin via mitigating TLR4/TRAF6 mediated NF- κ B and p38MAPK pathways in acetic acid-induced ulcerative colitis in mice. *Chem. Biol. Interact.* 331, 109276. doi:10.1016/j.cbi.2020.109276
- Ajayi, B. O., Adedara, I. A., and Farombi, E. O. (2018). Protective mechanisms of 6-gingerol in dextran sulfate sodium-induced chronic ulcerative colitis in mice. *Hum. Exp. Toxicol.* 37, 1054–1068. doi:10.1177/0960327117751235
- Alavala, S., Sangaraju, R., Nalban, N., Sahu, B. D., Jerald, M. K., Kilari, E. K., et al. (2019). Stevioside, a diterpenoid glycoside, shows anti-inflammatory property against Dextran Sulphate Sodium-induced ulcerative colitis in mice. *Eur. J. Pharmacol.* 855, 192–201. doi:10.1016/j.ejphar.2019.05.015
- Al-Rejaie, S. S., Abuhashish, H. M., Al-Enazi, M. M., Al-Assaf, A. H., Parmar, M. Y., and Ahmed, M. M. (2013). Protective effect of naringenin on acetic acid-induced ulcerative colitis in rats. *World J. Gastroenterol.* 19, 5633–5644. doi:10.3748/wjg.v19.i34.5633
- Aoyagi, Y., Nagata, S., Kudo, T., Fujii, T., Wada, M., Chiba, Y., et al. (2010). Peroxisome proliferator-activated receptor gamma 2 mutation may cause a subset of ulcerative colitis. *Pediatr. Int.* 52, 729–734. doi:10.1111/j.1442-200X.2010.03195.x
- Arduzzone, S., and Bianchi Porro, G. (2005). Biologic therapy for inflammatory bowel disease. *Drugs* 65, 2253–2286. doi:10.2165/00003495-200565160-00002
- Bagalagel, A., Diri, R., Noor, A., Almasri, D., Bakhs, H. T., Kutbi, H. I., et al. (2022). The therapeutic effects of cycloastragenol in ulcerative colitis by modulating SphK/MIP-1 α /miR-143 signalling. *Basic Clin. Pharmacol. Toxicol.* 131, 406–419. doi:10.1111/bcpt.13788
- Bai, X., Gou, X., Cai, P., Xu, C., Cao, L., Zhao, Z., et al. (2019). Sesamin enhances nrf2-mediated protective defense against oxidative stress and inflammation in colitis via AKT and ERK activation. *Oxid. Med. Cell. Longev.* 2019, 2432416. doi:10.1155/2019/2432416
- Bain, C. C., Scott, C. L., Uronen-Hansson, H., Gudjonsson, S., Jansson, O., Grip, O., et al. (2013). Resident and pro-inflammatory macrophages in the colon represent alternative context-dependent fates of the same Ly6Chi monocyte precursors. *Mucosal Immunol.* 6, 498–510. doi:10.1038/mi.2012.89
- Britton, G. J., Contijoch, E. J., Mogno, I., Vennaro, O. H., Llewellyn, S. R., Ng, R., et al. (2019). Microbiotas from humans with inflammatory bowel disease alter the balance of gut Th17 and ROR γ t+ regulatory T cells and exacerbate colitis in mice. *Immunity* 50, 212–224. doi:10.1016/j.immuni.2018.12.015
- Cai, J., Liu, J., Fan, P., Dong, X., Zhu, K., Liu, X., et al. (2021). Dioscin prevents DSS-induced colitis in mice with enhancing intestinal barrier function and reducing colon inflammation. *Int. Immunopharmacol.* 99, 108015. doi:10.1016/j.intimp.2021.108015
- Cao, H., Liu, J., Shen, P., Cai, J., Han, Y., Zhu, K., et al. (2018). Protective effect of naringin on DSS-induced ulcerative colitis in mice. *J. Agric. Food Chem.* 66, 13133–13140. doi:10.1021/acs.jafc.8b03942
- Cao, R., Ma, Y., Li, S., Shen, D., Yang, S., Wang, X., et al. (2020). 1,25(OH) $_2$ D-3 alleviates DSS-induced ulcerative colitis via inhibiting NLRP3 inflammasome activation. *J. Leukoc. Biol.* 108, 283–295. doi:10.1002/jlb.3ma0320-406rr
- Cao, R., Wu, X., Guo, H., Pan, X., Huang, R., Wang, G., et al. (2021). Naringin exhibited therapeutic effects against DSS-induced mice ulcerative colitis in intestinal barrier-dependent manner. *Molecules* 26, 6604. doi:10.3390/molecules26216604
- Cao, S., Ye, S., Wang, W., Wang, B., Zhang, T., and Pu, Y. (2019). Progress in active compounds effective on ulcerative colitis from Chinese medicines. *Chin. J. Nat. Med.* 17, 81–102. doi:10.1016/s1875-5364(19)30012-3
- Chamanara, M., Abdollahi, A., Rezayat, S. M., Ghazi-Khansari, M., Dehpour, A., Nassireslami, E., et al. (2019). Thymol reduces acetic acid-induced inflammatory response through inhibition of NF- κ B signaling pathway in rat colon tissue. *Inflammopharmacology* 27, 1275–1283. doi:10.1007/s10787-019-00583-8
- Che, L., Li, Y., Song, R., Qin, C., Hao, W., Wang, B., et al. (2019). Anti-inflammatory and anti-apoptosis activity of taraxasterol in ulcerative colitis *in vitro* and *in vivo*. *Exp. Ther. Med.* 18, 1745–1751. doi:10.3892/etm.2019.7736
- Chen, C., Liang, H., Wang, J., Ren, G., Li, R., Cui, Z.-G., et al. (2022a). Heterophyllin B an active cyclopeptide alleviates dextran sulfate sodium-induced colitis by modulating gut microbiota and repairing intestinal mucosal barrier via AMPK activation. *Mol. Nutr. Food Res.* 66, e2101169. doi:10.1002/mnfr.202101169
- Chen, L., Liu, D., Mao, M., Liu, W., Wang, Y., Liang, Y., et al. (2022b). Betaine ameliorates acute severe ulcerative colitis by inhibiting oxidative stress induced inflammatory pyroptosis. *Mol. Nutr. Food Res.* 66, e2200341. doi:10.1002/mnfr.202200341
- Chen, Y., Wang, J., Li, J., Zhu, J., Wang, R., Xi, Q., et al. (2021b). Astragalus polysaccharide prevents ferroptosis in a murine model of experimental colitis and human Caco-2 cells via inhibiting NRF2/HO-1 pathway. *Eur. J. Pharmacol.* 911, 174518. doi:10.1016/j.ejphar.2021.174518
- Chen, Y. E., Xu, S. J., Lu, Y. Y., Chen, S. X., Du, X. H., Hou, S. Z., et al. (2021a). Asperuloside suppressing oxidative stress and inflammation in DSS-induced chronic colitis and RAW 264.7 macrophages via Nrf2/HO-1 and NF- κ B pathways. *Chem. Biol. Interact.* 344, 109512. doi:10.1016/j.cbi.2021.109512
- Chen, Y.-X., Zhang, X.-Q., Yu, C.-G., Huang, S.-L., Xie, Y., Dou, X.-T., et al. (2019). Artesunate exerts protective effects against ulcerative colitis via suppressing Toll-like receptor 4 and its downstream nuclear factor-kappa B signaling pathways. *Mol. Med. Rep.* 20, 1321–1332. doi:10.3892/mmr.2019.10345
- Cheng, C., Zhang, W., Zhang, C., Ji, P., Wu, X., Sha, Z., et al. (2021). Hyperoside ameliorates DSS-induced colitis through MKRN1-mediated regulation of PPAR γ signaling and Th17/treg balance. *J. Agric. Food Chem.* 69, 15240–15251. doi:10.1021/acs.jafc.1c06292
- Cheng, H., Liu, J., Zhang, D., Wang, J., Tan, Y., Feng, W., et al. (2022a). Ginsenoside Rg1 alleviates acute ulcerative colitis by modulating gut microbiota and microbial tryptophan metabolism. *Front. Immunol.* 13, 817600. doi:10.3389/fimmu.2022.817600
- Cheng, J., Ma, X., Zhang, H., Wu, X., Li, M., Ai, G., et al. (2022b). 8-Oxypalmitate, a novel oxidative metabolite of palmitate, exhibits superior anti-colitis effect via regulating Nrf2 and NLRP3 inflammasome. *Biomed. Pharmacother.* 153, 113335. doi:10.1016/j.biopha.2022.113335
- Cheng, T., Xu, C., Wu, D., Yan, G., Wang, C., Wang, T., et al. (2023). Sodium houttuynonate derived from Houttuynia cordata Thunb improves intestinal malnutrition via maintaining gut microflora stability in *Candida albicans* overgrowth aggravated ulcerative colitis. *Food. Funct.* 14, 1072–1086. doi:10.1039/d2fo02369e
- Childers, R. E., Eluri, S., Vazquez, C., Weise, R. M., Bayless, T. M., and Hutfless, S. (2014). Family history of inflammatory bowel disease among patients with ulcerative colitis: a systematic review and meta-analysis. *J. Crohns Colitis* 8, 1480–1497. doi:10.1016/j.crohns.2014.05.008
- Cohen, R. D., Yu, A. P., Wu, E. Q., Xie, J., Mulani, P. M., and Chao, J. (2010). Systematic review: the costs of ulcerative colitis in Western countries. *Pharmacol. Ther.* 31, 693–707. doi:10.1111/j.1365-2036.2010.04234.x
- Conrad, K., Roggenbuck, D., and Laass, M. W. (2014). Diagnosis and classification of ulcerative colitis. *Autoimmun. Rev.* 13, 463–466. doi:10.1016/j.autrev.2014.01.028
- Consortium, U. I. G., Barrett, J. C., Lee, J. C., Lees, C. W., Prescott, N. J., Anderson, C. A., et al. (2009). Genome-wide association study of ulcerative colitis identifies three new susceptibility loci, including the HNF4A region. *Nat. Genet.* 41, 1330–1334. doi:10.1038/ng.483
- Cui, L., Feng, L., Zhang, Z. H., and Jia, X. B. (2014). The anti-inflammation effect of baicalin on experimental colitis through inhibiting TLR4/NF-kappa B pathway activation. *Int. Immunopharmacol.* 23, 294–303. doi:10.1016/j.intimp.2014.09.005
- da Silva, B. C., Lyra, A. C., Rocha, R., and Santana, G. O. (2014). Epidemiology, demographic characteristics and prognostic predictors of ulcerative colitis. *World J. Gastroenterol.* 20, 9458–9467. doi:10.3748/wjg.v20.i28.9458
- Diwan, B., and Sharma, R. (2022). Green tea EGCG effectively alleviates experimental colitis in middle-aged male mice by attenuating multiple aspects of oxi-inflammatory stress and cell cycle deregulation. *Biogerontology* 23, 789–807. doi:10.1007/s10522-022-09976-9
- Dixon, S. J., Lemberg, K. M., Lamprecht, M. R., Skouta, R., Zaitsev, E. M., Gleason, C. E., et al. (2012). Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* 149, 1060–1072. doi:10.1016/j.cell.2012.03.042
- Dong, L., Du, H., Zhang, M., Xu, H., Pu, X., Chen, Q., et al. (2022b). Anti-inflammatory effect of Rhein on ulcerative colitis via inhibiting PI3K/Akt/mTOR signaling pathway and regulating gut microbiota. *Phytother. Res.* 36, 2081–2094. doi:10.1002/ptr.7429
- Dong, S., Lu, Y., Peng, G., Li, J., Li, W., Li, M., et al. (2021). Furin inhibits epithelial cell injury and alleviates experimental colitis by activating the Nrf2-Gpx4 signaling pathway. *Dig. Liver Dis.* 53, 1276–1285. doi:10.1016/j.dld.2021.02.011
- Dong, Y., Fan, H., Zhang, Z., Jiang, F., Li, M., Zhou, H., et al. (2022c). Berberine ameliorates DSS-induced intestinal mucosal barrier dysfunction through microbiota-dependence and Wnt/ β -catenin pathway. *Int. J. Biol. Sci.* 18, 1381–1397. doi:10.7150/ijbs.65476
- Dong, Y., Huang, C., Yang, J., Zheng, Z., and Dai, Z. (2022a). Docosapentaenoic acid (DPA, 22:5n-3) alleviates ulcerative colitis via modification of gut microbiota and their metabolism. *Nutrients* 14, 4204. doi:10.3390/nu14194204
- Dou, B., Hu, W., Song, M., Lee, R. J., Zhang, X., and Wang, D. (2020). Anti-inflammation of Erianin in dextran sulphate sodium-induced ulcerative colitis mice model via collaborative regulation of TLR4 and STAT3. *Chemico-Biological Interact.* 324, 109089. doi:10.1016/j.cbi.2020.109089
- Dou, W., Zhang, J., Sun, A., Zhang, E., Ding, L., Mukherjee, S., et al. (2013). Protective effect of naringenin against experimental colitis via suppression of Toll-like receptor 4/NF-kappa B signalling. *Br. J. Nutr.* 110, 599–608. doi:10.1017/s0007114512005594
- Dou, Y. X., Zhou, J. T., Wang, T. T., Huang, Y. F., Chen, V. P., Xie, Y. L., et al. (2018). Self-nanoemulsifying drug delivery system of bruceine D: a new approach for anti-ulcerative colitis. *Int. J. Nanomedicine* 13, 5887–5907. doi:10.2147/IJN.174146

- Du, G., Xiong, L., Li, X., Zhuo, Z., Zhuang, X., Yu, Z., et al. (2020). Peroxisome elevation induces stem cell differentiation and intestinal epithelial repair. *Dev. Cell* 53, 169–184. doi:10.1016/j.devcel.2020.03.002
- Duan, S., Du, X., Chen, S., Liang, J., Huang, S., Hou, S., et al. (2020). Effect of vitexin on alleviating liver inflammation in a dextran sulfate sodium (DSS)-induced colitis model. *Biomed. Pharmacother.* 121, 109683. doi:10.1016/j.biopha.2019.109683
- Elmaksoud, H. A. A., Motawea, M. H., Desoky, A. A., Elharrif, M. G., and Ibrahim, A. (2021). Hydroxytyrosol alleviate intestinal inflammation, oxidative stress and apoptosis resulted in ulcerative colitis. *Biomed. Pharmacother.* 142, 112073. doi:10.1016/j.biopha.2021.112073
- Fang, J., Wang, H., Zhou, Y., Zhang, H., Zhou, H., and Zhang, X. (2021). Slimy partners: the mucus barrier and gut microbiome in ulcerative colitis. *Exp. Mol. Med.* 53, 772–787. doi:10.1038/s12276-021-00617-8
- Fang, W., Zhu, S., Niu, Z., and Yin, Y. (2019). The protective effect of syringic acid on dextran sulfate sodium-induced experimental colitis in BALB/c mice. *Drug Dev. Res.* 80, 731–740. doi:10.1002/ddr.21524
- Feng, J. S., Guo, C. C., Zhu, Y. Z., Pang, L. P., Yang, Z., Zou, Y., et al. (2014). Baicalin down regulates the expression of TLR4 and NF- κ B-p65 in colon tissue in mice with colitis induced by dextran sulfate sodium. *Int. J. Clin. Exp. Med.* 7, 4063–4072.
- Feng, Z., Zhou, P., Wu, X., Zhang, J., and Zhang, M. (2022). Hydroxysafflor yellow A protects against ulcerative colitis via suppressing TLR4/NF- κ B signaling pathway. *Chem. Biol. Drug Des.* 99, 897–907. doi:10.1111/cbdd.14045
- Foerster, E. G., Mukherjee, T., Cabral-Fernandes, L., Rocha, J. D. B., Girardin, S. E., and Philpott, D. J. (2022). How autophagy controls the intestinal epithelial barrier. *Autophagy* 18, 86–103. doi:10.1080/15548627.2021.1909406
- Formentini, L., Santacatterina, F., Nunez de Arenas, C., Stamatakis, K., Lopez-Martinez, D., Logan, A., et al. (2017). Mitochondrial ROS production protects the intestine from inflammation through functional M2 macrophage polarization. *Cell Rep.* 19, 1202–1213. doi:10.1016/j.celrep.2017.04.036
- Formiga, R. O., Alves Junior, E. B., Vasconcelos, R. C., Bernardo Guerra, G. C., de Araujo, A. A., de Carvalho, T. G., et al. (2020). p-Cymene and rosmarinic acid ameliorate TNBS-induced intestinal inflammation upkeeping ZO-1 and MUC-2: role of antioxidant system and immunomodulation. *Int. J. Mol. Sci.* 21, 5870. doi:10.3390/ijms21165870
- Fu, C., Hao, S., Xu, X., Zhou, J., Liu, Z., Lu, H., et al. (2019). Activation of SIRT1 ameliorates LPS-induced lung injury in mice via decreasing endothelial tight junction permeability. *Acta Pharmacol. Sin.* 40, 630–641. doi:10.1038/s41401-018-0045-3
- Fu, R., Wang, L., Meng, Y., Xue, W., Liang, J., Peng, Z., et al. (2022). Apigenin remodels the gut microbiota to ameliorate ulcerative colitis. *Front. Nutr.* 9, 1062961. doi:10.3389/fnut.2022.1062961
- Gai, L., Chu, L., Xia, R., Chen, Q., and Sun, X. (2019). Barbaloin attenuates mucosal damage in experimental models of rat colitis by regulating inflammation and the AMPK signaling pathway. *Med. Sci. Monit.* 25, 10045–10056. doi:10.12659/msm.918935
- Gao, H., Zhou, H., Zhang, Z., Gao, J., Li, J., and Li, X. (2023). Vitamin D3 alleviates inflammation in ulcerative colitis by activating the VDR-NLRP6 signaling pathway. *Front. Immunol.* 14, 1135930. doi:10.3389/fimmu.2023.1135930
- Gao, W., Wang, C., Yu, L., Sheng, T., Wu, Z., Wang, X., et al. (2019a). Chlorogenic acid attenuates dextran sulfate sodium-induced ulcerative colitis in mice through MAPK/ERK/JNK pathway. *Biomed. Res. Int.* 2019, 6769789. doi:10.1155/2019/6769789
- Gao, W. Y., Zhang, L. D., Wang, X. Q., Yu, L., Wang, C. H., and Gong, Y. (2018). The combination of indirubin and isatin attenuates dextran sulfate sodium-induced ulcerative colitis in mice. *Biochem. Cell Biol.* 96, 636–645. doi:10.1139/bcb-2018-0041
- Gao, X.-J., Tang, B., Liang, H.-H., Yi, L., and Wei, Z.-G. (2019b). The protective effect of nigezanine on dextran sulfate sodium-induced experimental colitis in mice and Caco-2 cells. *J. Cell. Physiol.* 234, 23398–23408. doi:10.1002/jcp.28909
- Giris, M., Depboylu, B., Dogru-Abbasoglu, S., Erbil, Y., Olgac, V., Alis, H., et al. (2008). Effect of taurine on oxidative stress and apoptosis-related protein expression in trinitrobenzene sulphonic acid-induced colitis. *Clin. Exp. Immunol.* 152, 102–110. doi:10.1111/j.1365-2249.2008.03599.x
- Glassner, K. L., Abraham, B. P., and Quigley, E. M. M. (2020). The microbiome and inflammatory bowel disease. *J. Allergy Clin. Immunol.* 145, 16–27. doi:10.1016/j.jaci.2019.11.003
- Gu, P., Zhu, L., Liu, Y., Zhang, L., Liu, J., and Shen, H. (2017). Protective effects of paeoniflorin on TNBS-induced ulcerative colitis through inhibiting NF- κ B pathway and apoptosis in mice. *Int. Immunopharmacol.* 50, 152–160. doi:10.1016/j.intimp.2017.06.022
- Guo, G., Shi, F., Zhu, J., Shao, Y., Gong, W., Zhou, G., et al. (2020). Piperine, a functional food alkaloid, exhibits inhibitory potential against TNBS-induced colitis via the inhibition of I κ B- α /NF- κ B and induces tight junction protein (claudin-1, occludin, and ZO-1) signaling pathway in experimental mice. *Hum. Exp. Toxicol.* 39, 477–491. doi:10.1177/0960327119892042
- Guo, R., Meng, Q., Wang, B., and Li, F. (2021). Anti-inflammatory effects of Platycodin D on dextran sulfate sodium (DSS) induced colitis and *E. coli* Lipopolysaccharide (LPS) induced inflammation. *Int. Immunopharmacol.* 94, 107474. doi:10.1016/j.intimp.2021.107474
- Guo, W., Wang, X., Liu, F., Chen, S., Wang, S., Zhang, Q., et al. (2022). Acteoside alleviates dextran sulphate sodium-induced ulcerative colitis via regulation of the HO-1/HMGB1 signaling pathway. *Mol. Med. Rep.* 26, 360. doi:10.3892/mmr.2022.12877
- Gutierrez-Orozco, F., Thomas-Ahner, J. M., Berman-Booty, L. D., Galley, J. D., Chitchumroonchokchai, C., Mace, T., et al. (2014). Dietary α -mangostin, a xanthone from mangosteen fruit, exacerbates experimental colitis and promotes dysbiosis in mice. *Mol. Nutr. Food Res.* 58, 1226–1238. doi:10.1002/mnfr.201300771
- Guvenc, M., Cellat, M., Ozkan, H., Tekeli, I. O., Uyar, A., Gokcek, I., et al. (2019). Protective effects of tyrosol against DSS-induced ulcerative colitis in rats. *Inflammation* 42, 1680–1691. doi:10.1007/s10753-019-01028-8
- Hagan, M., Hayee, B. H., and Rodriguez-Mateos, A. (2021). (Poly)phenols in inflammatory bowel disease and irritable bowel syndrome: a review. *Molecules* 26, 1843. doi:10.3390/molecules26071843
- Han, J., Li, W., Shi, G., Huang, Y., Sun, X., Sun, N., et al. (2022). Atractylenolide III improves mitochondrial function and protects against ulcerative colitis by activating AMPK/SIRT1/PGC-1 α . *Mediat. Inflamm.* 2022, 9129984. doi:10.1155/2022/9129984
- Hauso, Ø., Martinsen, T. C., and Waldum, H. (2015). 5-Aminosalicylic acid, a specific drug for ulcerative colitis. *Scand. J. Gastroenterol.* 50, 933–941. doi:10.3109/00365521.2015.1018937
- He, J., Liang, J., Zhu, S., Zhao, W., Zhang, Y., and Sun, W. (2011). Protective effect of taurohydroxychoyolic acid from *Pulsis Fellis Suis* on trinitrobenzene sulfonic acid induced ulcerative colitis in mice. *Eur. J. Pharmacol.* 670, 229–235. doi:10.1016/j.ejphar.2011.08.036
- He, J., Liu, L., Liu, X., Chen, H., Liu, K., Huang, N., et al. (2022). Epoxymicheliolide prevents dextran sulfate sodium-induced colitis in mice by inhibiting TAK1-NF- κ B pathway and activating Keap1-NRF2 signaling in macrophages. *Int. Immunopharmacol.* 113, 109404. doi:10.1016/j.intimp.2022.109404
- He, X., Liu, J., Long, G., Xia, X.-H., and Liu, M. (2021). 2,3,5,4'-Tetrahydroxystilbene-2-O- β -D-glucoside, a major bioactive component from *Polygoni multiflori Radix* (Heshouwu) suppresses DSS induced acute colitis in BALB/c mice by modulating gut microbiota. *Biomed. Pharmacother.* 137, 111420. doi:10.1016/j.biopha.2021.111420
- He, X., Zheng, Z., Yang, X., Lu, Y., Chen, N., and Chen, W. (2012). Tetramethylpyrazine attenuates PPAR- γ antagonist-deteriorated oxazolone-induced colitis in mice. *Mol. Med. Rep.* 5, 645–650. doi:10.3892/mmr.2011.721
- Hoivik, M. L., Moum, B., Solberg, I. C., Henriksen, M., Cvancarova, M., Bernklev, T., et al. (2013). Work disability in inflammatory bowel disease patients 10 years after disease onset: results from the IBSEN Study. *Gut* 62, 368–375. doi:10.1136/gutjnl-2012-302311
- Hossen, I., Hua, W., Ting, L., Mehmood, A., Jingyi, S., Duoxia, X., et al. (2020). Phytochemicals and inflammatory bowel diseases: a review. *Crit. Rev. Food Sci. Nutr.* 60, 1321–1345. doi:10.1080/10408398.2019.1570913
- Hou, Y.-C., Chu, C.-C., Ko, T.-L., Yeh, C.-L., and Yeh, S.-L. (2013). Effects of alanyl-glutamine dipeptide on the expression of colon-inflammatory mediators during the recovery phase of colitis induced by dextran sulfate sodium. *Eur. J. Nutr.* 52, 1089–1098. doi:10.1007/s00394-012-0416-3
- Hu, L., Wu, C., Zhang, Z., Liu, M., Prasad, E. M., Chen, Y., et al. (2019). Pinocembrin protects against dextran sulfate sodium-induced rats colitis by ameliorating inflammation, improving barrier function and modulating gut microbiota. *Front. Physiol.* 10, 908. doi:10.3389/fphys.2019.00908
- Hu, L.-H., Liu, J.-Y., and Yin, J.-B. (2021). Eriodictyol attenuates TNBS-induced ulcerative colitis through repressing TLR4/NF- κ B signaling pathway in rats. *Kaohsiung J. Med. Sci.* 37, 812–818. doi:10.1002/kjm.212400
- Hu, X., He, X., Peng, C., He, Y., Wang, C., Tang, W., et al. (2022). Improvement of ulcerative colitis by aspartate via RIPK pathway modulation and gut microbiota composition in mice. *Nutrients* 14, 3707. doi:10.3390/nu14183707
- Hua, Y., Liu, R., Lu, M., Guan, X., Zhuang, S., Tian, Y., et al. (2021). Juglone regulates gut microbiota and Th17/Treg balance in DSS-induced ulcerative colitis. *Int. Immunopharmacol.* 97, 107683. doi:10.1016/j.intimp.2021.107683
- Huang, B., Wang, L., Liu, M., Wu, X., Lu, Q., and Liu, R. (2022a). The underlying mechanism of A-type procyanidins from peanut skin on DSS-induced ulcerative colitis mice by regulating gut microbiota and metabolism. *J. Food Biochem.* 46, e14103. doi:10.1111/jfbc.14103
- Huang, J., Zhang, J., Ma, J., Ma, J., Liu, J., Wang, F., et al. (2022b). Inhibiting ferroptosis: a novel approach for ulcerative colitis therapeutics. *Oxid. Med. Cell. Longev.* 2022, 9678625. doi:10.1155/2022/9678625
- Huang, S., Fu, Y., Xu, B., Liu, C., Wang, Q., Luo, S., et al. (2020). Wogonoside alleviates intestinal epithelial barrier function via the MLCK/pMLC2 pathway. *Phytomedicine* 68, 153179. doi:10.1016/j.phymed.2020.153179
- Huang, X. R., Wang, C. D., Wang, R. X., and Jiao, H. X. (2016). Changes of colonic permeability and its correlation with TNF- α , NF- κ B p65 in ulceration colitis mice. *Chin. J. Appl. Physiol.* 32, 112–115. doi:10.13459/j.cnki.cjap.2016.02.005
- Islam, J., Sato, S., Watanabe, K., Watanabe, T., Ardiansyah, H. K., Aoyama, Y., et al. (2017). Dietary tryptophan alleviates dextran sodium sulfate-induced colitis through

- aryl hydrocarbon receptor in mice. *J. Nutr. Biochem.* 42, 43–50. doi:10.1016/j.jnutbio.2016.12.019
- Jeon, Y.-D., Lee, J.-H., Lee, Y.-M., and Kim, D.-K. (2020). Puerarin inhibits inflammation and oxidative stress in dextran sulfate sodium-induced colitis mice model. *Biomed. Pharmacother.* 124, 109847. doi:10.1016/j.biopha.2020.109847
- Jha, N. K., Sharma, C., Hashiesh, H. M., Arunachalam, S., Meeran, M. N., Javed, H., et al. (2021). β -Caryophyllene, A natural dietary CB2 receptor selective cannabinoid can be a candidate to target the trinity of infection, immunity, and inflammation in COVID-19. *Front. Pharmacol.* 12, 590201. doi:10.3389/fphar.2021.590201
- Jia, L., Xue, K., Liu, J., Habotta, O. A., Hu, L., and Abdel Moneim, A. E. (2020). Anticolitic effect of berberine in rat experimental model: impact of PGE2/p38 MAPK pathways. *Mediat. Inflamm.* 2020, 9419085. doi:10.1155/2020/9419085
- Jia, X., Gao, Y., Liu, L., Guo, Y., Wang, J., Ma, H., et al. (2022). Artemisinin alleviates intestinal inflammation and metabolic disturbance in ulcerative colitis rats induced by DSS. *Evid. Based Complement. Altern. Med.* 2022, 6211215. doi:10.1155/2022/6211215
- Jiang, H., Shi, G.-F., Fang, Y.-X., Liu, Y.-Q., Wang, Q., Zheng, X., et al. (2022). Aloin A prevents ulcerative colitis in mice by enhancing the intestinal barrier function via suppressing the Notch signaling pathway. *Phytomedicine* 106, 154403. doi:10.1016/j.phymed.2022.154403
- Jiang, M., Zhong, G., Zhu, Y., Wang, L., He, Y., Sun, Q., et al. (2021b). Retardant effect of dihydroartemisinin on ulcerative colitis in a JAK2/STAT3-dependent manner. *Acta Biochim. Biophys. Sin.* 53, 1113–1123. doi:10.1093/abbs/gmab097
- Jiang, Y., Zhao, L., Chen, Q., and Zhou, L. (2021a). Exploring the mechanism of berberine intervention in ulcerative colitis from the perspective of inflammation and immunity based on systemic pharmacology. *Evid. Based Complement. Altern. Med.* 2021, 9970240. doi:10.1155/2021/9970240
- Johansson, M. E. V., Ambort, D., Pelaseyed, T., Schütte, A., Gustafsson, J. K., Ermund, A., et al. (2011). Composition and functional role of the mucus layers in the intestine. *Cell. Mol. Life Sci.* 68, 3635–3641. doi:10.1007/s00018-011-0822-3
- Johansson, M. E. V., Gustafsson, J. K., Holmen-Larsson, J., Jabbar, K. S., Xia, L. J., Xu, H., et al. (2014). Bacteria penetrate the normally impenetrable inner colon mucus layer in both murine colitis models and patients with ulcerative colitis. *Gut* 63, 281–291. doi:10.1136/gutjnl-2012-303207
- Jostins, L., Ripke, S., Weersma, R. K., Duerr, R. H., McGovern, D. P., Hui, K. Y., et al. (2012). Host–microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* 491, 119–124. doi:10.1038/nature11582
- Kanneganti, T. D., Lamkanfi, M., and Nunez, G. (2007). Intracellular NOD-like receptors in host defense and disease. *Immunity* 27, 549–559. doi:10.1016/j.immuni.2007.10.002
- Kaser, A., Martinez-Naves, E., and Blumberg, R. S. (2010). Endoplasmic reticulum stress: implications for inflammatory bowel disease pathogenesis. *Curr. Opin. Gastroenterol.* 26, 318–326. doi:10.1097/MOG.0b013e32833a9ff1
- Kondo, K., Hiramoto, K., Yamate, Y., Goto, K., Sekijima, H., and Ooi, K. (2019). Ameliorative effect of high-dose vitamin C administration on dextran sulfate sodium-induced colitis mouse model. *Biol. Pharm. Bull.* 42, 954–959. doi:10.1248/bpb.b18-00967
- Kook, S.-H., Choi, K. C., Cho, S.-W., Chod, H.-K., Lee, K. D., and Lee, J.-C. (2015). Catechin-7-O- β -D-glucopyranoside isolated from the seed of *Phaseolus calcaratus* Roxburgh ameliorates experimental colitis in rats. *Int. Immunopharmacol.* 29, 521–527. doi:10.1016/j.intimp.2015.10.003
- Koppel, N., Rekdal, V. M., and Balskus, E. P. (2017). Chemical transformation of xenobiotics by the human gut microbiota. *Science* 356, eaag2770. doi:10.1126/science.aag2770
- Kostic, A. D., Xavier, R. J., and Gevers, D. (2014). The microbiome in inflammatory bowel disease: current status and the future ahead. *Gastroenterology* 146, 1489–1499. doi:10.1053/j.gastro.2014.02.009
- Kotipalli, R. S. S., Tirunavalli, S. K., Pote, A. B., Sahu, B. D., Kuncha, M., Jerald, M. K., et al. (2023). Sinigrin attenuates the dextran sulfate sodium-induced colitis in mice by modulating the MAPK pathway. *Inflammation* 46, 787–807. doi:10.1007/s10753-022-01780-4
- Krndija, D., El Marjoui, F., Guirao, B., Richon, S., Leroy, O., Bellaiche, Y., et al. (2019). Active cell migration is critical for steady-state epithelial turnover in the gut. *Science* 365, 705–710. doi:10.1126/science.aau3429
- Kuhnert, F., Davis, C. R., Wang, H. T., Chu, P., Lee, M., Yuan, J., et al. (2004). Essential requirement for Wnt signaling in proliferation of adult small intestine and colon revealed by adenoviral expression of Dickkopf-1. *Proc. Natl. Acad. Sci. U. S. A.* 101, 266–271. doi:10.1073/pnas.2536800100
- Larabi, A., Barnich, N., and Nguyen, H. T. T. (2020). New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy* 16, 38–51. doi:10.1080/15548627.2019.1635384
- Laukens, D., Devisscher, L., Van den Bossche, L., Hindryckx, P., Vandenbroucke, R. E., Vandewynckel, Y. P., et al. (2014). Tauroursodeoxycholic acid inhibits experimental colitis by preventing early intestinal epithelial cell death. *Lab. Invest.* 94, 1419–1430. doi:10.1038/labinvest.2014.117
- Li, B., Du, P., Du, Y., Zhao, D., Cai, Y., Yang, Q., et al. (2021a). Luteolin alleviates inflammation and modulates gut microbiota in ulcerative colitis rats. *Life Sci.* 269, 119008. doi:10.1016/j.lfs.2020.119008
- Li, C., Dong, N., Wu, B., Mo, Z., Xie, J., and Lu, Q. (2021b). Dihydroberberine, an isoquinoline alkaloid, exhibits protective effect against dextran sulfate sodium-induced ulcerative colitis in mice. *Phytomedicine* 90, 153631. doi:10.1016/j.phymed.2021.153631
- Li, C., Zhang, W., Wu, X., Cai, Q., Tan, Z., Hong, Z., et al. (2022c). Aromatic-turmerone ameliorates DSS-induced ulcerative colitis via modulating gut microbiota in mice. *Inflammopharmacology* 30, 1283–1294. doi:10.1007/s10787-022-01007-w
- Li, H., Fan, C., Lu, H., Feng, C., He, P., Yang, X., et al. (2020b). Protective role of berberine on ulcerative colitis through modulating enteric glial cells-intestinal epithelial cells-immune cells interactions. *Acta Pharm. Sin. B* 10, 447–461. doi:10.1016/j.apsb.2019.08.006
- Li, H., Pang, B., Nie, B., Qu, S., Zhang, K., Xu, J., et al. (2022g). Dioscin promotes autophagy by regulating the AMPK-mTOR pathway in ulcerative colitis. *Immunopharmacol. Immunotoxicol.* 44, 238–246. doi:10.1080/08923973.2022.2037632
- Li, M., Guo, W., Dong, Y., Wang, W., Tian, C., Zhang, Z., et al. (2022e). Beneficial effects of celastrol on immune balance by modulating gut microbiota in experimental ulcerative colitis mice. *Genomics Proteomics Bioinforma.* 20, 288–303. doi:10.1016/j.gpb.2022.05.002
- Li, M. Y., Zhang, Z. H., Wang, Z., Zuo, H. X., Wang, J. Y., Xing, Y., et al. (2019b). Convallatoxin protects against dextran sulfate sodium-induced experimental colitis in mice by inhibiting NF- κ B signaling through activation of PPAR γ . *Pharmacol. Res.* 147, 104355. doi:10.1016/j.phrs.2019.104355
- Li, N., Sun, W., Zhou, X., Gong, H., Chen, Y., Chen, D., et al. (2019a). Dihydroartemisinin protects against dextran sulfate sodium-induced colitis in mice through inhibiting the PI3K/AKT and NF- κ B signaling pathways. *Biomed. Res. Int.* 2019, 1415809–1415812. doi:10.1155/2019/1415809
- Li, P., Wu, M., Xiong, W., Li, J., An, Y., Ren, J., et al. (2020a). Saikosaponin-d ameliorates dextran sulfate sodium-induced colitis by suppressing NF- κ B activation and modulating the gut microbiota in mice. *Int. Immunopharmacol.* 81, 106288. doi:10.1016/j.intimp.2020.106288
- Li, R., Chen, C., Liu, B., Shi, W., Shimizu, K., and Zhang, C. (2022f). Bryodulcosigenin a natural cucurbitane-type triterpenoid attenuates dextran sulfate sodium (DSS)-induced colitis in mice. *Phytomedicine* 94, 153814. doi:10.1016/j.phymed.2021.153814
- Li, T. T., Zhang, J. F., Fei, S. J., Zhu, S. P., Zhu, J. Z., Qiao, X., et al. (2014). Glutamate microinjection into the hypothalamic paraventricular nucleus attenuates ulcerative colitis in rats. *Acta Pharmacol. Sin.* 35, 185–194. doi:10.1038/aps.2013.140
- Li, W., Zhang, L., Xu, Q., Yang, W., Zhao, J., Ren, Y., et al. (2022d). Taxifolin alleviates DSS-induced ulcerative colitis by acting on gut microbiome to produce butyric acid. *Nutrients* 14, 1069. doi:10.3390/nu14051069
- Li, X., Wu, X., Wang, Q., Xu, W., Zhao, Q., Xu, N., et al. (2022b). Sanguinarine ameliorates DSS induced ulcerative colitis by inhibiting NLRP3 inflammasome activation and modulating intestinal microbiota in C57BL/6 mice. *Phytomedicine* 104, 154321. doi:10.1016/j.phymed.2022.154321
- Li, X., Xu, S., Zhang, Y., Li, K., Gao, X.-J., and Guo, M.-Y. (2022a). Berberine depresses inflammation and adjusts smooth muscle to ameliorate ulcerative colitis of cats by regulating gut microbiota. *Microbiol. Spectr.* 10, e0320722. doi:10.1128/spectrum.03207-22
- Li, Y., Pan, X., Yin, M., Li, C., and Han, L. (2021c). Preventive effect of lycopene in dextran sulfate sodium-induced ulcerative colitis mice through the regulation of TLR4/TRIF/NF- κ B signaling pathway and tight junctions. *J. Agric. Food Chem.* 69, 13500–13509. doi:10.1021/acs.jafc.1c05128
- Liang, H., Cheng, R., Wang, J., Xie, H., Li, R., Shimizu, K., et al. (2021). Mogrol, an aglycone of mogrosides, attenuates ulcerative colitis by promoting AMPK activation. *Phytomedicine* 81, 153427. doi:10.1016/j.phymed.2020.153427
- Liang, S., Deng, X., Lei, L., Zheng, Y., Ai, J., Chen, L., et al. (2019). The comparative study of the therapeutic effects and mechanism of baicalin, baicalein, and their combination on ulcerative colitis rat. *Front. Pharmacol.* 10, 1466. doi:10.3389/fphar.2019.01466
- Lifei, L., Zhang, J., Li, X., Zhu, Y., Wang, Y., and Liu, D. (2023). Sericic acid ameliorates DSS-induced ulcerative colitis in mice by modulating the NF- κ B and Nrf2 pathways. *Curr. Mol. Pharmacol.* 16, 759–770. doi:10.2174/1874467215666220928100319
- Lin, Y., Jiang, M., Chen, W., Zhao, T., and Wei, Y. (2019). Cancer and ER stress: mutual crosstalk between autophagy, oxidative stress and inflammatory response. *Biomed. Pharmacother.* 118, 109249. doi:10.1016/j.biopha.2019.109249
- Lin, Y., Wu, Y., Su, J., Wang, M., Wu, X., Su, Z., et al. (2021). Therapeutic role of d-pinitol on experimental colitis via activating Nrf2/ARE and PPAR- γ /NF- κ B signaling pathways. *Food. Funct.* 12, 2554–2568. doi:10.1039/d0fo03139a
- Liu, B., Piao, X., Guo, L., Liu, S., Chai, F., and Gao, L. (2016). Ursolic acid protects against ulcerative colitis via anti-inflammatory and antioxidant effects in mice. *Mol. Med. Rep.* 13, 4779–4785. doi:10.3892/mmr.2016.5094

- Liu, C., Wang, J., Yang, Y., Liu, X., Zhu, Y., Zou, J., et al. (2018b). Ginsenoside Rd ameliorates colitis by inducing p62-driven mitophagy-mediated NLRP3 inflammasome inactivation in mice. *Biochem. Pharmacol.* 155, 366–379. doi:10.1016/j.bcp.2018.07.010
- Liu, D., Huo, X., Gao, L., Zhang, J., Ni, H., and Cao, L. (2018a). NF- κ B and Nrf2 pathways contribute to the protective effect of Licochalcone A on dextran sulphate sodium-induced ulcerative colitis in mice. *Biomed. Pharmacother.* 102, 922–929. doi:10.1016/j.biopha.2018.03.130
- Liu, D., Tian, Q., Liu, K., Ren, F., Liu, G., Zhou, J., et al. (2023c). Ginsenoside Rg3 ameliorates DSS-induced colitis by inhibiting NLRP3 inflammasome activation and regulating microbial homeostasis. *J. Agric. Food Chem.* 71, 3472–3483. doi:10.1021/acs.jafc.2c07766
- Liu, F., Yao, Y., Wang, Q., Zhang, F., Wang, M., Zhu, C., et al. (2023b). Nigakionone alleviates DSS-induced experimental colitis via regulating bile acid profile and FXR/NLRP3 signaling pathways. *Phytother. Res.* 37, 15–34. doi:10.1002/ptr.7588
- Liu, H., Xiong, J., He, T., Xiao, T., Li, Y., Yu, Y., et al. (2017). High uric acid-induced epithelial-mesenchymal transition of renal tubular epithelial cells via the TLR4/NF- κ B signaling pathway. *Am. J. Nephrol.* 46, 333–342. doi:10.1159/000481668
- Liu, J., Cai, J., Fan, P., Zhang, N., and Cao, Y. (2019). The abilities of salidroside on ameliorating inflammation, skewing the imbalanced nucleotide oligomerization domain-like receptor family pyrin domain containing 3/autophagy, and maintaining intestinal barrier are profitable in colitis. *Front. Pharmacol.* 10, 1385. doi:10.3389/fphar.2019.01385
- Liu, J., Shi, L., Huang, W., Zheng, Z., Huang, X., and Su, Y. (2022b). Homoharringtonine attenuates dextran sulfate sodium-induced colitis by inhibiting NF- κ B signaling. *Mediat. Inflamm.* 2022, 3441357. doi:10.1155/2022/3441357
- Liu, J. Z., van Sommeren, S., Huang, H., Ng, S. C., Alberts, R., Takahashi, A., et al. (2015). Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat. Genet.* 47, 979–986. doi:10.1038/ng.3359
- Liu, K., Li, G., Guo, W., and Zhang, J. (2020c). The protective effect and mechanism of pedunculoside on DSS (dextran sulfate sodium) induced ulcerative colitis in mice. *Int. Immunopharmacol.* 88, 107017. doi:10.1016/j.intimp.2020.107017
- Liu, S., Shen, H., Li, J., Gong, Y., Bao, H., Zhang, J., et al. (2020b). Loganin inhibits macrophage M1 polarization and modulates sirt1/NF- κ B signaling pathway to attenuate ulcerative colitis. *Bioengineered* 11, 628–639. doi:10.1080/21655979.2020.1774992
- Liu, X., Zhou, M., Dai, Z., Luo, S., Shi, Y., He, Z., et al. (2023a). Salidroside alleviates ulcerative colitis via inhibiting macrophage pyroptosis and repairing the dysbacteriosis-associated Th17/Treg imbalance. *Phytother. Res.* 37, 367–382. doi:10.1002/ptr.7636
- Liu, Y., Li, B. G., Su, Y. H., Zhao, R. X., Song, P., Li, H., et al. (2022a). Potential activity of traditional Chinese medicine against ulcerative colitis: a review. *J. Ethnopharmacol.* 289, 115084. doi:10.1016/j.jep.2022.115084
- Liu, Y., Wu, J., Chen, L., Wu, X., Gan, Y., Xu, N., et al. (2020a). β -patchoulene simultaneously ameliorated dextran sulfate sodium-induced colitis and secondary liver injury in mice via suppressing colonic leakage and flora imbalance. *Biochem. Pharmacol.* 182, 114260. doi:10.1016/j.bcp.2020.114260
- Long, J., Liu, X. K., Kang, Z. P., Wang, M. X., Zhao, H. M., Huang, J. Q., et al. (2022). Ginsenoside Rg1 ameliorated experimental colitis by regulating the balance of M1/M2 macrophage polarization and the homeostasis of intestinal flora. *Eur. J. Pharmacol.* 917, 174742. doi:10.1016/j.ejphar.2022.174742
- Lopes de Oliveira, G. A., Alarcon-de-la-Lastra, C., Angeles Rosillo, M., Castejon Martinez, M. L., Sanchez-Hidalgo, M., Rolim Medeiros, J. V., et al. (2019). Preventive effect of berginin against the development of TNBS-induced acute colitis in rats is associated with inflammatory mediators inhibition and NLRP3/ASC inflammasome signaling pathways. *Chemico-Biological Interact.* 297, 25–33. doi:10.1016/j.cbi.2018.10.020
- Lu, Y., Chen, J., He, X., Xu, S., Chen, Y.-E., Gao, J., et al. (2021). Combined administration of vitamin D-3 and geniposide is less effective than single use of vitamin D-3 or geniposide in the treatment of ulcerative colitis. *Front. Pharmacol.* 12, 714065. doi:10.3389/fphar.2021.714065
- Luca, S. V., Macovei, I., Bujor, A., Miron, A., Skalicka-Wozniak, K., Aprotosoia, A. C., et al. (2020). Bioactivity of dietary polyphenols: the role of metabolites. *Crit. Rev. Food Sci. Nutr.* 60, 626–659. doi:10.1080/10408398.2018.1546669
- Lucena, A. M. M., Souza, C. R. M., Jales, J. T., Guedes, P. M. M., de Miranda, G. E. C., de Moura, A. M. A., et al. (2018). The bisindole alkaloid caulerpin, from seaweeds of the genus caulerpa, attenuated colon damage in murine colitis model. *Mar. Drugs.* 16, 318. doi:10.3390/md16090318
- Luo, M., and Luo, Y. (2021). Imperatorin relieved ulcerative colitis by regulating the nrf-2/ARE/HO-1 pathway in rats. *Inflammation* 44, 558–569. doi:10.1007/s10753-020-01353-3
- Luo, S., He, J., Huang, S., Wang, X., Su, Y., Li, Y., et al. (2022). Emodin targeting the colonic metabolism via PPAR gamma alleviates UC by inhibiting facultative anaerobe. *Phytomedicine* 104, 154106. doi:10.1016/j.phymed.2022.154106
- Lv, L., Chen, Z., Bai, W., Hao, J., Heng, Z., Meng, C., et al. (2023). Taurohyodeoxycholic acid alleviates trinitrobenzene sulfonic acid induced ulcerative colitis via regulating Th1/Th2 and Th17/Treg cells balance. *Life Sci.* 318, 121501. doi:10.1016/j.lfs.2023.121501
- Lv, Q., Xing, Y., Liu, Y., Chen, Q., Xu, J., Hu, L., et al. (2021). Didymin switches M1-like toward M2-like macrophage to ameliorate ulcerative colitis via fatty acid oxidation. *Pharmacol. Res.* 169, 105613. doi:10.1016/j.phrs.2021.105613
- Lv, T., Shen, L., Yang, L., Diao, W., Yang, Z., Zhang, Y., et al. (2018). Polydatin ameliorates dextran sulfate sodium-induced colitis by decreasing oxidative stress and apoptosis partially via Sonic hedgehog signaling pathway. *Int. Immunopharmacol.* 64, 256–263. doi:10.1016/j.intimp.2018.09.009
- Ma, J., Yin, G., Lu, Z., Xie, P., Zhou, H., Liu, J., et al. (2018). Casticin prevents DSS induced ulcerative colitis in mice through inhibitions of NF- κ B pathway and ROS signaling. *Phytother. Res.* 32, 1770–1783. doi:10.1002/ptr.6108
- Mahid, S. S., Minor, K. S., Soto, R. E., Hornung, C. A., and Galandiuk, S. (2006). Smoking and inflammatory bowel disease: a meta-analysis. *Mayo Clin. Proc.* 81, 1462–1471. doi:10.4065/81.11.1462
- Mai, C.-T., Wu, M.-M., Wang, C.-L., Su, Z.-R., Cheng, Y.-Y., and Zhang, X.-J. (2019). Palmatine attenuated dextran sulfate sodium (DSS)-induced colitis via promoting mitophagy-mediated NLRP3 inflammasome inactivation. *Mol. Immunol.* 105, 76–85. doi:10.1016/j.molimm.2018.10.015
- Maria-Ferreira, D., Nascimento, A. M., Cipriani, T. R., Santana, A. P., Watanabe, P. D., Ana, D., et al. (2018). Rhamnogalacturonan, a chemically-defined polysaccharide, improves intestinal barrier function in DSS-induced colitis in mice and human Caco-2 cells. *Sci. Rep.* 8, 12261. doi:10.1038/s41598-018-30526-2
- Martinon, F., Burns, K., and Tschopp, J. (2002). The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL- β . *Mol. Cell* 10, 417–426. doi:10.1016/s1097-2765(02)00599-3
- Miao, F. (2022). Hydroxytyrosol alleviates dextran sodium sulfate-induced colitis by inhibiting NLRP3 inflammasome activation and modulating gut microbiota *in vivo*. *Nutrition* 97, 111579. doi:10.1016/j.nut.2021.111579
- Miao, S., Lu, Q., Zhou, Y., Chang, Y., Xu, T., and Zhu, M. (2022). Oral administration of octacosanol modulates the gut bacteria and protects the intestinal barrier in ulcerative colitis mice. *J. Food Biochem.* 46, e14284. doi:10.1111/jfbc.14284
- Miao, Y., Lv, Q., Qiao, S., Yang, L., Tao, Y., Yan, W., et al. (2019). Alpinetin improves intestinal barrier homeostasis via regulating AhR/suv39h1/TSC2/mTORC1/autophagy pathway. *Toxicol. Appl. Pharmacol.* 384, 114772. doi:10.1016/j.taap.2019.114772
- Mostafa, A. F., Elalfy, M. M., Shata, A., and Elhadidy, M. G. (2020). Prophylactic effect of aquatic extract of stevia on acetic acid induced-ulcerative colitis in male rats: a possible role of Nrf2 and PPAR γ . *J. Basic Clin. Physiol. Pharmacol.* 32, 1093–1104. doi:10.1515/jbcp-2020-0039
- Nakarai, H., Yamashita, A., Nagayasu, S., Iwashita, M., Kumamoto, S., Ohyama, H., et al. (2012). Adipocyte-macrophage interaction may mediate LPS-induced low-grade inflammation: potential link with metabolic complications. *Innate Immun.* 18, 164–170. doi:10.1177/1753425910393370
- Nanki, K., Fujii, M., Shimokawa, M., Matano, M., Nishikori, S., Date, S., et al. (2019). Somatic inflammatory gene mutations in human ulcerative colitis epithelium. *Nature* 577, 254–259. doi:10.1038/s41586-019-1844-5
- Niu, W., Chen, Y., Wang, L., Li, J., Cui, Z., Lv, J., et al. (2022). The combination of sodium alginate and chlorogenic acid enhances the therapeutic effect on ulcerative colitis by the regulation of inflammation and the intestinal flora. *Food. Funct.* 13, 10710–10723. doi:10.1039/d2fo01619b
- Niu, X., Zhang, H., Li, W., Wang, Y., Mu, Q., Wang, X., et al. (2015). Protective effect of cavidine on acetic acid-induced murine colitis via regulating antioxidant, cytokine profile and NF- κ B signal transduction pathways. *Chem. Biol. Interact.* 239, 34–45. doi:10.1016/j.cbi.2015.06.026
- Ohkusa, T., Okayasu, I., Ogihara, T., Morita, K., Ogawa, M., and Sato, N. (2003). Induction of experimental ulcerative colitis by *Fusobacterium varium* isolated from colonic mucosa of patients with ulcerative colitis. *Gut* 52, 79–83. doi:10.1136/gut.52.1.79
- Pagano, E., Romano, B., Iannotti, F. A., Parisi, O. A., D'Armiento, M., Pignatiello, S., et al. (2019). The non-euphoric phytocannabinoid cannabidiol counteracts intestinal inflammation in mice and cytokine expression in biopsies from UC pediatric patients. *Pharmacol. Res.* 149, 104464. doi:10.1016/j.phrs.2019.104464
- Pan, H.-H., Zhou, X.-X., Ma, Y.-Y., Pan, W.-S., Zhao, F., Yu, M.-S., et al. (2020). Resveratrol alleviates intestinal mucosal barrier dysfunction in dextran sulfate sodium-induced colitis mice by enhancing autophagy. *World J. Gastroenterol.* 26, 4945–4959. doi:10.3748/wjg.v26.i33.4945
- Pan, Y. Y., Deng, Y., Su, S., Yin, J. H., Chen, Y. H., Wang, L. C., et al. (2023). Structure composition and intracellular transport of clathrin-mediated intestinal transmembrane tight junction protein. *Inflammation* 46, 18–34. doi:10.1007/s10753-022-01724-y
- Pang, B., Jin, H., Liao, N., Li, J., Jiang, C., and Shi, J. (2021). Vitamin A supplementation ameliorates ulcerative colitis in gut microbiota-dependent manner. *Food Res. Int.* 148, 110568. doi:10.1016/j.foodres.2021.110568
- Park, M.-Y., Ji, G. E., and Sung, M.-K. (2012). Dietary kaempferol suppresses inflammation of dextran sulfate sodium-induced colitis in mice. *Dig. Dis. Sci.* 57, 355–363. doi:10.1007/s10620-011-1883-8

- Pastorelli, L., Pizarro, T. T., Cominelli, F., and Vecchi, M. (2009). Emerging drugs for the treatment of ulcerative colitis. *Drugs* 14, 505–521. doi:10.1517/14728210903146882
- Peng, L., Gao, X., Nie, L., Xie, J., Dai, T., Shi, C., et al. (2020). Astragaloside attenuates dextran sulfate sodium (DSS)-induced acute experimental colitis by alleviating gut microbiota dysbiosis and inhibiting NF- κ B activation in mice. *Front. Immunol.* 11, 2058. doi:10.3389/fimmu.2020.02058
- Prados, M. E., Garcia-Martin, A., Unciti-Broceta, J. D., Palomares, B., Collado, J. A., Minassi, A., et al. (2021). Betulinic acid hydroxamate prevents colonic inflammation and fibrosis in murine models of inflammatory bowel disease. *Acta Pharmacol. Sin.* 42, 1124–1138. doi:10.1038/s41401-020-0497-0
- Pu, Z., Liu, Y., Li, C., Xu, M., Xie, H., and Zhao, J. (2020). Using network pharmacology for systematic understanding of geniposide in ameliorating inflammatory responses in colitis through suppression of NLRP3 inflammasome in macrophage by AMPK/Sirt1 dependent signaling. *Am. J. Chin. Med.* 48, 1693–1713. doi:10.1142/s0192415x20500846
- Puppala, E. R., Aochenlar, S. L., Shantanu, P. A., Ahmed, S., Jannu, A. K., Jala, A., et al. (2022). Perillyl alcohol attenuates chronic restraint stress aggravated dextran sulfate sodium-induced ulcerative colitis by modulating TLR4/NF- κ B and JAK2/STAT3 signaling pathways. *Phytomedicine* 106, 154415. doi:10.1016/j.phymed.2022.154415
- Qian, B., Wang, C., Zeng, Z., Ren, Y., Li, D., and Song, J.-L. (2020). Ameliorative effect of sinapic acid on dextran sodium sulfate- (DSS-) induced ulcerative colitis in kunming (KM) mice. *Oxid. Med. Cell. Longev.* 2020, 8393504. doi:10.1155/2020/8393504
- Qu, B., Cao, T., Wang, M., Wang, S., Li, W., Li, H., et al. (2022b). Ginsenosides Rd monomer inhibits proinflammatory cytokines production and alleviates DSS-colitis by NF- κ B and P38MAPK pathways in mice. *Immunopharmacol. Immunotoxicol.* 44, 110–118. doi:10.1080/08923973.2021.2012482
- Qu, C., Yuan, Z. W., Yu, X. T., Huang, Y. F., Yang, G. H., Chen, J. N., et al. (2017). Patchouli alcohol ameliorates dextran sodium sulfate-induced experimental colitis and suppresses tryptophan catabolism. *Pharmacol. Res.* 121, 70–82. doi:10.1016/j.phrs.2017.04.017
- Qu, L., Lin, X., Liu, C., Ke, C., Zhou, Z., Xu, K., et al. (2021c). Atractylodin attenuates dextran sulfate sodium-induced colitis by alleviating gut microbiota dysbiosis and inhibiting inflammatory response through the MAPK pathway. *Front. Pharmacol.* 12, 665376. doi:10.3389/fphar.2021.665376
- Qu, L., Shi, K., Xu, J., Liu, C., Ke, C., Zhan, X., et al. (2022a). Atractylenolide-1 targets SPHK1 and B4GALT2 to regulate intestinal metabolism and flora composition to improve inflammation in mice with colitis. *Phytomedicine* 98, 153945. doi:10.1016/j.phymed.2022.153945
- Qu, S., Chen, L., Wen, X., Zuo, J., Wang, X., Lu, Z., et al. (2021b). Suppression of Th17 cell differentiation via sphingosine-1-phosphate receptor 2 by cinnamaldehyde can ameliorate ulcerative colitis. *Biomed. Pharmacother.* 134, 111116. doi:10.1016/j.biopha.2020.111116
- Qu, Y., Li, X., Xu, F., Zhao, S., Wu, X., Wang, Y., et al. (2021a). Kaempferol alleviates murine experimental colitis by restoring gut microbiota and inhibiting the LPS-TLR4-NF- κ B Axis. *Front. Immunol.* 12, 679897. doi:10.3389/fimmu.2021.679897
- Rapa, S. F., Di Paola, R., Cordaro, M., Siracusa, R., D'Amico, R., Fusco, R., et al. (2021). Plumericin protects against experimental inflammatory bowel disease by restoring intestinal barrier function and reducing apoptosis. *Biomedicines* 9, 67. doi:10.3390/biomedicines9010067
- Rashidian, A., Dejbani, P., Karami Fard, K., Abdollahi, A., Chamanara, M., Dehpour, A., et al. (2020). Bupropion ameliorates acetic acid-induced colitis in rat: the involvement of the TLR4/NF- κ B signaling pathway. *Inflammation* 43, 1999–2009. doi:10.1007/s10753-020-01273-2
- Rashidian, A., Muhammadnejad, A., Dehpour, A. R., Mehr, S. E., Akhavan, M. M., Shirkoobi, R., et al. (2016). Atorvastatin attenuates TNBS-induced rat colitis: the involvement of the TLR4/NF- κ B signaling pathway. *Inflammopharmacology* 24, 109–118. doi:10.1007/s10787-016-0263-6
- Ren, J., Yue, B., Wang, H., Zhang, B., Luo, X., Yu, Z., et al. (2021). Acacetin ameliorates experimental colitis in mice via inhibiting macrophage inflammatory response and regulating the composition of gut microbiota. *Front. Physiol.* 11, 577237. doi:10.3389/fphys.2020.577237
- Rogler, G. (2010). Gastrointestinal and liver adverse effects of drugs used for treating IBD. *Best. Pract. Res. Clin. Gastroenterol.* 24, 157–165. doi:10.1016/j.bpg.2009.10.011
- Saha, S., Shalova, I. N., and Biswas, S. K. (2017). Metabolic regulation of macrophage phenotype and function. *Immunol. Rev.* 280, 102–111. doi:10.1111/immr.12603
- Sahu, B. D., Kumar, J. M., and Sistla, R. (2016). Fisetin, a dietary flavonoid, ameliorates experimental colitis in mice: relevance of NF- κ B signaling. *J. Nutr. Biochem.* 28, 171–182. doi:10.1016/j.jnutbio.2015.10.004
- Sakthivel, K. M., and Guruvayoorappan, C. (2013). Amentoflavone inhibits iNOS, COX-2 expression and modulates cytokine profile, NF- κ B signal transduction pathways in rats with ulcerative colitis. *Int. Immunopharmacol.* 17, 907–916. doi:10.1016/j.intimp.2013.09.022
- Sanchez-Fidalgo, S., Villegas, I., Angeles Rosillo, M., Aparicio-Soto, M., and Alarcon-de-la-Lastra, C. (2015). Dietary squalene supplementation improves DSS-induced acute colitis by downregulating p38 MAPK and NF κ B signaling pathways. *Mol. Nutr. Food Res.* 59, 284–292. doi:10.1002/mnfr.201400518
- Sangaraju, R., Nalban, N., Alavala, S., Rajendran, V., Jerald, M. K., and Sistla, R. (2019). Protective effect of galangin against dextran sulfate sodium (DSS)-induced ulcerative colitis in Balb/c mice. *Inflamm. Res.* 68, 691–704. doi:10.1007/s00011-019-01252-w
- Satoh, Y., Ishiguro, Y., Sakuraba, H., Kawaguchi, S., Hiraga, H., Fukuda, S., et al. (2009). Cyclosporin regulates intestinal epithelial apoptosis via TGF- β -related signaling. *Am. J. Physiol. Gastrointest. Liver Physiol.* 297, G514–G519. doi:10.1152/ajpgi.90608.2008
- Seo, G. S., Jiang, W.-Y., Park, P.-H., Sohn, D. H., Cheon, J. H., and Lee, S. H. (2014). Hirsutenone reduces deterioration of tight junction proteins through EGFR/Akt and ERK1/2 pathway both converging to HO-1 induction. *Biochem. Pharmacol.* 90, 115–125. doi:10.1016/j.bcp.2014.05.006
- Sethuraman, N., Swaminathan, S., Nelson, S. B., Palaninathan, P. S., Gopalan, T. K., and Velayudham, P. (2015). Modulation of PPAR γ and TNF α by emu oil and glycyrrhizin in ulcerative colitis. *Inflammopharmacology* 23, 47–56. doi:10.1007/s10787-014-0226-8
- Shafik, N. M., Gaber, R. A., Mohamed, D. A., and Ebeid, A. M. (2019). Hesperidin modulates dextran sulfate sodium-induced ulcerative colitis in rats: targeting sphingosine kinase-1-sphingosine 1-phosphate signaling pathway, mitochondrial biogenesis, inflammation, and apoptosis. *J. Biochem. Mol. Toxicol.* 33, e22312. doi:10.1002/jbt.22312
- Shah, Y. M., Morimura, K., and Gonzalez, F. J. (2007). Expression of peroxisome proliferator-activated receptor- γ in macrophage suppresses experimentally induced colitis. *Am. J. Physiol. Gastrointest. Liver Physiol.* 292, G657–G666. doi:10.1152/ajpgi.00381.2006
- Shahid, M., Raish, M., Ahmad, A., Bin Jardan, Y. A., Ansari, M. A., Ahad, A., et al. (2022). Sinapic acid ameliorates acetic acid-induced ulcerative colitis in rats by suppressing inflammation, oxidative stress, and apoptosis. *Molecules* 27, 4139. doi:10.3390/molecules27134139
- Sheikh, S. Z., Hegazi, R. A., Kobayashi, T., Onyiah, J. C., Russo, S. M., Matsuoka, K., et al. (2011). An anti-inflammatory role for carbon monoxide and heme oxygenase-1 in chronic Th2-mediated murine colitis. *J. Immunol.* 186, 5506–5513. doi:10.4049/jimmunol.1002433
- Shen, J., Cheng, J., Zhu, S., Zhao, J., Ye, Q., Xu, Y., et al. (2019b). Regulating effect of baicalin on IKK/I κ B/NF- κ B signaling pathway and apoptosis-related proteins in rats with ulcerative colitis. *Int. Immunopharmacol.* 73, 193–200. doi:10.1016/j.intimp.2019.04.052
- Shen, J., Li, N., and Zhang, X. (2019c). Daidzein ameliorates dextran sulfate sodium-induced experimental colitis in mice by regulating NF- κ B signaling. *J. Environ. Pathol. Toxicol. Oncol.* 38, 29–39. doi:10.1615/EnvironPatholToxicolOncol.2018027531
- Shen, P., Zhang, Z., Zhu, K., Cao, H., Liu, J., Lu, X., et al. (2019a). Evodiamine prevents dextran sulfate sodium-induced murine experimental colitis via the regulation of NF- κ B and NLRP3 inflammasome. *Biomed. Pharmacother.* 110, 786–795. doi:10.1016/j.biopha.2018.12.033
- Shen, Y., Liu, Y., Wang, Z., Ruan, X., Li, S., Ni, S., et al. (2020). Effect of berberine from *Coptis chinensis* on apoptosis of intestinal epithelial cells in a mouse model of ulcerative colitis: role of endoplasmic reticulum stress. *Evid. Based Complement. Altern. Med.* 2020, 3784671. doi:10.1155/2020/3784671
- Sheng, Q., Li, F., Chen, G., Li, J., Li, J., Wang, Y., et al. (2021). Ursolic acid regulates intestinal microbiota and inflammatory cell infiltration to prevent ulcerative colitis. *J. Immunol. Res.* 2021, 6679316. doi:10.1155/2021/6679316
- Shi, B., Liu, S., Huang, A., Zhou, M., Sun, B., Cao, H., et al. (2021). Revealing the mechanism of friedelin in the treatment of ulcerative colitis based on network pharmacology and experimental verification. *Evid. Based Complement. Altern. Med.* 2021, 4451779. doi:10.1155/2021/4451779
- Socca, E. A., Luiz-Ferreira, A., de Faria, F. M., de Almeida, A. C., Dunder, R. J., Manzo, L. P., et al. (2014). Inhibition of tumor necrosis factor- α and cyclooxygenase-2 by Isatin: a molecular mechanism of protection against TNBS-induced colitis in rats. *Chem. Biol. Interact.* 209, 48–55. doi:10.1016/j.cbi.2013.11.019
- Song, C., Chen, J., Li, X., Yang, R., Cao, X., Zhou, L., et al. (2021). Limonin ameliorates dextran sulfate sodium-induced chronic colitis in mice by inhibiting PERK-ATF4-CHOP pathway of ER stress and NF- κ B signaling. *Int. Immunopharmacol.* 90, 107161. doi:10.1016/j.intimp.2020.107161
- Su, C. G., Wen, X., Bailey, S. T., Jiang, W., Rangwala, S. M., Keilbaugh, S. A., et al. (1999). A novel therapy for colitis utilizing PPAR- γ ligands to inhibit the epithelial inflammatory response. *J. Clin. Invest.* 104, 383–389. doi:10.1172/JCI7145
- Su, S., Wang, X., Xi, X., Zhu, L., Chen, Q., Zhang, H., et al. (2021). Phellodendrine promotes autophagy by regulating the AMPK/mTOR pathway and treats ulcerative colitis. *J. Cell. Mol. Med.* 25, 5707–5720. doi:10.1111/jcmm.16587
- Suzuki, T. (2013). Regulation of intestinal epithelial permeability by tight junctions. *Cell. Mol. Life Sci.* 70, 631–659. doi:10.1007/s00018-012-1070-x
- Tan, X., Wen, Y., Han, Z., Su, X., Peng, J., Chen, F., et al. (2023). Cinnamaldehyde ameliorates dextran sulfate sodium-induced colitis in mice by modulating TLR4/NF-

- kappa B signaling pathway and NLRP3 inflammasome activation. *Chem. Biodivers.* 20, e202200089. doi:10.1002/cbdv.202200089
- Tan, Y., and Zheng, C. (2018). Effects of alpinetin on intestinal barrier function, inflammation and oxidative stress in dextran sulfate sodium-induced ulcerative colitis mice. *Am. J. Med. Sci.* 355, 377–386. doi:10.1016/j.amjms.2018.01.002
- Tan, Z., Xie, N., Cui, H., Moellering, D. R., Abraham, E., Thannickal, V. J., et al. (2015). Pyruvate dehydrogenase kinase 1 participates in macrophage polarization via regulating glucose metabolism. *J. Immunol.* 194, 6082–6089. doi:10.4049/jimmunol.1402469
- Tang, B., Zhu, J., Fang, S., Wang, Y., Vinothkumar, R., Li, M., et al. (2021). Pharmacological inhibition of MELK restricts ferroptosis and the inflammatory response in colitis and colitis-propelled carcinogenesis. *Free Radic. Biol. Med.* 172, 312–329. doi:10.1016/j.freeradbiomed.2021.06.012
- Tang, B., Zhu, J., Zhang, B., Wu, F., Wang, Y., Weng, Q., et al. (2020b). Therapeutic potential of triptolide as an anti-inflammatory agent in dextran sulfate sodium-induced murine experimental colitis. *Front. Immunol.* 11, 592084. doi:10.3389/fimmu.2020.592084
- Tang, X., Huang, G., Zhang, T., and Li, S. (2020a). Elucidation of colon-protective efficacy of diosgenin in experimental TNBS-induced colitis: inhibition of NF- κ B/I κ B- α and Bax/Caspase-1 signaling pathways. *Biosci. Biotechnol. Biochem.* 84, 1903–1912. doi:10.1080/09168451.2020.1776590
- Tekeli, I. O., Atessahin, A., Sakin, F., Aslan, A., Ceribasi, S., and Yipel, M. (2018). Protective effects of conventional and colon-targeted lycopene and linalool on ulcerative colitis induced by acetic acid in rats. *Inflammopharmacology* 27, 313–322. doi:10.1007/s10787-018-0485-x
- Tian, M., Ma, P., Zhang, Y., Mi, Y., and Fan, D. (2020). Ginsenoside Rk3 alleviated DSS-induced ulcerative colitis by protecting colitis barrier and inhibiting NLRP3 inflammasome pathway. *Int. Immunopharmacol.* 85, 106645. doi:10.1016/j.intimp.2020.106645
- Tiana, X., Peng, Z., Luo, S., Zhang, S., Li, B., Zhou, C., et al. (2019). Aesculin protects against DSS-Induced colitis through activating PPAR γ and inhibiting NF- κ B pathway. *Eur. J. Pharmacol.* 857, 172453. doi:10.1016/j.ejphar.2019.172453
- Ungaro, R., Mehandru, S., Allen, P. B., Peyrin-Biroulet, L., and Colombel, J.-F. (2017). Ulcerative colitis. *Lancet* 389, 1756–1770. doi:10.1016/s0140-6736(16)32126-2
- Veltkamp, C., Tonkonogy, S. L., de Jong, Y. P., Albright, C., Grenther, W. B., Balish, E., et al. (2001). Continuous stimulation by normal luminal bacteria is essential for the development and perpetuation of colitis in Tg(epsilon26) mice. *Gastroenterology* 120, 900–913. doi:10.1053/gast.2001.22547
- Villani, A. C., Lemire, M., Fortin, G., Louis, E., Silverberg, M. S., Collette, C., et al. (2009). Common variants in the NLRP3 region contribute to Crohn's disease susceptibility. *Nat. Genet.* 41, 71–76. doi:10.1038/ng.285
- Vukelic, I., Detel, D., Baticic, L., Potocnjak, I., and Domitrovic, R. (2020). Luteolin ameliorates experimental colitis in mice through ERK-mediated suppression of inflammation, apoptosis and autophagy. *Food Chem. Toxicol.* 145, 111680. doi:10.1016/j.fct.2020.111680
- Wan, P., Chen, H., Guo, Y., and Bai, A. P. (2014). Advances in treatment of ulcerative colitis with herbs: from bench to bedside. *World J. Gastroenterol.* 20, 14099–14104. doi:10.3748/wjg.v20.i39.14099
- Wang, F., Zhang, S., Jeon, R., Vuckovic, I., Jiang, X., Lerman, A., et al. (2018b). Interferon gamma induces reversible metabolic reprogramming of M1 macrophages to sustain cell viability and pro-inflammatory activity. *EBioMedicine* 30, 303–316. doi:10.1016/j.ebiom.2018.02.009
- Wang, H., Zhang, M., Wen, X., He, L., Zhang, M., Zhang, J., et al. (2022b). Cepharranthine ameliorates dextran sulphate sodium-induced colitis through modulating gut microbiota. *Microb. Biotechnol.* 15, 2208–2222. doi:10.1111/1751-7915.14059
- Wang, H.-Y., Ge, W., Liu, S.-Q., Long, J., Jiang, Q.-Q., Zhou, W., et al. (2022d). Curcumin inhibits T follicular helper cell differentiation in mice with dextran sulfate sodium (DSS)-Induced colitis. *Am. J. Chin. Med.* 50, 275–293. doi:10.1142/s0192415x22500100
- Wang, K., Li, Y., Lv, Q., Li, X., Dai, Y., and Wei, Z. (2018a). Bergenin, acting as an agonist of PPAR gamma, ameliorates experimental colitis in mice through improving expression of SIRT1, and therefore inhibiting NF-kappa B-mediated macrophage activation. *Front. Pharmacol.* 8, 981. doi:10.3389/fphar.2017.00981
- Wang, L., Shao, L., Chen, M.-Y., Wang, L., Zhang, W., Tan, F.-B., et al. (2022c). Effect of ginsenoside compound K on alleviating colitis via modulating gut microbiota. *Chin. Med.* 17, 146. doi:10.1186/s13020-022-00701-9
- Wang, L., and Wang, J. (2022). Honokiol ameliorates DSS-induced mouse colitis by inhibiting inflammation and oxidative stress and improving the intestinal barrier. *Oxid. Med. Cell. Longev.* 2022, 1755608. doi:10.1155/2022/1755608
- Wang, M., Li, Y., Su, J., Bai, J., Zhao, Z., and Sun, Z. (2023c). Protective effects of 4-geranyloxy-2,6-dihydroxybenzophenone on DSS-induced ulcerative colitis in mice via regulation of cAMP/PKA/CREB and NF-kappa B signaling pathways. *Phytother. Res.* 37, 1330–1345. doi:10.1002/ptr.7689
- Wang, M., Lin, L., Chen, Y., Zhong, Y., Lin, Y., Li, P., et al. (2020a). Evodiamine has therapeutic efficacy in ulcerative colitis by increasing Lactobacillus acidophilus levels and acetate production. *Pharmacol. Res.* 159, 104978. doi:10.1016/j.phrs.2020.104978
- Wang, N., Kong, R., Han, W., Bao, W., Shi, Y., Ye, L., et al. (2022e). Honokiol alleviates ulcerative colitis by targeting PPAR-gamma-TLR4-NF-kappa B signaling and suppressing gasdermin-D-mediated pyroptosis *in vivo* and *in vitro*. *Int. Immunopharmacol.* 111, 109058. doi:10.1016/j.intimp.2022.109058
- Wang, S., Liu, W., Wang, J., and Bai, X. (2020b). Curculigoside inhibits ferroptosis in ulcerative colitis through the induction of GPX4. *Life Sci.* 259, 118356. doi:10.1016/j.lfs.2020.118356
- Wang, X., Su, L., Tan, J., Ding, T., and Yue, Y. (2023b). Albilflorin alleviates DSS-induced ulcerative colitis in mice by reducing inflammation and oxidative stress. *Iran. J. Basic Med. Sci.* 26, 48–56. doi:10.22038/ijbms.2022.66678.14624
- Wang, Y., Liu, J., Huang, Z., Li, Y., Liang, Y., Luo, C., et al. (2021a). Coptisine ameliorates DSS-induced ulcerative colitis via improving intestinal barrier dysfunction and suppressing inflammatory response. *Eur. J. Pharmacol.* 896, 173912. doi:10.1016/j.ejphar.2021.173912
- Wang, Y., Liu, K., Qi, Z., Chen, T., Yu, W., Jiang, Y., et al. (2021b). Therapeutic mechanism and effect of camptothecin on dextran sodium sulfate-induced ulcerative colitis in mice. *J. Immunol. Res.* 2021, 5556659. doi:10.1155/2021/5556659
- Wang, Y. J., Li, Q. M., Zha, X. Q., and Luo, J. P. (2022a). Intervention and potential mechanism of non-starch polysaccharides from natural resources on ulcerative colitis: a review. *Int. J. Biol. Macromol.* 210, 545–564. doi:10.1016/j.ijbiomac.2022.04.208
- Wang, Z.-J., Chen, L.-H., Xu, J., Xu, Q.-X., Xu, W., and Yang, X.-W. (2023a). Corylin ameliorates chronic ulcerative colitis via regulating the gut-brain axis and promoting 5-hydroxytryptophan production in the colon. *Phytomedicine* 110, 154651. doi:10.1016/j.phymed.2023.154651
- Wei, H. L., Li, J. T., Chen, Z. G., and Yan, S. G. (2022). Experimental study on effects of berberine combined with 6-shogaol on intestinal inflammation and flora in mice with ulcerative colitis. *China J. Chin. Mat. Med.* 47, 4418–4427. doi:10.19540/j.cnki.cjcm.20220413.401
- Wei, W., Ding, M., Zhou, K., Xie, H., Zhang, M., and Zhang, C. (2017). Protective effects of wedelolactone on dextran sodium sulfate induced murine colitis partly through inhibiting the NLRP3 inflammasome activation via AMPK signaling. *Biomed. Pharmacother.* 94, 27–36. doi:10.1016/j.biopha.2017.06.071
- Whyte, J. L., Smith, A. A., and Helms, J. A. (2012). Wnt signaling and injury repair. *Cold Spring Harb. Perspect. Biol.* 4, a008078. doi:10.1101/cshperspect.a008078
- Wu, H., Rao, Q., Ma, G. C., Yu, X. H., Zhang, C. E., and Ma, Z. J. (2020a). Effect of triptolide on dextran sodium sulfate-induced ulcerative colitis and gut microbiota in mice. *Front. Pharmacol.* 11, 1652. doi:10.3389/fphar.2019.01652
- Wu, K., Liu, X., Meng, X., Cao, L., Li, H., Bi, Y., et al. (2023). Sauchinone alleviates dextran sulfate sodium-induced ulcerative colitis via NAD(P)H dehydrogenase quinone 1/NF-kB pathway and gut microbiota. *Front. Microbiol.* 13, 1084257. doi:10.3389/fmicb.2022.1084257
- Wu, M., Li, P., An, Y., Ren, J., Yan, D., Cui, J., et al. (2019). Phloretin ameliorates dextran sulfate sodium-induced ulcerative colitis in mice by regulating the gut microbiota. *Pharmacol. Res.* 150, 104489. doi:10.1016/j.phrs.2019.104489
- Wu, M.-M., Wang, Q.-M., Huang, B.-Y., Mai, C.-T., Wang, C.-L., Wang, T.-T., et al. (2021). Dioscin ameliorates murine ulcerative colitis by regulating macrophage polarization. *Pharmacol. Res.* 172, 105796. doi:10.1016/j.phrs.2021.105796
- Wu, S., and Chen, Z. (2019). Astragaloside IV alleviates the symptoms of experimental ulcerative colitis *in vitro* and *in vivo*. *Exp. Ther. Med.* 18, 2877–2884. doi:10.3892/etm.2019.7907
- Wu, X., Yang, Y., Dou, Y., Ye, J., Bian, D., Wei, Z., et al. (2014). Arctigenin but not arctiin acts as the major effective constituent of *Arctium lappa* L. fruit for attenuating colonic inflammatory response induced by dextran sulfate sodium in mice. *Int. Immunopharmacol.* 23, 505–515. doi:10.1016/j.intimp.2014.09.026
- Wu, Y., Li, Y., Ruan, Z., Li, J., Zhang, L., Lu, H., et al. (2020b). Puerarin rebuilding the mucus layer and regulating mucin-utilizing bacteria to relieve ulcerative colitis. *J. Agric. Food Chem.* 68, 11402–11411. doi:10.1021/acs.jafc.0c04119
- Wu, Y. T., Zhong, L. S., Huang, C., Guo, Y. Y., Jin, F. J., Hu, Y. Z., et al. (2022). β -Caryophyllene acts as a ferroptosis inhibitor to ameliorate experimental colitis. *Int. J. Mol. Sci.* 23, 16055. doi:10.3390/ijms232416055
- Wu, Z., Zeng, H., Zhang, L., Pu, Y., Li, S., Yuan, Y., et al. (2020c). Patchouli alcohol: a natural sesquiterpene against both inflammation and intestinal barrier damage of ulcerative colitis. *Inflammation* 43, 1423–1435. doi:10.1007/s10753-020-01219-8
- Xie, J., Li, L., Deng, S., Chen, J., Gu, Q., Su, H., et al. (2020). Slit2/Robo1 mitigates DSS-induced ulcerative colitis by activating autophagy in intestinal stem cell. *Int. J. Biol. Sci.* 16, 1876–1887. doi:10.7150/ijbs.42331
- Xie, X., Zhao, M., Huang, S., Li, P., Chen, P., Luo, X., et al. (2022). Luteolin alleviates ulcerative colitis by restoring the balance of NCR(-)ILC3/NCR(+)ILC3 to repairing impaired intestinal barrier. *Int. Immunopharmacol.* 112, 109251. doi:10.1016/j.intimp.2022.109251
- Xing, J. F., Sun, J. N., Sun, J. Y., You, C. Y., Dong, K., Lv, J., et al. (2012). Protective effects of 3,4-oxo-isopropylidene-shikimic acid on experimental colitis induced by trinitrobenzenesulfonic acid in rats. *Dig. Dis. Sci.* 57, 2045–2054. doi:10.1007/s10620-012-2155-y

- Xu, B., Huang, S., Chen, Y., Wang, Q., Luo, S., Li, Y., et al. (2021). Synergistic effect of combined treatment with baicalin and emodin on DSS-induced colitis in mouse. *Phytother. Res.* 35, 5708–5719. doi:10.1002/ptr.7230
- Xu, B., Li, Y.-L., Xu, M., Yu, C.-C., Lian, M.-Q., Tang, Z.-Y., et al. (2017). Geniposide ameliorates TNBS-induced experimental colitis in rats via reducing inflammatory cytokine release and restoring impaired intestinal barrier function. *Acta Pharmacol. Sin.* 38, 688–698. doi:10.1038/aps.2016.168
- Xu, D., Zhuang, L., Gao, S., Ma, H., Cheng, J., Liu, J., et al. (2022a). Orally administered ginkgolide C attenuates DSS-induced colitis by maintaining gut barrier integrity, inhibiting inflammatory responses, and regulating intestinal flora. *J. Agric. Food Chem.* 70, 14718–14731. doi:10.1021/acs.jafc.2c06177
- Xu, X., Li, W., Yu, Z., Zhang, L., Duo, T., Zhao, Y., et al. (2022c). Berberine ameliorates dextran sulfate sodium-induced ulcerative colitis and inhibits the secretion of gut lysozyme via promoting autophagy. *Metabolites* 12, 676. doi:10.3390/metabo12080676
- Xu, X., Zhang, G., Peng, K., Gao, Y., Wang, J., Gao, C., et al. (2022b). Carnosol maintains intestinal barrier function and mucosal immune homeostasis in DSS-induced colitis. *Front. Nutr.* 9, 894307. doi:10.3389/fnut.2022.894307
- Xu, Z., Wei, C., Zhang, R., Yao, J., Zhang, D., and Wang, L. (2015). Epigallocatechin-3-gallate-induced inhibition of interleukin-6 release and adjustment of the regulatory T/T helper 17 cell balance in the treatment of colitis in mice. *Exp. Ther. Med.* 10, 2231–2238. doi:10.3892/etm.2015.2824
- Xuan, H., Ou, A., Hao, S., Shi, J., and Jin, X. (2020). Galangin protects against symptoms of dextran sodium sulfate-induced acute colitis by activating autophagy and modulating the gut microbiota. *Nutrients* 12, 347. doi:10.3390/nu12020347
- Xue, H., Li, J., Li, S., Guo, J., Yan, R., Chen, T., et al. (2023). Phyllygenin attenuated colon inflammation and improved intestinal mucosal barrier in DSS-induced colitis mice via TLR4/src mediated MAPK and NF-kappa B signaling pathways. *Int. J. Mol. Sci.* 24, 2238. doi:10.3390/ijms24032238
- Yan, H. Y., Wang, H. G., Zhang, X. L., Li, X. Q., and Yu, J. (2015). Ascorbic acid ameliorates oxidative stress and inflammation in dextran sulfate sodium-induced ulcerative colitis in mice. *Int. J. Clin. Exp. Med.* 8, 20245–20253.
- Yan, S., Hui, Y., Li, J., Xu, X., Li, Q., and Wei, H. (2020b). Glutamine relieves oxidative stress through PI3K/Akt signaling pathway in DSS-induced ulcerative colitis mice. *Iran. J. Basic Med. Sci.* 23, 1124–1129. doi:10.22038/ijbms.2020.39815.9436
- Yan, X., Lu, Q. G., Zeng, L., Li, X. H., Liu, Y., Du, X. F., et al. (2020a). Synergistic protection of astragalus polysaccharides and matrine against ulcerative colitis and associated lung injury in rats. *World J. Gastroenterol.* 26, 55–69. doi:10.3748/wjg.v26.i1.55
- Yang, H., Yue, Y., Li, Y., Su, L., and Yan, S. (2020). Geniposide attenuates dextran sulfate sodium-induced colitis in mice via Nf-2/HO-1/NF-kB pathway. *Ann. Palliat. Med.* 9, 2826–2836. doi:10.21037/ajpm.20-279
- Yang, N., Xia, Z., Shao, N., Li, B., Xue, L., Peng, Y., et al. (2017). Carnosic acid prevents dextran sulfate sodium-induced acute colitis associated with the regulation of the Keap1/Nrf2 pathway. *Sci. Rep.* 7, 11036. doi:10.1038/s41598-017-11408-5
- Yang, Y. X., Yuan, Y., and Xia, B. (2022). Cinnamtannin D1 ameliorates DSS-induced colitis by preventing Th17/Treg imbalance through activation of the AMPK/mTOR pathway. *Allergol. Immunopathol. (Madr.)* 50, 153–161. doi:10.15586/aei.v50i5.654
- Yao, H., Shi, Y., Yuan, J., Sa, R., Chen, W., and Wan, X. (2021). Matrine protects against DSS-induced murine colitis by improving gut barrier integrity, inhibiting the PPAR- α signaling pathway, and modulating gut microbiota. *Int. Immunopharmacol.* 100, 108091. doi:10.1016/j.intimp.2021.108091
- Yao, H., Yan, J., Yin, L., and Chen, W. (2022). Picoside II alleviates DSS-induced ulcerative colitis by suppressing the production of NLRP3 inflammasomes through NF-kB signaling pathway. *Immunopharmacol. Immunotoxicol.* 44, 437–446. doi:10.1080/08923973.2022.2054425
- Yao, J., Cao, X., Zhang, R., Li, Y.-X., Xu, Z.-L., Zhang, D.-G., et al. (2016). Protective effect of baicalin against experimental colitis via suppression of oxidant stress and apoptosis. *Pharmacogn. Mag.* 12, 225–234. doi:10.4103/0973-1296.186342
- Yao, J., Wang, J. Y., Liu, L., Zeng, W. S., Li, Y. X., Xun, A. Y., et al. (2011). Polydatin ameliorates DSS-induced colitis in mice through inhibition of nuclear factor-kappaB activation. *Planta Med.* 77, 421–427. doi:10.1055/s-0030-1250462
- Yeom, J., Kim, S., and Park, S. (2022). Regulation of the gut microbiota and inflammation by β -caryophyllene extracted from cloves in a dextran sulfate sodium-induced colitis mouse model. *Molecules* 27, 7782. doi:10.3390/molecules27227782
- Yin, S., Li, L., Tao, Y., Yu, J., Wei, S., Liu, M., et al. (2021). The inhibitory effect of artesunate on excessive endoplasmic reticulum stress alleviates experimental colitis in mice. *Front. Pharmacol.* 12, 629798. doi:10.3389/fphar.2021.629798
- Yin, S., Yang, H., Tao, Y., Wei, S., Li, L., Liu, M., et al. (2020). Artesunate ameliorates DSS-induced ulcerative colitis by protecting intestinal barrier and inhibiting inflammatory response. *Inflammation* 43, 765–776. doi:10.1007/s10753-019-01164-1
- Yin, Y., Liu, K., and Li, G. (2022). Protective effect of prim-O-glucosylcimifugin on ulcerative colitis and its mechanism. *Front. Pharmacol.* 13, 882924. doi:10.3389/fphar.2022.882924
- Yin, Z., Wang, Q., and Cheng, H. (2023). Synergistic protective effect of interactions of quercetin with lycopene against ochratoxin A-induced ulcerative colitis. *Appl. Biochem. Biotechnol.* 195, 5253–5266. doi:10.1007/s12010-022-04287-8
- You, B. H., Chae, H. S., Song, J., Ko, H. W., Chin, Y. W., and Choi, Y. H. (2017). α -Mangostin ameliorates dextran sulfate sodium-induced colitis through inhibition of NF-kB and MAPK pathways. *Int. Immunopharmacol.* 49, 212–221. doi:10.1016/j.intimp.2017.05.040
- Yu, S., Qian, H., Zhang, D., and Jiang, Z. (2023a). Ferulic acid relieved ulcerative colitis by inhibiting the TXNIP/NLRP3 pathway in rats. *Cell Biol. Int.* 47, 417–427. doi:10.1002/cbin.11935
- Yu, S., and Qian, H. H. (2021). Deoxyschizandrin treats mice with ulcerative colitis possibly via the TLR4/NF-kB signaling pathway. *Am. J. Transl. Res.* 13, 3856–3863.
- Yu, T.-Y., Feng, Y.-M., Kong, W.-S., Li, S.-N., Sun, X.-J., Zhou, G., et al. (2023b). Gallic acid ameliorates dextran sodium-induced ulcerative colitis in mice via inhibiting NLRP3 inflammasome. *Front. Pharmacol.* 14, 1095721. doi:10.3389/fphar.2023.1095721
- Yu, X. T., Xu, Y. F., Huang, Y. F., Qu, C., Xu, L. Q., Su, Z. R., et al. (2018). Berberine attenuates mucosal lesions and inflammation in dextran sodium sulfate-induced colitis in mice. *PLoS ONE* 13, e0194069. doi:10.1371/journal.pone.0194069
- Yuan, J., Cheng, W., Zhang, G., Ma, Q., Li, X., Zhang, B., et al. (2020). Protective effects of iridoid glycosides on acute colitis via inhibition of the inflammatory response mediated by the STAT3/NF-kB pathway. *Int. Immunopharmacol.* 81, 106240. doi:10.1016/j.intimp.2020.106240
- Yuan, S.-N., Wang, M.-X., Han, J.-L., Feng, C.-Y., Wang, M., Wang, M., et al. (2023). Improved colonic inflammation by nervonic acid via inhibition of NF-kB signaling pathway of DSS-induced colitis mice. *Phytomedicine* 112, 154702. doi:10.1016/j.phymed.2023.154702
- Yue, B., Ren, J. Y., Yu, Z. L., Luo, X. P., Ren, Y. J., Zhang, J., et al. (2020). Pinocembrin alleviates ulcerative colitis in mice via regulating gut microbiota, suppressing TLR4/MD2/NF-kappa B pathway and promoting intestinal barrier. *Biosci. Rep.* 40, 1010. doi:10.1042/bsr20200986
- Zeng, J., Zhang, D., Wan, X., Bai, Y., Yuan, C., Wang, T., et al. (2020). Chlorogenic acid suppresses miR-155 and ameliorates ulcerative colitis through the NF-kappa B/NLRP3 inflammasome pathway. *Mol. Nutr. Food Res.* 64, e2000452. doi:10.1002/mnfr.202000452
- Zhang, C., Zhu, H., Jie, H., Ding, H., and Sun, H. (2021d). Arbutin ameliorated ulcerative colitis of mice induced by dextran sodium sulfate (DSS). *Bioengineered* 12, 11707–11715. doi:10.1080/21655979.2021.2005746
- Zhang, H., Deng, A., Zhang, Z., Yu, Z., Liu, Y., Peng, S., et al. (2016c). The protective effect of epicatechin on experimental ulcerative colitis in mice is mediated by increasing antioxidation and by the inhibition of NF-kB pathway. *Pharmacol. Rep.* 68, 514–520. doi:10.1016/j.pharep.2015.12.011
- Zhang, H., Lang, W., Li, S., Xu, C., Wang, X., Li, Y., et al. (2023a). Corynoline ameliorates dextran sulfate sodium-induced colitis in mice by modulating Nrf2/NF-kB pathway. *Immunopharmacol. Immunotoxicol.* 45, 26–34. doi:10.1080/08923973.2022.2112218
- Zhang, H., Lang, W., Liu, X., Bai, J., Jia, Q., and Shi, Q. (2022e). Procyandin A1 alleviates DSS-induced ulcerative colitis via regulating AMPK/mTOR/p70S6K-mediated autophagy. *J. Physiol. Biochem.* 78, 213–227. doi:10.1007/s13105-021-00854-5
- Zhang, H., Wang, Y., Su, Y., Fang, X., and Guo, W. (2021c). The alleviating effect and mechanism of Bilobalide on ulcerative colitis. *Food. Funct.* 12, 6226–6239. doi:10.1039/d1fo01266e
- Zhang, H., Zhuo, S., Song, D., Wang, L., Gu, J., Ma, J., et al. (2021a). Icarin inhibits intestinal inflammation of DSS-induced colitis mice through modulating intestinal flora abundance and modulating p-p65/p65 molecule. *J. Gastroenterol.* 32, 382–392. doi:10.5152/tjg.2021.20282
- Zhang, J., Cao, L., Sun, Y., Qing, D.-G., Xu, X.-Q., Wang, J.-C., et al. (2021b). The regulatory effects of licochalcone A on the intestinal epithelium and gut microbiota in murine colitis. *Molecules* 26, 4149. doi:10.3390/molecules26144149
- Zhang, J., Liang, F., Chen, Z., Chen, Y., Yuan, J., Xiong, Q., et al. (2022c). Vitexin protects against dextran sodium sulfate-induced colitis in mice and its potential mechanisms. *J. Agric. Food Chem.* 70, 12041–12054. doi:10.1021/acs.jafc.2c05177
- Zhang, J., Xu, X., Li, N., Cao, L., Sun, Y., Wang, J., et al. (2022b). Licoflavone B, an isoprene flavonoid derived from licorice residue, relieves dextran sodium sulfate-induced ulcerative colitis by rebuilding the gut barrier and regulating intestinal microflora. *Eur. J. Pharmacol.* 916, 174730. doi:10.1016/j.ejphar.2021.174730
- Zhang, J. L., Zhang, M. N., Wang, H. G., Yang, X. Z., and Yu, C. G. (2022a). Jatrochazine alleviates ulcerative colitis via regulating gut microbiota and NOS2 expression. *Gut Pathog.* 14, 41. doi:10.1186/s13099-022-00514-z
- Zhang, L., Cao, N., Wang, Y., Wang, Y., Wu, C., Cheng, X., et al. (2020b). Improvement of oxazolone-induced ulcerative colitis in rats using andrographolide. *Molecules* 25, 76. doi:10.3390/molecules25010076
- Zhang, L., Wang, Y., Tong, L., Sun, S., Liu, W., Zang, S., et al. (2017b). Berberine alleviates dextran sodium sulfate-induced colitis by improving intestinal barrier function and reducing inflammation and oxidative stress. *Exp. Ther. Med.* 13, 3374–3382. doi:10.3892/etm.2017.4402

- Zhang, L., Xue, H., Zhao, G., Qiao, C., Sun, X., Pang, C., et al. (2019b). Curcumin and resveratrol suppress dextran sulfate sodium-induced colitis in mice. *Mol. Med. Rep.* 19, 3053–3060. doi:10.3892/mmr.2019.9974
- Zhang, M., Long, Y., Sun, Y., Wang, Y., Li, Q., Wu, H., et al. (2011). Evidence for the complementary and synergistic effects of the three-alkaloid combination regimen containing berberine, hyaconitine and skimmianine on the ulcerative colitis rats induced by trinitrobenzene-sulfonic acid. *Eur. J. Pharmacol.* 651, 187–196. doi:10.1016/j.ejphar.2010.10.030
- Zhang, Q., Wen, F., Sun, F., Xu, Z., Liu, Y., Tao, C., et al. (2023b). Efficacy and mechanism of quercetin in the treatment of experimental colitis using network pharmacology analysis. *Molecules* 28, 146. doi:10.3390/molecules28010146
- Zhang, Q., Xu, N., Hu, X., and Zheng, Y. (2020c). Anti-colitic effects of Physalin B on dextran sodium sulfate-induced BALB/c mice by suppressing multiple inflammatory signaling pathways. *J. Ethnopharmacol.* 259, 112956. doi:10.1016/j.jep.2020.112956
- Zhang, W., Wang, W., Shen, C., Wang, X., Pu, Z., and Yin, Q. (2021f). Network pharmacology for systematic understanding of Schisandrin B reduces the epithelial cells injury of colitis through regulating pyroptosis by AMPK/Nrf2/NLRP3 inflammasome. *Aging-Us* 13, 23193–23209. doi:10.18632/aging.203611
- Zhang, W. F., Yang, Y., Su, X., Xu, D. Y., Yan, Y. L., Gao, Q., et al. (2016b). Deoxychizandrin suppresses dss-induced ulcerative colitis in mice. *Saudi. J. Gastroenterol.* 22, 448–455. doi:10.4103/1319-3767.195552
- Zhang, X.-J., Yuan, Z.-W., Qu, C., Yu, X.-T., Huang, T., Chen, P. V., et al. (2018). Palmatine ameliorated murine colitis by suppressing tryptophan metabolism and regulating gut microbiota. *Pharmacol. Res.* 137, 34–46. doi:10.1016/j.phrs.2018.09.010
- Zhang, Y., Tan, L., Li, C., Wu, H., Ran, D., and Zhang, Z. (2020a). Sulforaphane alter the microbiota and mitigate colitis severity on mice ulcerative colitis induced by DSS. *Amb. Express* 10, 119. doi:10.1186/s13568-020-01053-z
- Zhang, Y., Zha, Z., Shen, W., Li, D., Kang, N., Chen, Z., et al. (2021e). Anemoside B4 ameliorates TNBS-induced colitis through S100A9/MAPK/NF- κ B signaling pathway. *Chin. Med.* 16, 11. doi:10.1186/s13020-020-00410-1
- Zhang, Z., Cui, Y., Liu, S., Huang, J., Liu, Y., Zhou, Y., et al. (2022d). Short-term treatment with zingerone ameliorates dextran sulfate sodium-induced mouse experimental colitis. *J. Sci. Food Agric.* 102, 4873–4882. doi:10.1002/jsfa.11850
- Zhang, Z., Li, S., Cao, H., Shen, P., Liu, J., Fu, Y., et al. (2019a). The protective role of phloretin against dextran sulfate sodium-induced ulcerative colitis in mice. *Food. Funct.* 10, 422–431. doi:10.1039/c8fo01699b
- Zhang, Z., Li, Y., Shen, P., Li, S., Lu, X., Liu, J., et al. (2017d). Administration of geniposide ameliorates dextran sulfate sodium-induced colitis in mice via inhibition of inflammation and mucosal damage. *Int. Immunopharmacol.* 49, 168–177. doi:10.1016/j.intimp.2017.05.033
- Zhang, Z., Shen, P., Lu, X., Li, Y., Liu, J., Liu, B., et al. (2017a). *In vivo* and *in vitro* study on the efficacy of Terpinen-4-ol in Dextran sulfate sodium-induced Mice experimental colitis. *Front. Immunol.* 8, 558. doi:10.3389/fimmu.2017.00558
- Zhang, Z., Wu, X., Cao, S., Wang, L., Wang, D., Yang, H., et al. (2016a). Caffeic acid ameliorates colitis in association with increased Akkermansia population in the gut microbiota of mice. *Oncotarget* 7, 31790–31799. doi:10.18632/oncotarget.9306
- Zhang, Z., Yang, L., Wang, B., Zhang, L., Zhang, Q., Li, D., et al. (2017c). Protective role of liriiodendrin in mice with dextran sulphate sodium-induced ulcerative colitis. *Int. Immunopharmacol.* 52, 203–210. doi:10.1016/j.intimp.2017.09.012
- Zhao, Y., Liu, P., Zhang, Y., Jiang, H., Luan, H., Xu, Y., et al. (2022b). Demethyleneberberine blocked the maturation of IL-1 β in inflammation by inhibiting TLR4-mitochondria signaling. *Int. Immunopharmacol.* 113, 109319. doi:10.1016/j.intimp.2022.109319
- Zhao, Y., Zhong, X., Yan, J., Sun, C., Zhao, X., and Wang, X. (2022a). Potential roles of gut microbes in biotransformation of natural products: an overview. *Front. Microbiol.* 13, 956378. doi:10.3389/fmicb.2022.956378
- Zhao, Z. J., Xiang, J. Y., Liu, L., Huang, X. L., and Gan, H. T. (2012). Parthenolide, an inhibitor of the nuclear factor- κ B pathway, ameliorates dextran sulfate sodium-induced colitis in mice. *Int. Immunopharmacol.* 12, 169–174. doi:10.1016/j.intimp.2011.11.007
- Zheng, C., Wang, Y., Xu, Y., Zhou, L., Hassan, S., Xu, G., et al. (2021a). Berberine inhibits dendritic cells differentiation in DSS-induced colitis by promoting Bacteroides fragilis. *Int. Immunopharmacol.* 101, 108329. doi:10.1016/j.intimp.2021.108329
- Zheng, J., Li, H., Zhang, P., Yue, S., Zhai, B., Zou, J., et al. (2022). Paeonol ameliorates ulcerative colitis in mice by modulating the gut microbiota and metabolites. *Metabolites* 12, 956. doi:10.3390/metabo12100956
- Zheng, J.-Y., Xu, J.-Y., Zhang, L., Wang, Z.-M., Yin, X.-B., and Qin, L.-Q. (2023). Effect of 3,3'-diselenodipropionic acid on dextran sodium sulfate-induced ulcerative colitis in mice. *Biol. Trace Elem. Res.* 201, 3961–3970. doi:10.1007/s12011-022-03491-1
- Zheng, K., Jia, J., Yan, S., Shen, H., Zhu, P., and Yu, J. (2020). Paeoniflorin ameliorates ulcerative colitis by modulating the dendritic cell-mediated T(H)17/T(reg) balance. *Inflammopharmacology* 28, 1705–1716. doi:10.1007/s10787-020-00722-6
- Zheng, S., Zhuang, T., Tang, Y., Wu, R., Xu, T., Leng, T., et al. (2021b). Leonurine protects against ulcerative colitis by alleviating inflammation and modulating intestinal microflora in mouse models. *Exp. Ther. Med.* 22, 1199. doi:10.3892/etm.2021.10633
- Zhong, Y., Liu, W., Xiong, Y., Li, Y., Wan, Q., Zhou, W., et al. (2022). Astragaloside IV alleviates ulcerative colitis by regulating the balance of Th17/Treg cells. *Phytomedicine* 104, 154287. doi:10.1016/j.phymed.2022.154287
- Zhou, F., Wang, N., Yang, L., Zhang, L.-C., Meng, L.-J., and Xia, Y.-C. (2019). Saikosaponin A protects against dextran sulfate sodium-induced colitis in mice. *Int. Immunopharmacol.* 72, 454–458. doi:10.1016/j.intimp.2019.04.024
- Zhou, J., Wang, T., Dou, Y., Huang, Y., Qu, C., Gao, J., et al. (2018a). Brusatol ameliorates 2, 4, 6-trinitrobenzenesulfonic acid-induced experimental colitis in rats: involvement of NF- κ B pathway and NLRP3 inflammasome. *Int. Immunopharmacol.* 64, 264–274. doi:10.1016/j.intimp.2018.09.008
- Zhou, K., Cheng, R., Liu, B., Wang, L., Xie, H., and Zhang, C. (2018b). Eupatilin ameliorates dextran sulphate sodium-induced colitis in mice partly through promoting AMPK activation. *Phytomedicine* 46, 46–56. doi:10.1016/j.phymed.2018.04.033
- Zhou, P., Lai, J., Li, Y., Deng, J., Zhao, C., Huang, Q., et al. (2022a). Methyl gallate alleviates acute ulcerative colitis by modulating gut microbiota and inhibiting TLR4/NF- κ B pathway. *Int. J. Mol. Sci.* 23, 14024. doi:10.3390/ijms232214024
- Zhou, Q., Zhang, W.-X., He, Z.-Q., Wu, B.-S., Shen, Z.-F., Shang, H.-T., et al. (2020). The possible anti-inflammatory effect of dehydrocostus lactone on DSS-induced colitis in mice. *Evid. Based Complement. Altern. Med.* 2020, 5659738. doi:10.1155/2020/5659738
- Zhou, Y., Chen, S., Dai, Y., Wu, L., Jin, M., Zhao, J., et al. (2023). Sinomenine attenuated dextran sulfate sodium-induced inflammatory responses by promoting 14-3-3 theta protein and inhibiting NF- κ B signaling. *J. Ethnopharmacol.* 303, 116037. doi:10.1016/j.jep.2022.116037
- Zhou, Y., Chen, S., Gu, W., Sun, X., Wang, L., and Tang, L. (2021). Sinomenine hydrochloride ameliorates dextran sulfate sodium-induced colitis in mice by modulating the gut microbiota composition whilst suppressing the activation of the NLRP3 inflammasome. *Exp. Ther. Med.* 22, 1287. doi:10.3892/etm.2021.10722
- Zhou, Y., Dou, F., Song, H., and Liu, T. (2022b). Anti-ulcerative effects of wogonin on ulcerative colitis induced by dextran sulfate sodium via Nrf2/TLR4/NF- κ B signaling pathway in BALB/c mice. *Environ. Toxicol.* 37, 954–963. doi:10.1002/tox.23457
- Zhu, L., Gu, P., and Shen, H. (2019a). Protective effects of berberine hydrochloride on DSS-induced ulcerative colitis in rats. *Int. Immunopharmacol.* 68, 242–251. doi:10.1016/j.intimp.2018.12.036
- Zhu, L., Gu, P., and Shen, H. (2019b). Gallic acid improved inflammation via NF- κ B pathway in TNBS-induced ulcerative colitis. *Int. Immunopharmacol.* 67, 129–137. doi:10.1016/j.intimp.2018.11.049
- Zhu, L., Song, Y., Liu, H., Wu, M., Gong, H., Lan, H., et al. (2021). Gut microbiota regulation and anti-inflammatory effect of β -carotene in dextran sulfate sodium-stimulated ulcerative colitis in rats. *J. Food Sci.* 86, 2118–2130. doi:10.1111/1750-3841.15684
- Zhu, L., Xu, L.-Z., Zhao, S., Shen, Z.-F., Shen, H., and Zhan, L.-B. (2020). Protective effect of baicalin on the regulation of Treg/Th17 balance, gut microbiota and short-chain fatty acids in rats with ulcerative colitis. *Appl. Microbiol. Biotechnol.* 104, 5449–5460. doi:10.1007/s00253-020-10527-w
- Zhuang, H., Lv, Q., Zhong, C., Cui, Y., He, L., Zhang, C., et al. (2021). Tiliroside ameliorates ulcerative colitis by restoring the M1/M2 macrophage balance via the HIF-1 α /glycolysis pathway. *Front. Immunol.* 12, 649463. doi:10.3389/fimmu.2021.649463
- Zohny, M. H., Alrouji, M., Alhajlah, S., AlOmeir, O., Ewees, M. G. E., Ghaffar, D. M. A., et al. (2022). Diacetyl-rhein, an anthraquinone antiarthritic agent, suppresses dextran sodium sulfate-induced inflammation in rats: a possible mechanism for a protective effect against ulcerative colitis. *Biomed. Pharmacother.* 154, 113651. doi:10.1016/j.biopha.2022.113651
- Zou, Y., Dai, S.-X., Chi, H.-G., Li, T., He, Z.-W., Wang, J., et al. (2015). Baicalin attenuates TNBS-induced colitis in rats by modulating the Th17/Treg paradigm. *Arch. Pharm. Res.* 38, 1873–1887. doi:10.1007/s12272-014-0486-2

Glossary

ACE	abundance-based coverage estimator	PDK1	Pyruvate dehydrogenase kinase 1
AMP	adenosine 5'-monophosphate	p-JAK2	phosphorylated Janus kinase 2
AMPK	AMP-activated protein kinase	PPARγ	peroxisome proliferator-activated receptor γ
CAT	catalase	p-STAT3	phosphorylated signal transducer and activator of transcription 3
CCL-5	chemokine ligand 5	p38	p38 kinase
CD	Crohn's disease	RhoA-ROCK	Ras homologous protein A-Rho kinase
CDH1	e-cadherin	ROS	reactive oxygen species
CHOP	C/EBP-homologous protein	ROCK1	Rho-associated kinase 1
COX-2	cyclooxygenase	STAT3	signal transducer and activator of transcription 3
Cxcl2	CXC motif chemokine ligand 2	SCFAs	short-chain fatty acids
ERK-1/2	extracellular-signal-regulated kinases 1/2	SNPs	single nucleotide polymorphisms
ER	endoplasmic reticulum	TA	Taurohyodeoxycholic acid
FITC-dextran	fluorescein isothiocyanate dextran	TCM	traditional Chinese medicine
GPCR	G-protein-coupled receptors	TJs	tight junctions
GPX4	glutathione peroxidase 4	Tff3	trefoil factor 3
GRP78	GSH glucose-regulated protein glutathione	TNF-α	tumor necrosis factor alpha
HNF4A	hepatocyte nuclear factor 4 alpha	TLR4	Toll-like receptor 4
HO-1	heme oxygenase-1	ULK1	unc-51 like autophagy activating kinase 1
IBD	inflammatory bowel disease	UC	Ulcerative colitis
IκB	IkappaB	ZO-1	zona occludens 1
IEC	intestinal epithelial cell	2-DG	2-deoxy-D-glucose
IFN-γ	interferon- γ		
IL-10	interleukin-10		
ILC	innate lymphoid cells		
iNOS	inducible nitric oxide synthase		
JAK/STAT	janus kinase/signal transducer and activator of transions		
JNK	c-Jun N-terminal kinase		
LOX	lipoxygenase		
mTOR	mammalian target of rapamycin		
MAPK	mitogen-activated protein kinase		
MDA	malondialdehyde		
MLC2	myosin light chain 2		
MPO	myeloperoxidase		
MTOR	mechanistic target of rapamycin kinase		
Muc1	mucin 1		
NK cells	natural killer cells		
NO	nitric oxide		
NF-κB	nuclear factor kappa-B		
NLRP3	NOD-like receptor protein 3		
NLR	NOD-like receptor		
PAMPs	pathogen-associated molecular patterns		