



OPEN ACCESS

EDITED BY

Gessica Sala,
University of Milano-Bicocca, Italy

REVIEWED BY

Nitesh Sanghai,
University of Manitoba, Canada

*CORRESPONDENCE

Priscila Pereira Sena
✉ priscila.sena@med.uni-tuebingen.de

RECEIVED 06 August 2025

ACCEPTED 29 September 2025

PUBLISHED 24 October 2025

CITATION

Pereira Sena P, Friedrich L, Villarreal A, Fath F, Sopco L, Hernández-Guillamon M, Saraiva-Pereira ML, Britton G, Weber JJ and Schmidt T (2025) Proteostasis disruption and lipid dyshomeostasis in neurodegeneration: exploring common druggable targets across sporadic and monogenic disorders. *Front. Mol. Neurosci.* 18:1681079. doi: 10.3389/fnmol.2025.1681079

COPYRIGHT

© 2025 Pereira Sena, Friedrich, Villarreal, Fath, Sopco, Hernández-Guillamon, Saraiva-Pereira, Britton, Weber and Schmidt. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Proteostasis disruption and lipid dyshomeostasis in neurodegeneration: exploring common druggable targets across sporadic and monogenic disorders

Priscila Pereira Sena^{1*}, Lea Friedrich¹, Alcibiades Villarreal^{2,3}, Florian Fath^{1,4,5}, Liubovi Sopco⁶, Mar Hernández-Guillamon⁶, Maria Luiza Saraiva-Pereira^{7,8}, Gabrielle Britton^{2,3,9}, Jonasz Jeremiasz Weber^{1,4} and Thorsten Schmidt¹

¹Institute of Medical Genetics and Applied Genomics, Eberhard Karls University Tübingen, Tübingen, Germany, ²Instituto de Investigaciones Científicas y Servicios de Alta Tecnología (INDICASAT-AIP), Ciudad del Saber, Panama City, Panama, ³Sistema Nacional de Investigación (SNI) in Panama, Secretaría Nacional de Ciencia Tecnología e Innovación (SENACYT), Panama City, Panama, ⁴Department of Human Genetics, Ruhr University Bochum, Bochum, Germany, ⁵Department of Neurology, Huntington-Center NRW, St. Josef-Hospital Bochum, Ruhr University Bochum, Bochum, Germany, ⁶Neurovascular Research Laboratory, Vall d'Hebron Research Institute (VHIR), Barcelona, Spain, ⁷Serviço de Genética Médica, Hospital de Clínicas de Porto Alegre, Porto Alegre, Brazil, ⁸Departamento de Bioquímica, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil, ⁹Centro de Vacunación e Investigación (CEVAXIN), Panama City, Panama

Neurodegenerative disorders pose an increasing burden in the aging society. These conditions share several molecular pathomechanisms, some of which may offer opportunities for therapeutic intervention. In this review, we explore a representative selection of sporadic and hereditary neurodegenerative diseases—namely Alzheimer's disease, cerebral amyloid angiopathy, and the polyQ disorders spinocerebellar ataxia types 2 and 3, as well as Huntington's disease—which all feature the accumulation of intra- or extracellular protein deposits as a hallmark. We place particular emphasis on dysregulations in proteostasis—underlying the formation of these aggregates—and the less commonly addressed disturbances in lipid metabolism. By highlighting potential mechanistic links across different classes of neurodegenerative diseases, we aim to provide new insights that may guide the identification of shared druggable targets and the development of broad-spectrum therapeutic strategies.

KEYWORDS

apolipoprotein E, ApoE, aggregates, autophagy, amyloid β , cholesterol, plaques, polyglutamine

1 Introduction

With increasing life expectancy, the heterogeneous group of neurodegenerative disorders presents a significant and growing challenge to healthcare systems worldwide. Despite decades of intensive research, effective treatment options remain limited, underscoring the need for a deeper understanding of the diverse molecular mechanisms that ultimately lead to irreversible neuronal damage and death. This is likely due to

the high heterogeneity of such conditions, which encompass both sporadic forms, such as most manifestations of Alzheimer's Disease (AD), Amyotrophic Lateral Sclerosis (ALS) or Parkinson's Disease (PD), and monogenic forms such as the hereditary triplet repeat disorders, including the polyglutamine (polyQ) diseases. These conditions vary in their age of onset, affected tissues, clinical manifestations, and underlying molecular pathways (Wilson et al., 2023; Kelser et al., 2024). For instance, while AD primarily affects memory and cognition through cortical and hippocampal pathology, PD is characterized by motor symptoms driven by dopaminergic neurodegeneration in the substantia nigra, and spinocerebellar ataxias (SCAs) predominantly impair motor circuits through cerebellar degeneration (Knopman et al., 2021; Poewe et al., 2017; Klockgether et al., 2019). This clinical and pathological diversity complicates diagnosis, treatment, and the development of broadly effective therapies.

Current treatment options remain largely symptomatic and disease-specific. In AD, acetylcholinesterase inhibitors and the NMDA receptor antagonist memantine can provide modest symptomatic relief but do not alter disease progression (Zhang et al., 2024). PD management relies heavily on dopaminergic replacement therapies such as levodopa, which improve motor symptoms but often lose effectiveness over time and do not halt neurodegeneration (Charvin et al., 2018). In ALS, drugs such as riluzole and edaravone extend survival only modestly (Jaiswal, 2019). For hereditary polyQ disorders like Huntington's disease (HD) and SCAs, unfortunately, no approved disease-modifying treatments exist to date (Tenchov et al., 2024). Collectively, these limitations highlight the urgent need for therapies that address the root causes of neurodegeneration.

At the molecular level, several pathogenic processes have been identified, including dysfunctional proteostasis leading to protein aggregation, mitochondrial and synaptic dysfunction, oxidative stress, neuroinflammation, and disturbances in lipid metabolism (Knopman et al., 2021; Poewe et al., 2017; Klockgether et al., 2019). While these mechanisms are well-studied individually, a major gap remains in understanding how they converge and interact across different neurodegenerative conditions.

In this review, we examine a selection of neurodegenerative disorders encompassing both sporadic and monogenic forms with proteopathic characteristics. By focusing on shared molecular features—particularly dysfunctional proteostasis as the driver of the hallmark protein aggregation—we draw attention to dysregulated lipid metabolism as a common contributor for this impairment. Through this perspective, we aim to support ongoing efforts toward the development of unifying therapeutic strategies capable of targeting multiple neurodegenerative conditions.

2 Neurodegenerative disorders

Neurodegenerative disorders are caused by progressive neuronal loss across multiple brain regions. With variable clinical and pathological presentations, this group consists of largely sporadic disorders such as Alzheimer's and Parkinson's disease (AD and PD, respectively) (Bali et al., 2012; Schulze et al., 2018) and other inherited diseases such as the polyglutamine disorders, which result from constitutional mutations of single genes, thus named

monogenic hereditary disorders (Pihlström et al., 2017). One of the most common hallmarks of neurodegenerative disorders is the aggregation of misfolded proteins into insoluble inclusion bodies within the nucleus or cytoplasm, such as Lewy bodies in PD, neurofibrillary tangles in AD, or polyQ aggregates in HD, as well as the formation of extracellular deposits in neuronal tissue, including neuritic amyloid plaques in AD (Klockgether et al., 2019; Knopman et al., 2021; Ross and Poirier, 2004). These deposits are a consequence of disruptions in a process collectively known as proteostasis—an intricate network of mechanisms that regulate protein synthesis, folding, trafficking, and degradation to maintain cellular protein homeostasis. Numerous molecular pathways involved in proteostasis have been identified and characterized in the context of neurodegeneration, either contributing to disease pathogenesis or acting as disease modifiers (Yerbury et al., 2016).

One potential, yet not fully understood, contributor is lipid metabolism, which is essential for both neuronal and glial function. When disturbed, it increases the risk of neurological disease, as strikingly demonstrated by the association of the apolipoprotein E (ApoE) allele $\epsilon 4$ (*APOE* $\epsilon 4$) with late-onset AD (Strittmatter et al., 1993; Kawade and Yamanaka, 2024; Yang et al., 2023). Although a direct link between protein aggregation and lipid metabolism in neurodegeneration may not be immediately apparent, emerging intersections suggest a relevant interplay (Hernandez-Diaz and Soukup, 2020), which will be explored in the following sections.

2.1 Sporadic neurodegenerative disorders

The vast majority of neurodegenerative disorders have a sporadic etiology, with only about 10% of cases considered hereditary (Dillitt et al., 2021). Non-genetic components—such as lifestyle and environmental factors—have been reported to either contribute to or protect against the development of common neurodegenerative conditions in the elderly, including AD, PD, and cerebral amyloid angiopathy (CAA) (Jäkel et al., 2022; Mentis et al., 2021).

Given the well-established role of lipid metabolism and ApoE in neurodegenerative dementias, we focus on AD and CAA as prototypical examples of predominantly sporadic neurodegenerative disorders, which are further characterized by the deposition of amyloidogenic proteins.

2.1.1 Alzheimer's disease (AD)

AD, the most common form of dementia, is characterized by cognitive impairment and neuronal loss that progress through several stages, each defined by distinct pathological and clinical features. AD is estimated to affect around 130 million individuals by 2050 if no therapies become available (Ju and Tam, 2022). A histopathological hallmark of AD is the accumulation of abnormal protein aggregates, particularly intracellular neurofibrillary tangles formed by hyperphosphorylated tau and extracellular amyloid- β (A β) plaques—composed of A β peptides generated by proteolytic cleavage of the amyloid precursor protein—in the limbic and neocortical regions (Chen et al., 2017; Zhao and Huai, 2023) (Figure 1). The microtubule-stabilizing protein tau exists in six isoforms generated by alternative splicing of the *MAPT* gene.

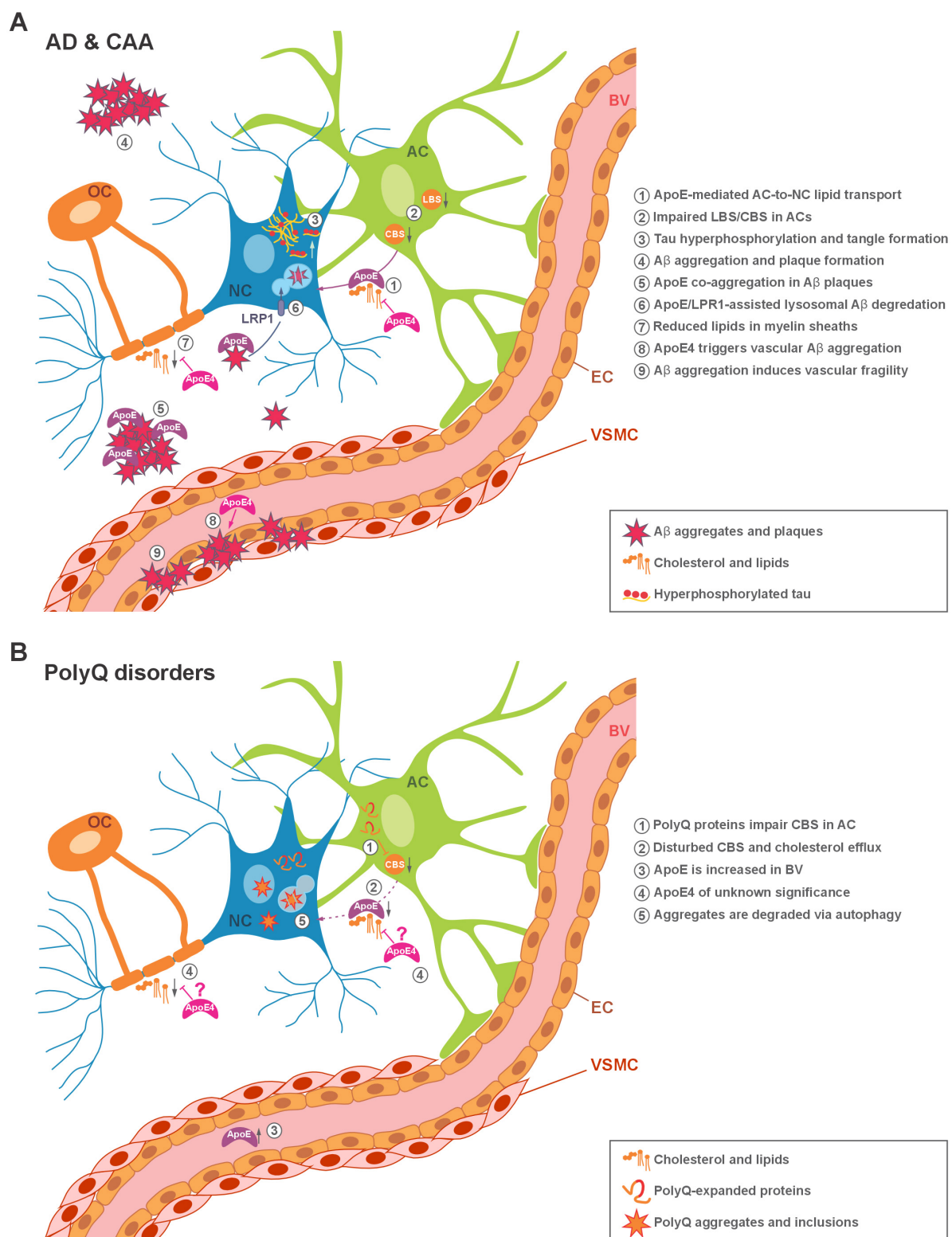


FIGURE 1

Illustration of pathogenic protein aggregate sites in the monogenic and sporadic neurodegenerative disorders covered in this review, and the known contribution of lipid metabolism and ApoE in the respective disorders. **(A)** In Alzheimer's disease (AD), neurofibrillary tangles composed of hyperphosphorylated tau accumulate within neurons, while Aβ deposits in the form of amyloid plaques are found in the extracellular space, where activated astrocytes may contribute to their uptake and removal. In cerebral amyloid angiopathy (CAA), amyloid plaques accumulate in the walls of blood vessels. **(B)** In Huntington's disease (HD) and spinocerebellar ataxias (SCAs) types 2 and 3, the respective disease-associated polyglutamine (polyQ)-expanded proteins form intracellular aggregates in the cytoplasm or nucleus of affected neurons. Characteristic events are numbered and described in the respective panel. AC, astrocyte; NC, neuronal cell; OC, oligodendrocyte; LBS, lipid biosynthesis; CBS, cholesterol biosynthesis; BV, blood vessel; EC, endothelial cells; VSMC, vascular smooth muscle cells; LRP1, lipoprotein receptor-related protein 1.

Under physiological conditions, the ratio of three-repeat (3R) to four-repeat (4R) tau isoforms is tightly regulated, but this balance becomes disrupted in tau-related neurodegenerative disorders, altering tau's phosphorylation status and aggregation propensity (Rawat et al., 2022). Hyperphosphorylation causes tau to dissociate from microtubules, after which it aggregates into tangles with neurotoxic properties. Among the 85 identified phosphorylation sites, threonine 217 has emerged as a particularly important biomarker in AD. Phosphorylation at this residue produces p-tau217, whose elevated plasma levels are associated with early stages of the disease and correlate with cognitive decline. These findings suggest that p-tau217 not only reflects underlying tau pathology but also offers promise for early diagnosis and monitoring of disease progression (Hirota et al., 2025; Martin et al., 2013). The impairment of A β clearance mechanisms seems to be a major contributor to the accumulation of A β and tau in the brain, thus reflecting a failure of the cellular machinery responsible for protein quality control (Mawuenyega et al., 2010). The development of these plaques and tangles can also be a result of an imbalance between the production and degradation of proteins (He et al., 2020).

In neurons, whose functionality greatly depends on an exactly maintained proteome, disruption of proteostasis and accumulation of toxic aggregates is particularly detrimental, as it impairs crucial cellular functions and renders neurons vulnerable to stressors such as oxidative stress, chronic inflammation, and endogenous neurotoxins (e.g., quinolinic acid), which can precipitate neurodegeneration. Over the past decade, research has increasingly focused on elucidating the molecular consequences of proteostasis disturbances in AD. For example, impairment of key intracellular mechanisms such as the ubiquitin-proteasome system (UPS) reduces the degradation of misfolded proteins, notably A β and tau, as evidenced by decreased proteasomal complex subunits and defective nuclear localization of Nrf1, which normally promotes proteasome gene expression. Concurrently, molecular chaperones show diminished activity, further compromising proper protein folding and facilitating aggregation (Batko et al., 2024). Together, these molecular failures exacerbate proteotoxic stress, contributing to a vicious cycle of neuronal dysfunction and degeneration, and highlighting potential therapeutic targets aimed at restoring UPS activity and chaperone function.

Aging is a major risk factor for AD and is closely associated with a decline in proteostasis capacity (Meller and Shalgi, 2021). As organisms age, the efficiency of protein folding, trafficking, and degradation decreases, creating an environment that supports the accumulation of misfolded proteins such as A β , tau, α -synuclein, and TAR DNA-binding protein 43 (TDP43) (Wolozin and Ivanov, 2019). Major processes, including oxidative stress, mitochondrial dysfunction, and neuroinflammation, are connected to this malfunction of proteostasis mechanisms and can lead to cell death (Jellinger, 2010). The dysfunction of protein degradation pathways, such as the UPS and autophagy, contributes to the increase of misfolded proteins in neurodegenerative diseases, including AD (Jiang et al., 2025; Nixon and Rubinsztein, 2024; Nixon and Yang, 2011; Rao et al., 2015).

Lipid metabolism and cholesterol homeostasis play a significant role in AD pathogenesis. Disruption of these processes—including

altered lipid composition and cholesterol transport—have been linked to AD pathology and disease progression. Such disturbances may exacerbate A β accumulation and tau phosphorylation, and contribute to neuroinflammation as well as myelin abnormalities (Ahmed et al., 2024; He et al., 2025; Kawade and Yamanaka, 2024; Mi et al., 2023). Importantly, restoring dysregulated lipid metabolism has been demonstrated to ameliorate AD-related pathologies (He et al., 2025; Litvinchuk et al., 2024).

One crucial modifying factor in AD is the fat-binding protein ApoE, which is central in transporting cholesterol and other lipids from astrocytes to neurons (Raulin et al., 2022; Windham and Cohen, 2024). Variants in the coding *APOE* gene have significant implications for AD and its treatment, especially in its late-onset sporadic form which occurs after the age of 65, where *APOE* is a key genetic risk factor (Islam et al., 2025). The three primary *APOE* allelic variants differ in two amino acid residues at positions 112 and 158 in the encoded protein ApoE — ϵ 2 (Cys112; Cys158), ϵ 3 (Cys112; Arg158), and ϵ 4 (Arg112; Arg158) (Serrano-Pozo et al., 2021). Strong evidence from clinical and basic research indicates that the *APOE* ϵ 4 allele is associated with an increased risk of AD (Corder et al., 1993), while the *APOE* ϵ 2 allele is linked to a decreased risk compared to the more common *APOE* ϵ 3 allele (Bu, 2009; Corder et al., 1994; Farrer et al., 1997). Mechanistically, *APOE* ϵ 4 was implicated in disturbances of the lipid metabolism, primarily associated with impaired function in lipid and cholesterol efflux from astrocytes and neurons, leading to detrimental accumulation of lipid deposits (Lin et al., 2018; Raulin et al., 2022; Sienski et al., 2021). Notably, ApoE is known to accumulate in A β plaques and trigger tau hyperphosphorylation as well as its deposition (Hou et al., 2020; Namba et al., 1991; Therriault et al., 2020; Xia et al., 2024). Moreover, ApoE seems to play a protective role by mediating the removal of A β via receptor-mediated clearance and extracellular proteolytic machineries, while the ApoE ϵ 4 presented impaired functionality in these pathways, contributing to disease-associated accumulation of extracellular plaques (Jiang et al., 2008; Kanekiyo et al., 2013; Van Acker et al., 2019).

The variations in ApoE, particularly the protective effects of the *APOE* ϵ 2 allele and the risk associated with *APOE* ϵ 4, present critical insights for AD research. Understanding these genetic factors can enhance diagnostic precision and guide the development of targeted therapies, offering significant potential for more effective treatment strategies (Hou et al., 2020; Namba et al., 1991; Therriault et al., 2020; Xia et al., 2024).

From the perspective of AD progression, all the factors mentioned above contribute and define the pathological stages of this disorder. In the asymptomatic phase, amyloid-beta (A β) plaques begin to accumulate, a process strongly influenced by lipid metabolism and ApoE function (Raulin et al., 2022). During the prodromal or mild cognitive impairment (MCI) phase, tau pathology emerges, and subtle cognitive deficits appear, with ongoing ApoE- and lipid-mediated effects on protein clearance, membrane composition, and synaptic function. In the dementia phase, widespread neuronal loss, synaptic dysfunction, and cognitive decline occur, with dysregulated ApoE and lipid homeostasis further exacerbating proteostasis impairment, inflammation, and neurodegeneration (Jack et al., 2010; Dubois et al., 2016; Zhang et al., 2024).

2.1.2 Cerebral amyloid angiopathy (CAA)

CAA is one of the main causes of lobar intracerebral hemorrhage (ICH) in the elderly, causing 5–20% of spontaneous ICH in older adults (de Bruin et al., 2024). The prevalence of CAA increases significantly with age and is observed in approximately 80–90% of individuals with AD pathology (Yamada, 2015). CAA is characterized by vascular deposition of A β in the walls of small leptomeningeal arteries and cortical blood vessels (de Bruin et al., 2024) (Figure 1). Despite its close pathological overlap with AD, CAA shows distinct clinical features and may thus act as a pathological bridge linking cerebrovascular dysfunction with other neurodegenerative diseases (Cordonnier and van der Flier, 2011).

Structurally, CAA differs from AD in terms of the predominant A β isoforms involved. Unlike in AD, shorter A β fragments are more abundantly deposited in CAA. While A β 42 is primarily deposited in parenchymal neuritic plaques in AD, CAA is characterized by the more abundant deposition of A β 40. Moreover, several studies have demonstrated that shorter A β isoforms—such as A β 37, A β 38, and A β 39—are also present in vascular deposits (Kakuda et al., 2017; Reinert et al., 2016). These more soluble isoforms are thought to follow perivascular drainage pathways, which may contribute to their selective vascular accumulation (Greenberg et al., 2020; van den Berg et al., 2024). Over time, A β accumulation leads to the loss of vascular smooth muscle cells (VSMCs). These contractile cells, believed to be of mesenchymal origin (Sinha et al., 2014), are located in the tunica media (middle layer) of small arteries and arterioles, where they play a crucial role in maintaining vessel tone, regulating cerebral blood flow, and preserving vascular integrity. In CAA, their gradual depletion—particularly in leptomeningeal and cortical arteries—weakens the vessel wall and increases its risk of rupture. Despite these changes, the precise trigger initiating peptide deposition remains unknown. However, it is widely believed that these deposits result from impaired A β clearance, rather than overproduction, in the vascular walls, ultimately compromising vessel integrity (Koemans et al., 2023; Qi and Ma, 2017).

Similar to observations in AD, proteostasis and proteolytic mechanisms critical for A β generation and clearance are disrupted in CAA (Krohn et al., 2011; Ma et al., 2010; Monro et al., 2002; Savar et al., 2024; Qi and Ma, 2017). Moreover, vascular A β accumulation appears to involve lipid components (de Oliveira et al., 2025) and autophagy, an essential mechanism that maintains cellular health and homeostasis by removing damaged proteins and organelles (Liu et al., 2023).

Autophagy is characterized by membrane structures that form the autophagosomes—double-membrane vesicles that engulf cellular material for degradation. The fusion of autophagosomes with lysosomes vesicles containing hydrolytic enzymes enables the breakdown of cellular components (He and Klionsky, 2009). Experimental studies suggest that activating autophagy may have beneficial effects in CAA (Ma et al., 2010). This is mediated by the transmembrane lipoprotein receptor-related protein 1 (LRP1), which plays a key role in A β uptake and lysosomal degradation (Bell et al., 2009; Cheung et al., 2014; Kanekiyo et al., 2012). Notably, LRP1 is also a major neuronal ApoE receptor and is involved in modulating A β pathology (Na et al., 2023; Shinohara et al., 2017; Strickland and Holtzman, 2019; Tachibana et al., 2019). Previous *in vitro* and *in vivo* studies have shown that the ApoE

protein influences multiple aspects of A β pathology, including its accumulation, clearance, conformational state, and toxicity (Rannikmäe et al., 2013). The APOE ϵ 4 allele is associated with an increased risk of CAA, as it impairs A β clearance from the brain and promotes vascular deposition. In contrast, the ϵ 2 allele—although considered protective in AD—may increase the risk of vessel wall fragility in CAA due to A β accumulation, thus predisposing to ICH recurrence. Interestingly, individuals with the APOE ϵ 2/ ϵ 4 genotype may experience a compounded pathological effect, with both enhanced A β deposition and increased vascular fragility, leading to a higher risk of early ICH recurrence (Greenberg et al., 2020; Yang et al., 2025).

These findings show that the APOE gene influences the development of CAA and could be important for diagnosis as well as potential treatment.

2.2 Inherited neurodegenerative disorders

Strictly hereditary neurodegenerative disorders, which account for approximately 10% of all cases, are genetically heterogeneous and involve mutations in genes such as *presenilin-1* (*PSEN1*) in familial AD, *leucine-rich repeat kinase 2* (*LRRK2*) and *parkin* (*PRKN*) in dominant or recessive forms of PD, and *C9orf72* in Frontotemporal Dementia (FTD) and amyotrophic lateral sclerosis (ALS) (Dilliot et al., 2021; Pihlström et al., 2017). A distinct group within inherited neurodegenerative diseases comprise the so-called polyglutamine (polyQ) disorders, which include the following nine conditions: spinobulbar muscular atrophy (SBMA), dentatorubral-pallidoluysian atrophy (DRPLA), Huntington's disease (HD), and six spinocerebellar ataxias (SCA1, SCA2, SCA3, SCA6, SCA7, and SCA17). All of these diseases follow an autosomal dominant pattern of transmission, except for SBMA, which is X-linked recessive (Orr and Zoghbi, 2007).

2.2.1 Polyglutamine (polyQ) disorders

PolyQ diseases are characterized by a CAG trinucleotide expansion in the coding region of the affected gene, leading to an elongated polyQ tract in the translated protein. The length of the CAG repeat directly influences the age at onset, with longer CAG repeat expansions correlating with earlier disease onset and more severe manifestation of symptoms (Tandon et al., 2024). The polyQ expansion alters properties of the affected protein, induces its misfolding, and results ultimately in its accumulation in the form of neuronal intracellular aggregates (Figure 1). These aggregates may contain not only the entire protein or its polyQ stretch-containing fragments, but also additional components such as chaperones, ubiquitin, ubiquitin-binding proteins, proteasomal subunits, and other vital factors such as transcription-related proteins (Havel et al., 2009; Lieberman et al., 2019). Although the disease-causative proteins of all polyQ diseases are ubiquitously expressed across different cell types, aggregate formation and cell loss is restricted to neurons, while the affected brain region differs depending on the disease. Apart from the expanded polyQ stretch, the affected proteins lack sequence homology and participate in diverse cellular processes, including transcription regulation, RNA

metabolism, protein homeostasis, and protein-protein interactions (Maiuri et al., 2017; Paulson et al., 2017). Despite differences in the affected neuronal subpopulations and the varied functions of the disease-causing proteins, polyQ diseases share several pathological features (Figure 1B), which may represent common targets for therapeutic development.

Below, we present a selected overview of spinocerebellar ataxias (SCAs) 2 and 3, along with HD, as representative polyQ disorders, and highlight their common targetable pathways, with a focus on the impact of ApoE and lipid metabolism on pathology and proteostasis.

2.2.2 Spinocerebellar ataxia type 2 (SCA2)

SCA2 (OMIM: #183090) is caused by an expansion of a CAG tract in exon 1 of the *ATXN2* gene that encodes the protein ataxin-2 (Pulst, 1993). Repeat lengths up to 31 CAG repeats are considered normal alleles (de Castilhos et al., 2014; Gardiner et al., 2019), with a high prevalence of 22 CAG repeats, being found in 90.1% of the general population (Andrés et al., 2003), while expansions of 33 and above are considered as fully penetrant for SCA2. Repeat lengths around this threshold and CAA interruptions have been additionally associated with other neurodegenerative disorders, causing recessive SCA2 or representing a risk factor for ALS and PD (Charles et al., 2007; Elden et al., 2010; Gwinn-Hardy et al., 2000; Tojima et al., 2018). Expanded CAG repeats and consequently longer polyQ tracts in ataxin-2 lead to toxicity and neurodegeneration in the cerebellum and brainstem (Bunting et al., 2022). Clinical symptoms frequently observed are progressive ataxia and dysarthria, slow saccadic eye movements, and peripheral neuropathy (Pulst, 1993).

The known physiological role of wild-type ataxin-2 includes its posttranscriptional regulatory function in RNA metabolism and translation, and its involvement in cytoplasmic stress granules (Carmo-Silva et al., 2017; Costa et al., 2024). While the polyQ expansion in ataxin-2 likely alters its function, promoting toxicity and aggregation, the role of ataxin-2 aggregates—primarily due to their infrequent nuclear localization—was initially considered of minor pathological relevance. Notably, neuronal loss and intranuclear inclusions are not necessarily concomitant in SCA2 (Huynh et al., 2000; Koyano et al., 2014). However, the presence of cytoplasmic aggregated polyQ-expanded ataxin-2, visible as granular staining, has been shown to correlate with disease progression in the SCA2 patient brain (Koyano et al., 2014; Seidel et al., 2017). Dysregulation of autophagy has been observed in both SCA2 mouse models and patient-derived samples, indicating disease-related disruptions that may impair the toxic protein clearance and promote aggregation. Conversely, activation of this degradation pathway or its upstream regulators was found to ameliorate SCA2 pathology (Afonso et al., 2022; Liu et al., 2024; Marcelo et al., 2021; Paul et al., 2018; Wardman et al., 2020).

Ataxin-2 also plays a role in lipid metabolism. Knockout models revealed various perturbations, including deficits in lipid and cholesterol metabolism (Lastres-Becker et al., 2008). In the brains of SCA2 knock-in mice, reduced levels of myelin lipids and downregulation of enzymes essential for cholesterol

biosynthesis were observed, accompanied by decreased levels of cholesterol precursor metabolites (Canet-Pons et al., 2021; Sen et al., 2019). Similarly, the primary cholesterol elimination product, 24S-hydroxycholesterol, was found to be reduced in the brains of SCA2 patients (Locci et al., 2023). However, little is known about the involvement of ApoE in SCA2, although some studies have reported increased ApoE protein or expression levels in patient blood or fibroblasts (Cornelius et al., 2017; Swarup et al., 2013).

2.2.3 Spinocerebellar ataxia type 3 (SCA3)/Machado-Joseph disease (MJD)

SCA3, or Machado-Joseph disease (MJD) (OMIM: #109150), represents the second most common polyQ disease after HD, and the most common SCA worldwide (Durr, 2010; Gardiner et al., 2019; Klockgether et al., 2019). It is caused by a CAG repeat expansion in exon 10 of the *ATXN3* gene. Repeat lengths of 12 to 44 CAGs are found in non-affected individuals, while around 56 to 87 repeats are associated with clinical manifestation of the disease. For intermediate repeat lengths, incomplete penetrance of symptoms has been reported (McLoughlin et al., 2020). Symptoms include progressive cerebellar ataxia with motor deficiencies such as gait abnormalities, coordination problems, impaired balance or oculomotor impairments, but can also include parkinsonism, sleep disturbances or sensory damage (Rüb et al., 2013).

At the molecular level, the polyQ-expanded SCA3 disease-related protein ataxin-3 is abnormally folded and accumulates as intracellular protein aggregates, which works as a bait for other proteins, ultimately disrupting multiple cellular processes (Yang et al., 2014). Ataxin-3 is a deubiquitinase and mediates protein quality control pathways such as autophagy and the UPS, which are compromised in SCA3 and can be targeted for improving the molecular phenotype (Ashkenazi et al., 2017; Blount et al., 2014; Costa Mdo and Paulson, 2012; Menzies et al., 2010; Onofre et al., 2016; Pereira Sena et al., 2021). Apart from this function, ataxin-3 has also been associated with DNA damage repair (Gao et al., 2015; Pfeiffer et al., 2017) and transcriptional regulation (Li et al., 2002). Therefore, the ramifications of mutant ataxin-3 on protein homeostasis are not limited to aggregation and aberrant degradation of the actual disease protein but also impact the turnover of multiple other cellular proteins.

Studies on pathological dysfunctions in lipid and cholesterol metabolism in SCA3 remain limited but consistently highlight their significant impact, as demonstrated in various models and patient materials (Campos et al., 2022; Putka et al., 2025; Toonen et al., 2018). Notably, restoration of cholesterol levels by viral administration of cholesterol 24-hydroxylase (CYP46A1) in SCA3 mice activated autophagy, enhanced aggregate clearance, and concurrently alleviated both neuropathology and motor deficits (Nóbrega et al., 2019).

While the main factor contributing to age at onset is the length of the CAG repeat, genetic modifiers have been identified in SCA3 (de Mattos et al., 2019; Raposo et al., 2022; Weber et al., 2024), including the *APOE* genotype. It was demonstrated that carriers of the *APOE* ϵ 2 allele present an earlier disease onset (Bettencourt et al., 2011a; Peng et al., 2014), although a later study did not come

to this conclusion (Zhou et al., 2014). There is, however, evidence that the *APOE* $\epsilon 4$ allele is associated with better performance in language and visual memory in SCA3 patients, while being also associated with rather severe speech disturbances (Chen et al., 2025). A further case report of two SCA3 patients presenting parkinsonism identified a common ApoE genotype, namely *APOE* $\epsilon 2/\epsilon 3$ (Bettencourt et al., 2011b). Since *APOE* $\epsilon 2$ has previously been associated with PD (Huang et al., 2004; Jo et al., 2021; Pang et al., 2018), it is reasonable to suggest that *APOE* $\epsilon 2$ is linked to parkinsonism in SCA3. However, the exact repercussions of the *APOE* genotype on the molecular pathogenesis of SCA3, in particular in the proteostatic networks, have not been addressed yet.

2.2.4 Huntington's disease (HD)

With an estimated global prevalence of 4.88 per 100,000 individuals, HD (OMIM: #143100) is the most common inherited neurodegenerative disease and polyQ disorder (Arrasate and Finkbeiner, 2012; Medina et al., 2022). An aberrant expansion of the glutamine-coding CAG repeat region in exon 1 of the *HTT* gene leads to disease onset, with full disease penetrance to be expected above 39 CAG repeats (Jiang et al., 2023). Although symptoms vary between affected patients, HD is typically characterized by progressive motor disability and cognitive decline, chorea, personality changes and mood disorders, speech difficulties, and impaired gait, balance, and coordination (Lieberman et al., 2019; Saudou and Humbert, 2016). Neuropathologically, HD involves early degeneration of striatal GABAergic medium spiny neurons, leading to striatal atrophy and cortical thinning years before symptom onset (Aylward et al., 2011; Ehrlich, 2012; Rosas et al., 2008; Ross and Tabrizi, 2011).

The precise molecular function of wild-type HTT protein is still unknown. However, it is suggested to be a multivalent structural scaffolding hub for proteins by mediating crucial intra- and inter-molecular protein interactions via its HEAT domains (Saudou and Humbert, 2016). The interplay of these interactors with HTT dictates its physiological role in vesicle trafficking and recycling, cell division, ciliogenesis, endocytosis, autophagy, and transcriptional regulation, with many of these pathways being compromised upon polyQ expansion (Ehrnhoefer et al., 2011; Saudou and Humbert, 2016).

In the nucleus and cytoplasm of HD brain neurons, large intracellular aggregates of polyQ-expanded HTT, termed inclusion bodies, were reported (Difiglia et al., 1997; Gutekunst et al., 1999). Although aggregates are generally believed to have detrimental effects on cell viability, a causative relation between aggregate formation and cell death has not yet been drawn. However, it is hypothesized that inclusion bodies may mediate their toxic function through their sequestration of important cellular proteins, such as transcription factors or UPS-components, essentially rendering them dysfunctional (Leverenz et al., 2007; Lutz and Peng, 2018; Riguet et al., 2021; Shahmoradian et al., 2019; Stewart and Radford, 2017).

Consequently, multiple studies have investigated ways to activate autophagy or the UPS to eliminate detrimental soluble or aggregated forms of the polyQ-expanded HTT (Bailus et al., 2021; Bhat et al., 2014; Ravikumar et al., 2004; Williams et al., 2008).

As the CAG repeat length explains only 50% of the total variance in age at HD onset, other factors, such as a perturbed lipid metabolism and, in particular, ApoE, have been proposed to contribute to HD progression and molecular pathogenesis (Figure 1B) (Block et al., 2010; Panas et al., 1999). Interestingly, clinical data on HD revealed that the *APOE* $\epsilon 4$ allele in patient carriers delayed the age at onset of HD by a mean difference of 13.6 years, compared to $\epsilon 3/\epsilon 3$ patients (Panas et al., 1999). Regardless of specific isoforms, a reduction of ApoE synthesis and secretion in HD astrocytes were found in various rodent models of HD (Valenza et al., 2010, 2015). As neurons in the adult brain are mostly dependent on astrocyte synthesis and efflux of cholesterol (Saher and Stumpf, 2015), increasing ApoE-mediated cholesterol efflux from astrocytes could potentially lessen cholesterol-dependent neuronal damage in HD (Valenza et al., 2010). Furthermore, gene-therapeutic delivery of CYP46A1 into the striatum of HD affected mice not only improves disease pathology and reduces mutant HTT aggregates, but also enhances cholesterol metabolism by upregulating the expression of cholesterologenic enzymes and ApoE (Boussicault et al., 2016; Kacher et al., 2019, 2022). Additionally, dysregulation of cholesterol metabolism in HD models has been shown to alter mitochondrial membrane (MM) fluidity, whereas administration of the neuroprotective cholesterol derivative olesoxime exerted restorative effects, potentially by enhancing MM cholesterol levels (Eckmann et al., 2014; Weber et al., 2019). HTT itself has been found to associate with lipids and undergo lipidation, and its interactions with cholesterol, lipids, or lipid membranes have been shown to influence its aggregation (Beasley et al., 2021; Lemarié et al., 2023; Stonebraker et al., 2023). These findings in HD models suggest a potentially broader link between lipid metabolism, ApoE, and proteostasis.

3 Potential points of intervention

Although still incurable, multiple therapeutic strategies have been explored for the sporadic and monogenic neurodegenerative disorders discussed in this review. These approaches include molecules acting on known dysregulated neuronal signaling pathways, small molecules targeting protein aggregation, antisense oligonucleotides or RNA-based therapies to reduce toxic protein expression, gene therapy strategies aimed at restoring normal protein function, and immunotherapies directed against extracellular aggregates (Zhang et al., 2024; de Sousa-Loureño et al., 2024; Tenchov et al., 2024). Notably, a recent report from the pharmaceutical company UniQure announced a first-time slowdown in HD progression by 75% using a surgical, microRNA-based strategy aiming at lowering the mutant HTT protein (<https://www.clinicaltrials.gov/study/NCT04120493>). Despite promising preclinical and early clinical results, most of these interventions have yet to achieve clear disease-modifying effects in patients in a practicable manner.

Since protein aggregation is a hallmark across multiple sporadic and monogenic neurodegenerative disorders, current efforts focus on reducing these intra- or extracellular deposits or their sources. Strategies to activate often-compromised proteolytic systems, such as autophagy and the UPS, include genetic and pharmacologic

TABLE 1 Selected pathway implications, ApoE involvement, and experimental therapeutic strategies targeting lipid homeostasis in sporadic and monogenic neurodegenerative disorders.

Disease (etiological form)	Impaired pathways of interest			ApoE as GM	LH-targeting therapeutic strategy for enhancing proteostasis
	Proteostasis	LH	ApoE		
AD (sporadic)	X	X	X	Yes	Rescue of lipid metabolism He et al., 2025 ; Litvinchuk et al., 2024
CAA (sporadic)	X		(X)	Yes	N/A
SCA2 (monogenic)	X	X	X	N/A	N/A
SCA3 (monogenic)	X		(X)	Yes	Restoration of cholesterol levels via CYP46A1 Nóbrega et al., 2019
HD (monogenic)	X	X	X	Yes	Restoration of cholesterol levels via CYP46A1 Boussicault et al., 2016 ; Kacher et al., 2019, 2022 Olesoxime administration Eckmann et al., 2014 ; Weber et al., 2019

AD, Alzheimer's disease; CAA, cerebral amyloid angiopathy; SCA2, spinocerebellar ataxia type 2; SCA3, spinocerebellar ataxia type 3; HD, Huntington's disease; GM, genetic modifier; LH, lipid homeostasis; N/A, not available; X, implicated; (X), limited implication.

manipulation of pathway-related genes, modification of their upstream regulators or effector elements, and the targeting of substrate proteins via posttranslational modifications ([Dantuma and Bott, 2014](#); [Le Guerroué and Youle, 2021](#); [Nixon and Rubinsztein, 2024](#)). However, thinking beyond conventional approaches by including additional dysregulated, targetable pathways can broaden the strategic range. This may enable the development of novel, potentially more feasible interventions, that not only restore proteostasis but also address other pathologically impaired molecular mechanisms. Here, observations in monogenic diseases with a clearer molecular etiology can offer an advantage in assessing the robustness of identified points of action. One such compromised, broader pathway may be lipid metabolism—particularly the involvement of one of its key components, ApoE, across various neurodegenerative disorders ([Estes et al., 2021](#); [Fernández-Calle et al., 2022](#)). Numerous studies have demonstrated that lipid metabolism and autophagy modulate each other reciprocally, a relationship that becomes especially apparent when considering that autophagosomes and lysosomes are lipid-membrane vesicles, and that one autophagy protein, LC3, requires lipidation for activation ([Jarocki et al., 2024](#); [Xie et al., 2020](#)). The regulatory effects of lipids on the UPS are less well understood. However, ubiquitination and the UPS play important roles in regulating lipid biosynthesis and turnover ([Jiang and Song, 2014](#); [Loix et al., 2024](#)). Consequently, direct or indirect enhancement of autophagy may alleviate the burden on an overwhelmed UPS by restoring proteostasis—while simultaneously exerting beneficial effects on UPS-controlled lipid homeostasis. Exemplary studies conducted in SCA3, HD and, to a lesser extent, SCA2 models convincingly demonstrated that modifying cholesterol biosynthesis can ameliorate disease symptoms *in vivo* by enhancing autophagy and the UPS ([Kacher et al., 2019](#); [Nóbrega et al., 2019](#)) ([Table 1](#)). Similarly beneficial effects were observed with small molecules that counteracted the autophagy-suppressing influence of the ApoE ε4 allele ([Balasubramaniam et al., 2024](#); [Parcon et al., 2018](#)).

Pursuing comparable strategies across different neurodegenerative diseases and rigorously evaluating their

effects and involved pathways will be essential to assess their potential as targets for unified therapeutic approaches.

4 Conclusion

The complexity and heterogeneity of the pathways affected in neurodegenerative disorders present major challenges for therapeutic development, contributing to the ongoing lack of effective treatments for many of these diseases. While pathways are well elaborated for AD, monogenic models such as polyQ disorders represent a better paradigm for analyzing molecular pathogenesis, since they are likely less challenging to be reproduced in cell and animal models.

The identification of common molecular targets—such as ApoE—across interconnected pathways like lipid homeostasis and proteostasis, and across multiple disorders ([Table 1](#)), has renewed interest in the search for effective therapies. A key challenge in this context will be to unravel the distinct contributions of ApoE's different variants to the disruption or maintenance of proteostasis, specially in monogenic neurodegenerative disorders—where the role of lipid metabolism and ApoE still remain underinvestigated. Once clarified, ApoE may emerge as a central player and promising target for therapeutic intervention in neurodegeneration.

Author contributions

PP: Conceptualization, Visualization, Writing – original draft, Writing – review & editing. LF: Conceptualization, Visualization, Writing – original draft, Writing – review & editing. AV: Conceptualization, Visualization, Writing – original draft, Writing – review & editing. FF: Visualization, Writing – original draft, Writing – review & editing. LS: Writing – original draft, Writing – review & editing. MH: Funding acquisition, Writing – review & editing. MS-P: Funding acquisition, Writing – original draft, Writing – review & editing. GB: Funding acquisition, Writing – review & editing. JW: Conceptualization, Writing – original draft,

Writing – review & editing. TS: Funding acquisition, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. The ELAPSE-ND (European-Latin American and Caribbean collaborative project on personalized medicine for neurological disorders) network was funded by the Rio Grande do Sul State Research Support Foundation (FAPERGS, Brazil, Grant ID 23/2551-0000978-1), the Federal Ministry of Education and Research (BMBF, Germany, Grant ID 01DN23016), the Institute of Health Carlos III (ISCIII, Spain, Grant ID AC22/00051), and the National Secretary of Science, Technology and Innovation (SENACYT, Panama, Grant ID EU-LAC-2022-198/ELAPSE-ND) within the EU-LAC framework (4th EU-LAC Joint Call in STI 2022). AV and GB received funding from INDICASAT AIP (grant ID: IGI-2021-002). JJW received funding from the German Research Foundation (DFG; research grant number WE 6585/1-1). MLSP is funded by the National Council for Scientific and Technological Development (CNPq; research ID 316994/2021-9). LS received funding from the European Union's Horizon Europe research and innovation programme under the Marie Skłodowska-Curie grant agreement No 101126533.

Acknowledgments

We acknowledge support from the Open Access Publication Fund of the University of Tübingen.

References

- Afonso, I. T., Lima, P., Conceição, A., Matos, C. A., and Nóbrega, C. (2022). Mutant ataxin-2 expression in aged animals aggravates neuropathological features associated with spinocerebellar ataxia type 2. *Int. J. Mol. Sci.* 23:1896. doi: 10.3390/ijms231911896
- Ahmed, H., Wang, Y., Griffiths, W. J., Levey, A. I., Pikuleva, I., Liang, S. H., et al. (2024). Brain cholesterol and Alzheimer's disease: challenges and opportunities in probe and drug development. *Brain* 147, 1622–1635. doi: 10.1093/brain/awae028
- Andrés, A. M., Lao, O., Soldevila, M., Calafell, F., and Bertranpetit, J. (2003). Dynamics of CAG repeat loci revealed by the analysis of their variability. *Hum. Mutat.* 21, 61–70. doi: 10.1002/humu.10151
- Arrasate, M., and Finkbeiner, S. (2012). Protein aggregates in Huntington's disease. *Exp. Neurol.* 238, 1–11. doi: 10.1016/j.expneurol.2011.12.013
- Ashkenazi, A., Bento, C. F., Rickerts, T., Vicinanza, M., Siddiqi, F., Pavel, M., et al. (2017). Polyglutamine tracts regulate beclin 1-dependent autophagy. *Nature* 545, 108–111. doi: 10.1038/nature22078
- Aylward, E. H., Nopoulos, P. C., Ross, C. A., Langbehn, D. R., Pierson, R. K., Mills, J. A., et al. (2011). Longitudinal change in regional brain volumes in prodromal Huntington disease. *J. Neurol. Neurosurg. Psychiatr.* 82, 405–410. doi: 10.1136/jnnp.2010.208264
- Bailus, B. J., Scheeler, S. M., Simons, J., Sanchez, M. A., Tshilenge, K. T., Creus-Muncunill, J., et al. (2021). Modulating FKBP5/FKBP51 and autophagy lowers HTT (huntingtin) levels. *Autophagy* 17, 4119–4140. doi: 10.1080/15548627.2021.1904489
- Balasubramaniam, M., Narasimhappagari, J., Liu, L., Ganne, A., Ayyadevara, S., Atturi, R., et al. (2024). Rescue of ApoE4-related lysosomal autophagic failure in Alzheimer's disease by targeted small molecules. *Commun. Biol.* 7:60. doi: 10.1038/s42003-024-05767-9
- Bali, J., Gheinani, A. H., Zurbriggen, S., and Rajendran, L. (2012). Role of genes linked to sporadic Alzheimer's disease risk in the production of β -amyloid peptides. *Proc. Natl. Acad. Sci. USA.* 109, 15307–15311. doi: 10.1073/pnas.1201632109
- Batko, J., Antosz, K., Miśków, W., Pszczółowska, M., Walczak, K., and Leszek, J. (2024). Chaperones-A new class of potential therapeutic targets in Alzheimer's disease. *Int. J. Mol. Sci.* 25:ijms25063401. doi: 10.3390/ijms25063401
- Beasley, M., Groover, S., Valentine, S. J., and Legleiter, J. (2021). Lipid headgroups alter huntingtin aggregation on membranes. *Biochim. Biophys. Acta Biomembr.* 1863:183497. doi: 10.1016/j.bbamem.2020.183497
- Bell, R. D., Deane, R., Chow, N., Long, X., Sagare, A., Singh, I., et al. (2009). SRF and myocardin regulate LRP-mediated amyloid-beta clearance in brain vascular cells. *Nat. Cell Biol.* 11, 143–153. doi: 10.1038/ncb1819
- Bettencourt, C., Raposo, M., Kazachkova, N., Cymbron, T., Santos, C., Kay, T., et al. (2011a). The APOE ϵ 2 allele increases the risk of earlier age at onset in Machado-Joseph disease. *Arch. Neurol.* 68, 1580–3. doi: 10.1001/archneurol.2011.636
- Bettencourt, C., Santos, C., Coutinho, P., Rizzu, P., Vasconcelos, J., Kay, T., et al. (2011b). Parkinsonian phenotype in Machado-Joseph disease (MJD/SCA3): a two-case report. *BMC Neurol.* 11:131. doi: 10.1186/1471-2377-11-131
- Bhat, K. P., Yan, S., Wang, C. E., Li, S., and Li, X. J. (2014). Differential ubiquitination and degradation of huntingtin fragments modulated by ubiquitin-protein ligase E3A. *Proc. Natl. Acad. Sci. USA.* 111, 5706–5711. doi: 10.1073/pnas.1402215111
- Block, R. C., Dorsey, E. R., Beck, C. A., Brenna, J. T., and Shoulson, I. (2010). Altered cholesterol and fatty acid metabolism in Huntington disease. *J. Clin. Lipidol.* 4, 17–23. doi: 10.1016/j.jacl.2009.11.003

Conflict of interest

GB was employed by Centro de Vacunación e Investigación (CEVAXIN).

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Generative AI statement

The author(s) declare that no Gen AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- Blount, J. R., Tsou, W. L., Ristic, G., Burr, A. A., Ouyang, M., Galante, H., et al. (2014). Ubiquitin-binding site 2 of ataxin-3 prevents its proteasomal degradation by interacting with Rad23. *Nat. Commun.* 5:4638. doi: 10.1038/ncomms5638
- Boussicault, L., Alves, S., Lamazière, A., Planques, A., Heck, N., Moumné, L., et al. (2016). CYP46A1, the rate-limiting enzyme for cholesterol degradation, is neuroprotective in Huntington's disease. *Brain* 139, 953–970. doi: 10.1093/brain/awv384
- Bu, G. (2009). Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat. Rev. Neurosci.* 10, 333–344. doi: 10.1038/nrn2620
- Bunting, E. L., Hamilton, J., and Tabrizi, S. J. (2022). Polyglutamine diseases. *Curr. Opin. Neurobiol.* 72, 39–47. doi: 10.1016/j.conb.2021.07.001
- Campos, A. B., Duarte-Silva, S., Fernandes, B., Das Neves, S. P., Marques, F., Teixeira-Castro, A., et al. (2022). Profiling microglia in a mouse model of Machado-Joseph disease. *Biomedicine* 10:10020237. doi: 10.3390/biomedicine10020237
- Canet-Pons, J., Sen, N. E., Arsović, A., Almaguer-Mederos, L. E., Halbach, M. V., Key, J., et al. (2021). Atxn2-CAG100-KnockIn mouse spinal cord shows progressive TDP43 pathology associated with cholesterol biosynthesis suppression. *Neurobiol. Dis.* 152:105289. doi: 10.1002/cne.25228
- Carmo-Silva, S., Nobrega, C., Pereira de Almeida, L., and Cavadas, C. (2017). Unraveling the Role of Ataxin-2 in Metabolism. *Trends Endocrinol. Metab.* 28, 309–318. doi: 10.1016/j.tem.2016.12.006
- Charles, P., Camuzat, A., Benammar, N., Sellal, F., Destée, A., Bonnet, A. M., et al. (2007). Are interrupted SCA2 CAG repeat expansions responsible for parkinsonism? *Neurology* 69, 1970–1975. doi: 10.1212/01.wnl.0000269323.21969.db
- Charvin, D., Medori, R., Hauser, R. A., and Rascol, O. (2018). Therapeutic strategies for Parkinson disease: beyond dopaminergic drugs. *Nat. Rev. Drug Discov.* 17:844. doi: 10.1038/nrd.2018.184
- Chen, G. F., Xu, T. H., Yan, Y., Zhou, Y. R., Jiang, Y., Melcher, K., et al. (2017). Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol. Sin.* 38, 1205–1235. doi: 10.1038/aps.2017.28
- Chen, X., Lin, K., Ye, Z., Qiu, L., Qiu, Y., Yuan, R., et al. (2025). Apolipoprotein E epsilon4 allele is associated with better performance language and visual memory in spinocerebellar ataxia type 3. *Eur. J. Neurol.* 32:e70017. doi: 10.1111/ene.70017
- Cheung, C., Goh, Y. T., Zhang, J., Wu, C., and Guccione, E. (2014). Modeling cerebrovascular pathophysiology in amyloid- β metabolism using neural-crest-derived smooth muscle cells. *Cell Rep.* 9, 391–401. doi: 10.1016/j.celrep.2014.08.065
- Corder, E. H., Saunders, A. M., Risch, N. J., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., et al. (1994). Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. *Nat. Genet.* 7, 180–184. doi: 10.1038/ng0694-180
- Corder, E. H., Saunders, A. M., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., Small, G. W., et al. (1993). Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261, 921–923. doi: 10.1126/science.8346443
- Cordonnier, C., and van der Flier, W. M. (2011). Brain microbleeds and Alzheimer's disease: innocent observation or key player? *Brain* 134, 335–344. doi: 10.1093/brain/awq321
- Cornelius, N., Wardman, J. H., Hargreaves, I. P., Neergheen, V., Bie, A. S., Tümer, Z., et al. (2017). Evidence of oxidative stress and mitochondrial dysfunction in spinocerebellar ataxia type 2 (SCA2) patient fibroblasts: effect of coenzyme Q10 supplementation on these parameters. *Mitochondrion* 34, 103–114. doi: 10.1016/j.mito.2017.03.001
- Costa Mdo, C., and Paulson, H. L. (2012). Toward understanding Machado-Joseph disease. *Prog. Neurobiol.* 97, 239–257. doi: 10.1016/j.pneurobio.2011.11.006
- Costa, R. G., Conceição, A., Matos, C. A., and Nóbrega, C. (2024). The polyglutamine protein ATXN2: from its molecular functions to its involvement in disease. *Cell Death Dis.* 15:415. doi: 10.1038/s41419-024-06812-5
- Dantuma, N. P., and Bott, L. C. (2014). The ubiquitin-proteasome system in neurodegenerative diseases: precipitating factor, yet part of the solution. *Front. Mol. Neurosci.* 7:70. doi: 10.3389/fnmol.2014.00070
- de Bruin, O. F., Voigt, S., Schoones, J. W., Moojen, W. A., Van Etten, E. S., and Wermer, M. J. H. (2024). Surgical intervention for cerebral amyloid angiopathy-related lobar intracerebral hemorrhage: a systematic review. *J. Neurosurg.* 141, 955–965. doi: 10.3171/2024.1.JNS231852
- de Castilhos, R. M., Furtado, G. V., Gheno, T. C., Schaeffer, P., Russo, A., Barsottini, O., et al. (2014). Spinocerebellar ataxias in Brazil—frequencies and modulating effects of related genes. *Cerebellum* 13, 17–28. doi: 10.1007/s12311-013-010-3
- de Mattos, E. P., Kolbe Muskopf, M., Bielefeldt Leotti, V., Saraiva-Pereira, M. L., and Jardim, L. B. (2019). Genetic risk factors for modulation of age at onset in Machado-Joseph disease/spinocerebellar ataxia type 3: a systematic review and meta-analysis. *J. Neurol. Neurosurg. Psychiatr.* 90, 203–210. doi: 10.1136/jnnp-2018-319200
- de Oliveira, A. P., Baghel, D., Holcombe, B., Chase, W., Ward, T., Wang, S. J., et al. (2025). Lipid mediated formation of antiparallel aggregates in cerebral amyloid angiopathy. *Acta Neuropathol.* 150:3. doi: 10.1007/s00401-025-02911-5
- de Sousa-Lourenço, J., Silva, A. C., Pereira De Almeida, L., and Nobre, R. J. (2024). Molecular therapy for polyQ disorders: from bench to clinical trials. *Trends Mol. Med.* 30, 804–808. doi: 10.1016/j.molmed.2024.05.004
- Difiglia, M., Sapp, E., Chase, K. O., Davies, S. W., Bates, G. P., Vonsattel, J. P., et al. (1997). Aggregation of huntingtin in neuronal intranuclear inclusions and dystrophic neurites in brain. *Science* 277, 1990–1993. doi: 10.1126/science.277.5334.1990
- Dillio, A. A., Abdelhady, A., Sunderland, K. M., Farhan, S. M. K., Abrahao, A., Binns, M. A., et al. (2021). Contribution of rare variant associations to neurodegenerative disease presentation. *NPJ Genom. Med.* 6:80. doi: 10.1038/s41525-021-00243-3
- Dubois, B., Hampel, H., Feldman, H. H., Scheltens, P., Aisen, P., Andrieu, S., et al. (2016). Preclinical Alzheimer's disease: Definition, natural history, and diagnostic criteria. *Alzheimers. Dement.* 12, 292–323. doi: 10.1016/j.jalz.2016.02.002
- Durr, A. (2010). Autosomal dominant cerebellar ataxias: polyglutamine expansions and beyond. *Lancet Neurol.* 9, 885–894. doi: 10.1016/S1474-4422(10)70183-6
- Eckmann, J., Clemens, L. E., Eckert, S. H., Hagl, S., Yu-Taeger, L., Bordet, T., et al. (2014). Mitochondrial membrane fluidity is consistently increased in different models of Huntington disease: restorative effects of oleosxime. *Mol. Neurobiol.* 50, 107–118. doi: 10.1007/s12035-014-8663-3
- Ehrlich, M. E. (2012). Huntington's disease and the striatal medium spiny neuron: cell-autonomous and non-cell-autonomous mechanisms of disease. *Neurotherapeutics* 9, 270–284. doi: 10.1007/s13311-012-0112-2
- Ehrhoefer, D. E., Sutton, L., and Hayden, M. R. (2011). Small changes, big impact: posttranslational modifications and function of huntingtin in Huntington disease. *Neuroscientist* 17, 475–492. doi: 10.1177/1073858410390378
- Elden, A. C., Kim, H. J., Hart, M. P., Chen-Plotkin, A. S., Johnson, B. S., Fang, X., et al. (2010). Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature* 466, 1069–1075. doi: 10.1038/nature09320
- Estes, R. E., Lin, B., Khera, A., and Davis, M. Y. (2021). Lipid Metabolism Influence on Neurodegenerative Disease Progression: is the vehicle as important as the cargo? *Front. Mol. Neurosci.* 14:788695. doi: 10.3389/fnmol.2021.788695
- Farrer, L. A., Cupples, L. A., Haines, J. L., Hyman, B., Kukull, W. A., Mayeux, R., et al. (1997). Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. a meta-analysis. APOE and Alzheimer disease meta analysis consortium. *JAMA* 278, 1349–1356.
- Fernández-Calle, R., Konings, S. C., Frontiñán-Rubio, J., García-Revilla, J., Camprubi-Ferrer, L., Svensson, M., et al. (2022). APOE in the bulleye of neurodegenerative diseases: impact of the APOE genotype in Alzheimer's disease pathology and brain diseases. *Mol. Neurodegener.* 17:62. doi: 10.1186/s13024-022-00566-4
- Gao, R., Liu, Y., Silva-Fernandes, A., Fang, X., Paulucci-Holthauzen, A., Chatterjee, A., et al. (2015). Inactivation of PNKP by mutant ATXN3 triggers apoptosis by activating the DNA damage-response pathway in SCA3. *PLoS Genet.* 11:e1004834. doi: 10.1371/journal.pgen.1004834
- Gardiner, S. L., Boogaard, M. W., Trompet, S., De Mutsert, R., Rosendaal, F. R., Gussekloo, J., et al. (2019). Prevalence of carriers of intermediate and pathological polyglutamine disease-associated alleles among large population-based cohorts. *JAMA Neurol.* 76, 650–656. doi: 10.1001/jamaneuro.2019.0423
- Greenberg, S. M., Bacskai, B. J., Hernandez-Guillamon, M., Pruzin, J., Sperling, R., and Van Veluw, S. J. (2020). Cerebral amyloid angiopathy and Alzheimer disease - one peptide, two pathways. *Nat. Rev. Neurol.* 16, 30–42. doi: 10.1038/s41582-019-0281-2
- Gutkunst, C. A., Li, S. H., Yi, H., Mulroy, J. S., Kuemmerle, S., Jones, R., et al. (1999). Nuclear and neuropil aggregates in Huntington's disease: relationship to neuropathology. *J. Neurosci.* 19, 2522–2534. doi: 10.1523/JNEUROSCI.19-07-02522.1999
- Gwinn-Hardy, K., Chen, J. Y., Liu, H. C., Liu, T. Y., Boss, M., Seltzer, W., et al. (2000). Spinocerebellar ataxia type 2 with parkinsonism in ethnic Chinese. *Neurology* 55, 800–805. doi: 10.1212/wnl.55.6.800
- Havel, L. S., Li, S., and Li, X. J. (2009). Nuclear accumulation of polyglutamine disease proteins and neuropathology. *Mol. Brain* 2:21. doi: 10.1186/1756-6606-2-21
- He, C., and Klionsky, D. J. (2009). Regulation mechanisms and signaling pathways of autophagy. *Annu. Rev. Genet.* 43, 67–93. doi: 10.1146/annurev-genet-102808-114910
- He, S., Xu, Z., and Han, X. (2025). Lipidome disruption in Alzheimer's disease brain: detection, pathological mechanisms, and therapeutic implications. *Mol. Neurodegener.* 20:11. doi: 10.1186/s13024-025-00803-6
- He, Z., Yang, Y., Xing, Z., Zuo, Z., Wang, R., Gu, H., et al. (2020). Intraperitoneal injection of IFN- γ restores microglial autophagy, promotes amyloid- β clearance and improves cognition in APP/PS1 mice. *Cell Death Dis.* 11:440. doi: 10.1038/s41419-020-2644-4
- Hernandez-Diaz, S., and Soukup, S. F. (2020). The role of lipids in autophagy and its implication in neurodegeneration. *Cell Stress* 4, 167–186. doi: 10.15698/cst2020.07.225
- Hirota, Y., Sakakibara, Y., Morishima, M., Sano, T., Hara, M., Arakawa, A., et al. (2025). Biomarker-related phospho-tau217 appears in synapses around A β plaques prior to tau tangle in cerebral cortex of preclinical Alzheimer's disease. *Cell Rep.* 44:116203. doi: 10.1016/j.celrep.2025.116203

- Hou, T. T., Han, Y. D., Cong, L., Liu, C. C., Liang, X. Y., Xue, F. Z., et al. (2020). Apolipoprotein E facilitates amyloid- β oligomer-induced tau phosphorylation. *J. Alzheimers. Dis.* 74, 521–534. doi: 10.3233/JAD-190711
- Huang, X., Chen, P. C., and Poole, C. (2004). APOE-[epsilon]2 allele associated with higher prevalence of sporadic Parkinson disease. *Neurology* 62, 2198–2202. doi: 10.1212/01.wnl.0000130159.28215.6a
- Huynh, D. P., Figueroa, K., Hoang, N., and Pulst, S. M. (2000). Nuclear localization or inclusion body formation of ataxin-2 are not necessary for SCA2 pathogenesis in mouse or human. *Nat. Genet.* 26, 44–50. doi: 10.1038/79162
- Islam, S., Noorani, A., Sun, Y., Michikawa, M., and Zou, K. (2025). Multifunctional role of apolipoprotein E in neurodegenerative diseases. *Front. Aging Neurosci.* 17:1535280. doi: 10.3389/fnagi.2025.1535280
- Jack, C. R., Jr., Knopman, D. S., Jagust, W. J., Shaw, L. M., et al. (2010). Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol.* 9, 119–128. doi: 10.1016/S1474-4422(09)70299-6
- Jaiswal, M. K. (2019). Riluzole and edaravone: a tale of two amyotrophic lateral sclerosis drugs. *Med. Res. Rev.* 39, 733–748. doi: 10.1002/med.21528
- Jäkel, L., De Kort, A. M., Klijn, C. J. M., Schreuder, F., and Verbeek, M. M. (2022). Prevalence of cerebral amyloid angiopathy: a systematic review and meta-analysis. *Alzheimers. Dement.* 18, 10–28. doi: 10.1002/alz.12366
- Jarocki, M., Turek, K., Saczko, J., Tarek, M., and Kulbacka, J. (2024). Lipids associated with autophagy: mechanisms and therapeutic targets. *Cell Death Discov* 10:460. doi: 10.1038/s41420-024-02224-8
- Jellinger, K. A. (2010). Basic mechanisms of neurodegeneration: a critical update. *J. Cell. Mol. Med.* 14, 457–487. doi: 10.1111/j.1582-4934.2010.01010.x
- Jiang, A., Handley, R. R., Lehnert, K., and Snell, R. G. (2023). From pathogenesis to therapeutics: a review of 150 years of huntington's disease research. *Int. J. Mol. Sci.* 24:ijms241613021. doi: 10.3390/ijms241613021
- Jiang, Q., Lee, C. Y., Mandrekar, S., Wilkinson, B., Cramer, P., Zelcer, N., et al. (2008). ApoE promotes the proteolytic degradation of Abeta. *Neuron* 58, 681–693. doi: 10.1016/j.neuron.2008.04.010
- Jiang, S., Srikanth, M., Serpe, R., Yavari, S., Gaur, P., Collins, G. A., et al. (2025). Early proteasome downregulation and dysfunction drive proteostasis failure in Alzheimer's disease. *Brain* 9:awaf222. doi: 10.1093/brain/awaf222
- Jiang, W., and Song, B. L. (2014). Ubiquitin ligases in cholesterol metabolism. *Diabetes Metab. J.* 38, 171–180. doi: 10.4093/dmj.2014.38.3.171
- Jo, S., Kim, S. O., Park, K. W., Lee, S. H., Hwang, Y. S., and Chung, S. J. (2021). The role of APOE in cognitive trajectories and motor decline in Parkinson's disease. *Sci. Rep.* 11:7819. doi: 10.1038/s41598-021-86483-w
- Ju, Y., and Tam, K. Y. (2022). Pathological mechanisms and therapeutic strategies for Alzheimer's disease. *Neural. Regen. Res.* 17, 543–549. doi: 10.4103/1673-5374.320970
- Kacher, R., Lamazière, A., Heck, N., Kappes, V., Mounier, C., Despres, G., et al. (2019). CYP46A1 gene therapy deciphers the role of brain cholesterol metabolism in Huntington's disease. *Brain* 142, 2432–2450. doi: 10.1093/brain/awz174
- Kacher, R., Mounier, C., Caboche, J., and Betuing, S. (2022). Altered cholesterol homeostasis in Huntington's Disease. *Front. Aging Neurosci.* 14:797220. doi: 10.3389/fnagi.2022.797220
- Kakuda, N., Miyasaka, T., Iwasaki, N., Nirasawa, T., Wada-Kakuda, S., Takahashi-Fujigasaki, J., et al. (2017). Distinct deposition of amyloid- β species in brains with Alzheimer's disease pathology visualized with MALDI imaging mass spectrometry. *Acta Neuropathol. Commun.* 5:73. doi: 10.1186/s40478-017-0477-x
- Kanekiyo, T., Cirrito, J. R., Liu, C. C., Shinohara, M., Li, J., Schuler, D. R., et al. (2013). Neuronal clearance of amyloid- β by endocytic receptor LRP1. *J. Neurosci.* 33, 19276–19283. doi: 10.1523/JNEUROSCI.3487-13.2013
- Kanekiyo, T., Liu, C. C., Shinohara, M., Li, J., and Bu, G. (2012). LRP1 in brain vascular smooth muscle cells mediates local clearance of Alzheimer's amyloid- β . *J. Neurosci.* 32, 16458–16465. doi: 10.1523/JNEUROSCI.3987-12.2012
- Kawade, N., and Yamanaka, K. (2024). Novel insights into brain lipid metabolism in Alzheimer's disease: Oligodendrocytes and white matter abnormalities. *FEBS Open Bio.* 14, 194–216. doi: 10.1002/2211-5463.13661
- Kelser, B. M., Teichner, E. M., Subtilre, R. C., and Hoss, K. N. (2024). A review of proposed mechanisms for neurodegenerative disease. *Front. Aging Neurosci.* 16:1370580. doi: 10.3389/fnagi.2024.1370580
- Klockgether, T., Mariotti, C., and Paulson, H. L. (2019). Spinocerebellar ataxia. *Nat. Rev. Dis. Primers* 5:24. doi: 10.1038/s41572-019-0074-3
- Knopman, D. S., Amieva, H., Petersen, R. C., Chételat, G., Holtzman, D. M., Hyman, B. T., et al. (2021). Alzheimer disease. *Nat. Rev. Dis. Primers* 7:33. doi: 10.1038/s41572-021-00269-y
- Koemans, E. A., Chhatwal, J. P., Van Veluw, S. J., Van Etten, E. S., Van Osch, M. J. P., Van Walderveen, M. A. A., et al. (2023). Progression of cerebral amyloid angiopathy: a pathophysiological framework. *Lancet Neurol.* 22, 632–642. doi: 10.1016/S1474-4422(23)00114-X
- Koyano, S., Yagishita, S., Kuroiwa, Y., Tanaka, F., and Uchiyama, T. (2014). Neuropathological staging of spinocerebellar ataxia type 2 by semiquantitative 1C2-positive neuron typing: nuclear translocation of cytoplasmic 1C2 underlies disease progression of spinocerebellar ataxia type 2. *Brain Pathol.* 24, 599–606. doi: 10.1111/bpa.12146
- Krohn, M., Lange, C., Hofrichter, J., Scheffler, K., Stenzel, J., Steffen, J., et al. (2011). Cerebral amyloid- β proteostasis is regulated by the membrane transport protein ABCG1 in mice. *J. Clin. Invest.* 121, 3924–3931. doi: 10.1172/JCI57867
- Lastres-Becker, I., Brodesser, S., Lütjohann, D., Azizov, M., Buchmann, J., Hintermann, E., et al. (2008). Insulin receptor and lipid metabolism pathology in ataxin-2 knock-out mice. *Hum. Mol. Genet.* 17, 1465–1481. doi: 10.1093/hmg/ddn035
- Le Guerroué, F., and Youle, R. J. (2021). Ubiquitin signaling in neurodegenerative diseases: an autophagy and proteasome perspective. *Cell Death Differ.* 28, 439–454. doi: 10.1038/s41418-020-00667-x
- Lemarié, F. L., Sanders, S. S., Nguyen, Y., Martin, D. D. O., and Hayden, M. R. (2023). Full-length huntingtin is palmitoylated at multiple sites and post-translationally myristoylated following caspase-cleavage. *Front. Physiol.* 14:1086112. doi: 10.3389/fphys.2023.1086112
- Leverenz, J. B., Umar, I., Wang, Q., Montine, T. J., Mcmillan, P. J., Tsuang, D. W., et al. (2007). Proteomic identification of novel proteins in cortical lewy bodies. *Brain Pathol.* 17, 139–145. doi: 10.1111/j.1750-3639.2007.00048.x
- Li, F., Macfarlan, T., Pittman, R. N., and Chakravarti, D. (2002). Ataxin-3 is a histone-binding protein with two independent transcriptional corepressor activities. *J. Biol. Chem.* 277, 45004–45012. doi: 10.1074/jbc.M205259200
- Lieberman, A. P., Shakkottai, V. G., and Albin, R. L. (2019). Polyglutamine Repeats in Neurodegenerative Diseases. *Annu. Rev. Pathol.* 14, 1–27. doi: 10.1146/annurev-pathmechdis-012418-012857
- Lin, Y. T., Seo, J., Gao, F., Feldman, H. M., Wen, H. L., Penney, J., et al. (2018). APOE4 causes widespread molecular and cellular alterations associated with Alzheimer's disease phenotypes in human iPSC-derived brain cell types. *Neuron* 98, 1141–1154. doi: 10.1016/j.neuron.2018.05.008
- Litvinchuk, A., Suh, J. H., Guo, J. L., Lin, K., Davis, S. S., Bien-Ly, N., et al. (2024). Amelioration of Tau and ApoE4-linked glial lipid accumulation and neurodegeneration with an LXR agonist. *Neuron* 112, 384–403. doi: 10.1016/j.neuron.2023.10.023
- Liu, S., Yao, S., Yang, H., Liu, S., and Wang, Y. (2023). Autophagy: regulator of cell death. *Cell Death Dis.* 14:648. doi: 10.1038/s41419-023-06154-8
- Liu, Y. J., Wang, J. Y., Zhang, X. L., Jiang, L. L., and Hu, H. Y. (2024). Ataxin-2 sequesters raptor into aggregates and impairs cellular mTORC1 signaling. *FEBS J.* 291, 1795–1812. doi: 10.1111/febs.17081
- Locci, S., Nidiaci, V., De Stefano, N., Leoni, V., and Mignarri, A. (2023). 24S-Hydroxycholesterol and Cerebellar Degeneration: Insights from SCA2. *Cerebellum* 22, 1020–1022. doi: 10.1007/s12311-022-01448-7
- Loix, M., Zelcer, N., Bogie, J. F. J., and Hendriks, J. J. A. (2024). The ubiquitous role of ubiquitination in lipid metabolism. *Trends Cell Biol.* 34, 416–429. doi: 10.1016/j.tcb.2023.09.001
- Lutz, B. M., and Peng, J. (2018). Deep profiling of the aggregated proteome in Alzheimer's Disease: from pathology to disease mechanisms. *Proteomes* 6:46. doi: 10.3390/proteomes6040046
- Ma, J. F., Wang, H. M., Li, Q. Y., Zhang, Y., Pan, J., Qiang, Q., et al. (2010). Starvation triggers Abeta42 generation from human umbilical vascular endothelial cells. *FEBS Lett.* 584, 3101–3106. doi: 10.1016/j.febslet.2010.05.048
- Maiuri, T., Mocle, A. J., HunG, C. L., Xia, J., Van Roon-Mom, W. M., and Truant, R. (2017). Huntingtin is a scaffolding protein in the ATM oxidative DNA damage response complex. *Hum. Mol. Genet.* 26, 395–406. doi: 10.1093/hmg/ddw395
- Marcelo, A., Afonso, I. T., Afonso-Reis, R., Brito, D. V. C., Costa, R. G., Rosa, A., et al. (2021). Autophagy in Spinocerebellar ataxia type 2, a dysregulated pathway, and a target for therapy. *Cell Death Dis.* 12:1117. doi: 10.1038/s41419-021-04404-1
- Martin, L., Latypova, X., Wilson, C. M., Magnaudeix, A., Perrin, M. L., Yardin, C., et al. (2013). Tau protein kinases: involvement in Alzheimer's disease. *Ageing Res. Rev.* 12, 289–309. doi: 10.1016/j.arr.2012.06.003
- Mawuenyega, K. G., Sigurdson, W., Ovod, V., Munsell, L., Kasten, T., Morris, J. C., et al. (2010). Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science* 330:1774. doi: 10.1126/science.1197623
- McLoughlin, H. S., Moore, L. R., and Paulson, H. L. (2020). Pathogenesis of SCA3 and implications for other polyglutamine diseases. *Neurobiol. Dis.* 134:104635. doi: 10.1016/j.nbd.2019.104635
- Medina, A., Mahjoub, Y., Shaver, L., and Pringsheim, T. (2022). Prevalence and Incidence of huntington's disease: an updated systematic review and meta-analysis. *Mov. Disord.* 37, 2327–2335. doi: 10.1002/mds.29228
- Meller, A., and Shalgi, R. (2021). The aging proteostasis decline: from nematode to human. *Exp. Cell Res.* 399:112474. doi: 10.1016/j.yexcr.2021.112474
- Mentis, A. A., Dardiotis, E., Efthymiou, V., and Chrousos, G. P. (2021). Non-genetic risk and protective factors and biomarkers for neurological disorders:

- a meta-umbrella systematic review of umbrella reviews. *BMC Med.* 19:6. doi: 10.1186/s12916-020-01873-7
- Menzies, F. M., Huebener, J., Renna, M., Bonin, M., Riess, O., and Rubinsztein, D. C. (2010). Autophagy induction reduces mutant ataxin-3 levels and toxicity in a mouse model of spinocerebellar ataxia type 3. *Brain* 133, 93–104. doi: 10.1093/brain/a wp292
- Mi, Y., Qi, G., Vitali, F., Shang, Y., Raikes, A. C., Wang, T., et al. (2023). Loss of fatty acid degradation by astrocytic mitochondria triggers neuroinflammation and neurodegeneration. *Nat. Metab.* 5, 445–465. doi: 10.1038/s42255-023-0 0756-4
- Monro, O. R., Mackic, J. B., Yamada, S., Segal, M. B., Ghiso, J., Maurer, C., et al. (2002). Substitution at codon 22 reduces clearance of Alzheimer's amyloid-beta peptide from the cerebrospinal fluid and prevents its transport from the central nervous system into blood. *Neurobiol. Aging* 23, 405–412. doi: 10.1016/s0197-4580(01)00317-7
- Na, H., Yang, J. B., Zhang, Z., Gan, Q., Tian, H., Rajab, I. M., et al. (2023). Peripheral apolipoprotein E proteins and their binding to LRP1 antagonize Alzheimer's disease pathogenesis in the brain during peripheral chronic inflammation. *Neurobiol. Aging* 127, 54–69. doi: 10.1016/j.neurobiolaging.2023.02.013
- Namba, Y., Tomonaga, M., Kawasaki, H., Otomo, E., and Ikeda, K. (1991). Apolipoprotein E immunoreactivity in cerebral amyloid deposits and neurofibrillary tangles in Alzheimer's disease and kuru plaque amyloid in Creutzfeldt-Jakob disease. *Brain Res.* 541, 163–166. doi: 10.1016/0006-8993(91)91092-f
- Nixon, R. A., and Rubinsztein, D. C. (2024). Mechanisms of autophagy-lysosome dysfunction in neurodegenerative diseases. *Nat. Rev. Mol. Cell Biol.* 25, 926–946. doi: 10.1038/s41580-024-00757-5
- Nixon, R. A., and Yang, D. S. (2011). Autophagy failure in Alzheimer's disease—locating the primary defect. *Neurobiol. Dis.* 43, 38–45. doi: 10.1016/j.nbd.2011.01.021
- Nóbrega, C., Mendonça, L., Marcelo, A., Lamazière, A., Tomé, S., Despres, G., et al. (2019). Restoring brain cholesterol turnover improves autophagy and has therapeutic potential in mouse models of spinocerebellar ataxia. *Acta Neuropathol.* 138, 837–858. doi: 10.1007/s00401-019-02019-7
- Onofre, I., Mendonça, N., Lopes, S., Nobre, R., De Melo, J. B., Carreira, I. M., et al. (2016). Fibroblasts of Machado Joseph Disease patients reveal autophagy impairment. *Sci. Rep.* 6:28220. doi: 10.1038/srep28220
- Orr, H. T., and Zoghbi, H. Y. (2007). Trinucleotide repeat disorders. *Annu. Rev. Neurosci.* 30, 575–621. doi: 10.1146/annurev.neuro.29.051605.113042
- Panas, M., Avramopoulos, D., Karadima, G., Petersen, M. B., and Vassilopoulos, D. (1999). Apolipoprotein E and presenilin-1 genotypes in Huntington's disease. *J. Neurol.* 246, 574–577. doi: 10.1007/s004150050406
- Pang, S., Li, J., Zhang, Y., and Chen, J. (2018). Meta-analysis of the relationship between the apoE gene and the onset of parkinson's disease dementia. *Parkinsons Dis.* 2018:9497147. doi: 10.1155/2018/9497147
- Parcon, P. A., Balasubramaniam, M., Ayyadevara, S., Jones, R. A., Liu, L., Shmookler Reis, R. J., et al. (2018). Apolipoprotein E4 inhibits autophagy gene products through direct, specific binding to CLEAR motifs. *Alzheimers. Dement.* 14, 230–242. doi: 10.1016/j.jalz.2017.07.754
- Paul, S., Dansithong, W., Figueroa, K. P., Scoles, D. R., and Pulst, S. M. (2018). Staufen1 links RNA stress granules and autophagy in a model of neurodegeneration. *Nat. Commun.* 9:3648. doi: 10.1038/s41467-018-06041-3
- Paulson, H. L., Shakkottai, V. G., Clark, H. B., and Orr, H. T. (2017). Polyglutamine spinocerebellar ataxias - from genes to potential treatments. *Nat. Rev. Neurosci.* 18, 613–626. doi: 10.1038/nrn.2017.92
- Peng, H., Wang, C., Chen, Z., Sun, Z., Jiao, B., Li, K., et al. (2014). APOE ε2 allele may decrease the age at onset in patients with spinocerebellar ataxia type 3 or Machado-Joseph disease from the Chinese Han population. *Neurobiol. Aging* 35, e15–e18. doi: 10.1016/j.neurobiolaging.2014.03.020
- Pereira Sena, P., Weber, J. J., Watchon, M., Robinson, K. J., Wassouf, Z., Hauser, S., et al. (2021). Pathophysiological interplay between O-GlcNAc transferase and the Machado-Joseph disease protein ataxin-3. *Proc. Natl. Acad. Sci. USA.* 118:e2025810118. doi: 10.1073/pnas.2025810118
- Pfeiffer, A., Luijsterburg, M. S., Acs, K., Wiegant, W. W., Helfrich, A., Herzog, L. K., et al. (2017). Ataxin-3 consolidates the MDC1-dependent DNA double-strand break response by counteracting the SUMO-targeted ubiquitin ligase RNF4. *EMBO J.* 36, 1066–1083. doi: 10.15252/embj.201695151
- Pihlström, L., Wiethoff, S., and Houlden, H. (2017). Genetics of neurodegenerative diseases: an overview. *Handb. Clin. Neurol.* 145, 309–323. doi: 10.1016/B978-0-12-802395-2.00022-5
- Poewe, W., Seppi, K., Tanner, C. M., Halliday, G. M., Brundin, P., Volkman, J., et al. (2017). Parkinson disease. *Nat. Rev. Dis. Primers* 3:17013. doi: 10.1038/nrdp.2017.13
- Pulst, S. M. (1993). Spinocerebellar ataxia type 2. In *GeneReviews*, eds. Adam, M. P., Feldman, J., Mirza, G. M., Pagon, R. A., Wallace, S. E. and Amemiya, A (Seattle, WA: University of Washington, Seattle)
- Putka, A. F., Mohanty, V., Cologna, S. M., and McLoughlin, H. S. (2025). Cerebellar lipid dysregulation in SCA3: a comparative study in patients and mice. *Neurobiol. Dis.* 206:106827. doi: 10.1016/j.nbd.2025.106827
- Qi, X. M., and Ma, J. F. (2017). The role of amyloid beta clearance in cerebral amyloid angiopathy: more potential therapeutic targets. *Transl. Neurodegener.* 6:22. doi: 10.1186/s40035-017-0091-7
- Rannikmäe, K., Samarasekera, N., Martínez-González, N. A., Al-Shahi Salman, R., and Sudlow, C. L. (2013). Genetics of cerebral amyloid angiopathy: systematic review and meta-analysis. *J. Neurol. Neurosurg. Psychiatr.* 84, 901–908. doi: 10.1136/jnnp-2012-303898
- Rao, G., Croft, B., Teng, C., and Awasthi, V. (2015). Ubiquitin-proteasome system in neurodegenerative disorders. *J. Drug Metab. Toxicol.* 6:187. doi: 10.4172/2157-7609.1000187
- Raposo, M., Bettencourt, C., Melo, A. R. V., Ferreira, A. F., Alonso, I., Silva, P., et al. (2022). Novel Machado-Joseph disease-modifying genes and pathways identified by whole-exome sequencing. *Neurobiol. Dis.* 162:105578. doi: 10.1016/j.nbd.2021.105578
- Raulin, A. C., Doss, S. V., Trottier, Z. A., Ikezu, T. C., Bu, G., and Liu, C. C. (2022). ApoE in Alzheimer's disease: pathophysiology and therapeutic strategies. *Mol. Neurodegener.* 17:72. doi: 10.1186/s13024-022-00574-4
- Ravikumar, B., Vacher, C., Berger, Z., Davies, J. E., Luo, S., Oroz, L. G., et al. (2004). Inhibition of mTOR induces autophagy and reduces toxicity of polyglutamine expansions in fly and mouse models of Huntington disease. *Nat. Genet.* 36, 585–595. doi: 10.1038/ng1362
- Rawat, P., Sehar, U., Bisht, J., Selman, A., Culbertson, J., and Reddy, P. H. (2022). Phosphorylated tau in Alzheimer's disease and other tauopathies. *Int. J. Mol. Sci.* 23:12841. doi: 10.3390/ijms232112841
- Reinert, J., Richard, B. C., Klafki, H. W., Friedrich, B., Bayer, T. A., Wiltfang, J., et al. (2016). Deposition of C-terminally truncated Aβ species Aβ37 and Aβ39 in Alzheimer's disease and transgenic mouse models. *Acta Neuropathol. Commun.* 4:24. doi: 10.1186/s40478-016-0294-7
- Riguet, N., Mahul-Mellier, A. L., Maharjan, N., Burtscher, J., Croisier, M., Knott, G., et al. (2021). Nuclear and cytoplasmic huntingtin inclusions exhibit distinct biochemical composition, interactome and ultrastructural properties. *Nat. Commun.* 12:579. doi: 10.1038/s41467-021-26684-z
- Rosas, H. D., Salat, D. H., Lee, S. Y., Zaleta, A. K., Pappu, V., Fischl, B., et al. (2008). Cerebral cortex and the clinical expression of Huntington's disease: complexity and heterogeneity. *Brain* 131, 1057–1068. doi: 10.1093/brain/awn025
- Ross, C. A., and Poirier, M. A. (2004). Protein aggregation and neurodegenerative disease. *Nat. Med.* 10, S10–S17. doi: 10.1038/nm1066
- Ross, C. A., and Tabrizi, S. J. (2011). Huntington's disease: from molecular pathogenesis to clinical treatment. *Lancet Neurol.* 10, 83–98. doi: 10.1016/S1474-4422(10)70245-3
- Rüb, U., Schöls, L., Paulson, H., Auburger, G., Kermer, P., Jen, J. C., et al. (2013). Clinical features, neurogenetics and neuropathology of the polyglutamine spinocerebellar ataxias type 1, 2, 3, 6 and 7. *Prog. Neurobiol.* 104, 38–66. doi: 10.1016/j.pneurobio.2013.01.001
- Saher, G., and Stumpf, S. K. (2015). Cholesterol in myelin biogenesis and hypomyelinating disorders. *Biochim. Biophys. Acta* 1851, 1083–1094. doi: 10.1016/j.bbalip.2015.02.010
- Saudou, F., and Humbert, S. (2016). The biology of Huntingtin. *Neuron* 91, 910–926. doi: 10.1016/j.neuron.2016.02.003
- Savar, S. M., Ma, B., Hone, E., Jahan, F., Markovic, S., Pedrini, S., et al. (2024). Fluid biomarkers in cerebral amyloid angiopathy. *Front. Neurosci.* 18:1347320. doi: 10.3389/fnins.2024.1347320
- Schulze, M., Sommer, A., Plötz, S., Farrell, M., Winner, B., Grosch, J., et al. (2018). Sporadic Parkinson's disease derived neuronal cells show disease-specific mRNA and small RNA signatures with abundant deregulation of piRNAs. *Acta Neuropathol. Commun.* 6:58. doi: 10.1186/s40478-018-0561-x
- Seidel, K., Siswanto, S., Friedrich, M., Bouzrou, M., Den Dunnen, W. F. A., Özden, I., et al. (2017). On the distribution of intranuclear and cytoplasmic aggregates in the brainstem of patients with spinocerebellar ataxia type 2 and 3. *Brain Pathol.* 27, 345–355. doi: 10.1111/bpa.12412
- Sen, N. E., Arsovic, A., Meierhofer, D., Brodesser, S., Oberschmidt, C., Canet-Pons, J., et al. (2019). In human and mouse spinocerebellar tissue, ataxin-2 expansion affects ceramide-sphingomyelin metabolism. *Int. J. Mol. Sci.* 20:5854. doi: 10.3390/ijms20235854
- Serrano-Pozo, A., Das, S., and Hyman, B. T. (2021). APOE and Alzheimer's disease: advances in genetics, pathophysiology, and therapeutic approaches. *Lancet Neurol.* 20, 68–80. doi: 10.1016/S1474-4422(20)30412-9
- Shahmoradian, S. H., Lewis, A. J., Genoud, C., Hench, J., Moors, T. E., Navarro, P. P., et al. (2019). Lewy pathology in Parkinson's disease consists of crowded organelles and lipid membranes. *Nat. Neurosci.* 22, 1099–1109. doi: 10.1038/s41593-019-0423-1
- Shinohara, M., Tachibana, M., Kanekiyo, T., and Bu, G. (2017). Role of LRP1 in the pathogenesis of Alzheimer's disease: evidence from clinical and preclinical studies. *J. Lipid Res.* 58, 1267–1281. doi: 10.1194/jlr.R075796
- Sienski, G., Narayan, P., Bonner, J. M., Kory, N., Boland, S., Arczewska, A. A., et al. (2021). APOE4 disrupts intracellular lipid homeostasis in human iPSC-derived glia. *Sci. Transl. Med.* 13:eaa4564. doi: 10.1126/scitranslmed.aaz4564

- Sinha, S., Iyer, D., and Granata, A. (2014). Embryonic origins of human vascular smooth muscle cells: implications for in vitro modeling and clinical application. *Cell. Mol. Life Sci.* 71, 2271–2288. doi: 10.1007/s00018-013-1554-3
- Stewart, K. L., and Radford, S. E. (2017). Amyloid plaques beyond A β : a survey of the diverse modulators of amyloid aggregation. *Biophys. Rev.* 9, 405–419. doi: 10.1007/s12551-017-0271-9
- Stonebraker, A. R., Beasley, M., Massinople, S., Wunder, M., Li, P., Valentine, S. J., et al. (2023). Cholesterol impacts the formation of huntingtin/lipid complexes and subsequent aggregation. *Protein Sci.* 32:e4642. doi: 10.1002/pro.4642
- Strickland, M. R., and Holtzman, D. M. (2019). Dr. Jekyll and Mr. Hyde: APOE explains opposing effects of neuronal LRP1. *J. Clin. Invest.* 129, 969–971. doi: 10.1172/JCI127578
- Strittmatter, W. J., Saunders, A. M., Schmechel, D., Pericak-Vance, M., Enghild, J., Salvesen, G. S., et al. (1993). Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. USA.* 90, 1977–1981. doi: 10.1073/pnas.90.5.1977
- Swarup, V., Srivastava, A. K., Padma, M. V., and Moganty, R. R. (2013). Quantitative profiling and identification of plasma proteins of spinocerebellar ataxia type 2 patients. *Neurodegener. Dis.* 12, 199–206. doi: 10.1159/000346585
- Tachibana, M., Holm, M. L., Liu, C. C., Shinohara, M., Aikawa, T., Oue, H., et al. (2019). APOE4-mediated amyloid- β pathology depends on its neuronal receptor LRP1. *J. Clin. Invest.* 129, 1272–1277. doi: 10.1172/JCI124853
- Tandon, S., Aggarwal, P., and Sarkar, S. (2024). Polyglutamine disorders: pathogenesis and potential drug interventions. *Life Sci.* 344:122562. doi: 10.1016/j.lfs.2024.122562
- Tenchov, R., Sasso, J. M., and Zhou, Q. A. (2024). Polyglutamine (PolyQ) diseases: navigating the landscape of neurodegeneration. *ACS Chem. Neurosci.* 15, 2665–2694. doi: 10.1021/acschemneuro.4c00184
- Therriault, J., Benedet, A. L., Pascoal, T. A., Mathotaarachchi, S., Chamoun, M., Savard, M., et al. (2020). Association of apolipoprotein E ϵ 4 with medial temporal tau independent of amyloid- β . *JAMA Neurol.* 77, 470–479. doi: 10.1001/jamaneurol.2019.4421
- Tojima, M., Murakami, G., Hikawa, R., Yamakado, H., Yamashita, H., Takahashi, R., et al. (2018). Homozygous 31 trinucleotide repeats in the SCA2 allele are pathogenic for cerebellar ataxia. *Neurol. Genet.* 4:e283. doi: 10.1212/NXG.0000000000000283
- Toonen, L. J. A., Overzier, M., Evers, M. M., Leon, L. G., Van Der Zeeuw, S. A. J., Mei, H., et al. (2018). Transcriptional profiling and biomarker identification reveal tissue specific effects of expanded ataxin-3 in a spinocerebellar ataxia type 3 mouse model. *Mol. Neurodegener.* 13:31. doi: 10.1186/s13024-018-0261-9
- Valenza, M., Leoni, V., Karasinska, J. M., Petricca, L., Fan, J., Carroll, J., et al. (2010). Cholesterol defect is marked across multiple rodent models of Huntington's disease and is manifest in astrocytes. *J. Neurosci.* 30, 10844–10850. doi: 10.1523/JNEUROSCI.0917-10.2010
- Valenza, M., Marullo, M., Di Paolo, E., Cesana, E., Zuccato, C., Biella, G., et al. (2015). Disruption of astrocyte-neuron cholesterol cross talk affects neuronal function in Huntington's disease. *Cell Death Differ.* 22, 690–702. doi: 10.1038/cdd.2014.162
- Van Acker, Z. P., Bretou, M., and Annaert, W. (2019). Endo-lysosomal dysregulations and late-onset Alzheimer's disease: impact of genetic risk factors. *Mol. Neurodegener.* 14:20. doi: 10.1186/s13024-019-0323-7
- van den Berg, E., Kersten, I., Brinkmalm, G., Johansson, K., De Kort, A. M., Klijn, C. J. M., et al. (2024). Profiling amyloid- β peptides as biomarkers for cerebral amyloid angiopathy. *J. Neurochem.* 168, 1254–1264. doi: 10.1111/jnc.16074
- Wardman, J. H., Henriksen, E. E., Marthaler, A. G., Nielsen, J. E., and Nielsen, T. T. (2020). Enhancement of autophagy and solubilization of ataxin-2 alleviate apoptosis in spinocerebellar ataxia Type 2 patient cells. *Cerebellum* 19, 165–181. doi: 10.1007/s12311-019-01092-8
- Weber, J. J., Clemensson, L. E., Schiöth, H. B., and Nguyen, H. P. (2019). Olesoxime in neurodegenerative diseases: Scrutinising a promising drug candidate. *Biochem. Pharmacol.* 168, 305–318. doi: 10.1016/j.bcp.2019.07.002
- Weber, J. J., Czisch, L., Pereira Sena, P., Fath, F., Huridou, C., Schwarz, N., et al. (2024). The parkin V380L variant is a genetic modifier of Machado-Joseph disease with impact on mitophagy. *Acta Neuropathol.* 148:14. doi: 10.1007/s00401-024-02762-6
- Williams, A., Sarkar, S., Cuddon, P., Ttoli, E. K., Saiki, S., Siddiqi, F. H., et al. (2008). Novel targets for Huntington's disease in an mTOR-independent autophagy pathway. *Nat. Chem. Biol.* 4, 295–305. doi: 10.1038/nchembio.79
- Wilson, D. M., 3R. D., Cookson, M. R., Van Den Bosch, L., Zetterberg, H., et al. (2023). Hallmarks of neurodegenerative diseases. *Cell* 186, 693–714. doi: 10.1016/j.cell.2022.12.032
- Windham, I. A., and Cohen, S. (2024). The cell biology of APOE in the brain. *Trends Cell Biol.* 34, 338–348. doi: 10.1016/j.tcb.2023.09.004
- Wolozin, B., and Ivanov, P. (2019). Stress granules and neurodegeneration. *Nat. Rev. Neurosci.* 20, 649–666. doi: 10.1038/s41583-019-0222-5
- Xia, Z., Prescott, E. E., Urbanek, A., Wareing, H. E., King, M. C., Olerinyova, A., et al. (2024). Co-aggregation with apolipoprotein E modulates the function of amyloid- β in Alzheimer's disease. *Nat. Commun.* 15:4695. doi: 10.1038/s41467-024-49028-z
- Xie, Y., Li, J., Kang, R., and Tang, D. (2020). Interplay between lipid metabolism and autophagy. *Front. Cell Dev. Biol.* 8:431. doi: 10.3389/fcell.2020.00431
- Yamada, M. (2015). Cerebral amyloid angiopathy: emerging concepts. *J. Stroke* 17, 17–30. doi: 10.5853/jos.2015.17.1.17
- Yang, H., Li, J. J., Liu, S., Zhao, J., Jiang, Y. J., Song, A. X., et al. (2014). Aggregation of polyglutamine-expanded ataxin-3 sequesters its specific interacting partners into inclusions: implication in a loss-of-function pathology. *Sci. Rep.* 4:6410. doi: 10.1038/srep06410
- Yang, L. G., March, Z. M., Stephenson, R. A., and Narayan, P. S. (2023). Apolipoprotein E in lipid metabolism and neurodegenerative disease. *Trends Endocrinol. Metab.* 34, 430–445. doi: 10.1016/j.tem.2023.05.0
- Yang, Q., Zeng, X., Tang, L., Liu, X., Xia, K., Gao, F., et al. (2025). Association of APOE genotype with CT markers of cerebral amyloid angiopathy in spontaneous intracerebral haemorrhage. *Stroke. Vasc. Neurol.* 10:e003477. doi: 10.1136/svn-2024-003477
- Yerbury, J. J., Ooi, L., Dillin, A., Saunders, D. N., Hatters, D. M., Beart, P. M., et al. (2016). Walking the tightrope: proteostasis and neurodegenerative disease. *J. Neurochem.* 137, 489–505. doi: 10.1111/jnc.13575
- Zhang, J., Zhang, Y., Wang, J., Xia, Y., Zhang, J., and Chen, L. (2024). Recent advances in Alzheimer's disease: Mechanisms, clinical trials and new drug development strategies. *Signal Transduct. Target Ther.* 9:211. doi: 10.1038/s41392-024-01911-3
- Zhao, J., and Huai, J. (2023). Role of primary aging hallmarks in Alzheimer's disease. *Theranostics* 13, 197–230. doi: 10.7150/thno.79535
- Zhou, Q., Ni, W., Dong, Y., Wang, N., Gan, S. R., and Wu, Z. Y. (2014). The role of apolipoprotein E as a risk factor for an earlier age at onset for Machado-Joseph disease is doubtful. *PLoS ONE* 9:e111356. doi: 10.1371/journal.pone.0111356