

OPEN ACCESS

EDITED BY Robert John Vandenberg, The University of Sydney, Australia

REVIEWED BY
Nitish Mittal,
CereNexis Ventures, United States
Ankit Uniyal,
Johns Hopkins University, United States

*CORRESPONDENCE
Jing-Yan Lin

☑ linjingyan@nsmc.edu.cn

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 15 June 2025 ACCEPTED 07 October 2025 PUBLISHED 30 October 2025

CITATION

Jia SY, Tang X-J, Su Y, Liu Y-N, Ming Z and Lin J-Y (2025) The role and mechanisms of matrix metalloproteinase-9 in peripheral neuropathic pain. Front. Mol. Neurosci. 18:1647316. doi: 10.3389/fnmol.2025.1647316

COPYRIGHT

© 2025 Jia, Tang, Su, Liu, Ming and Lin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The role and mechanisms of matrix metalloproteinase-9 in peripheral neuropathic pain

Shuai Ying Jia[†], Xiao-Jun Tang[†], Yao Su, Yu-Ning Liu, Zhi Ming and Jing-Yan Lin*

Department of Anesthesiology, The Affiliated Hospital of North Sichuan Medical College, Nanchong, China

Peripheral neuropathic pain is a chronic, secondary pain state caused by damage or diseases of the peripheral nervous system, typically accompanied by edema, inflammatory responses, increased neuronal excitability, and glutamate accumulation. Matrix metalloproteinase-9 (MMP-9), an important enzyme, plays a key role in various physiological and pathological processes, primarily by degrading the extracellular matrix. Recent studies have shown that MMP-9 plays a crucial role in the onset and progression of central nervous system disorders, particularly neuropathic pain. This review discusses the mechanisms underlying the involvement of MMP-9 in various models of peripheral neuropathic pain, with the aim of exploring its potential as a therapeutic target.

KEYWORDS

matrix metalloproteinase-9, peripheral neuropathic pain, nerve injury, pathophysiology, treatment strategy

1 Introduction

Neuropathic pain is a complex condition that results from damage or dysfunction within the somatosensory nervous system. It affects about 7–10% of people globally and is particularly common among middle-aged individuals and women (van Hecke et al., 2014). There are two main types of neuropathic pain based on the underlying cause: central and peripheral. Central neuropathic pain arises from damage to the brain or spinal cord, while peripheral neuropathic pain is caused by injury to the peripheral nervous system (Scholz et al., 2019). Peripheral neuropathic pain is characterized by a wide range of clinical symptoms and a complex pathophysiology. These mechanisms are distinct from the pain caused by tissue damage or disease. Common symptoms include radicular pain, burning sensations, tingling, and numbness (Meacham et al., 2017). In addition to persistent pain, patients often experience sleep disturbances, depression, anxiety, and significant difficulties in daily life. These factors collectively reduce the overall quality of life for individuals suffering from the condition (Smith and Torrance, 2012). Furthermore, peripheral neuropathic pain often does not respond well to standard analgesic treatments, which highlights the urgent need for more effective therapeutic strategies (Finnerup et al., 2021).

Matrix metalloproteinase-9 (MMP-9), also known as gelatinase B, is a zinc-dependent endopeptidase in the matrix metalloproteinase family. MMP-9 regulates interactions between cells and the extracellular matrix by breaking down its components. This process influences cell migration, inflammation, proliferation, and apoptosis (Cabral-Pacheco et al., 2020). During inflammatory responses, MMP-9 plays a key role in the migration and activation of leukocytes. Recent studies have highlighted its crucial involvement in the development and progression of several neuropathic conditions, including trigeminal neuralgia, diabetic neuropathy, postherpetic neuralgia, and chronic neuropathic pain following peripheral nerve

injury (Yin et al., 2019; Singh et al., 2020; Pan et al., 2018; Fan et al., 2018). Elevated MMP-9 activity is closely linked to post-injury inflammation, neuron–glia interactions, and pain signal transmission (Umbricht, 2022; Huang et al., 2021; Remacle et al., 2015). Consequently, MMP-9 has emerged as a key therapeutic target in neuropathic pain research.

MMP-9 is encoded by the gelatinase B gene located on human chromosome 20. This gene consists of 13 exons and 12 introns. The protein encoded by MMP-9 has a complex structure, including a signal peptide, propeptide, hinge region, catalytic domain, and a hemopexin-like domain (Nagase et al., 2006). The catalytic domain contains a fibronectin type II domain, an active site, and a zinc-binding region that relies on zinc ions for its catalytic function. MMP-9's primary role is the degradation of the extracellular matrix (ECM) (Agrawal et al., 2008). Its structure is shown in Figure 1. Additionally, MMP-9 is involved in important biological processes such as tissue remodeling, wound healing, skeletal development, and immune responses (Qorri et al., 2018; Hingorani et al., 2018; Malemud, 2006; Bratcher et al., 2012; McClellan et al., 2006).

The expression of MMP-9 is regulated at multiple levels, including gene transcription and post-translational modifications. At the transcriptional level, transcription factors like activator protein 1 (AP-1) and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) regulate MMP-9 expression in response to external stimuli such as growth factors, cytokines, and stress signals (Wang T. et al., 2018; Li et al., 2020; Murthy et al., 2010). Epigenetic modifications, including DNA methylation and histone acetylation, also play a significant role in regulating MMP-9 expression (Duraisamy et al., 2017; Yeh et al., 2016). At the post-translational level, MMP-9 is secreted as an inactive proenzyme (Pro-MMP-9) and is activated through cleavage by specific proteases. Membrane-type matrix metalloproteinase removes the inhibitory sequence from the proenzyme, thus activating its catalytic function (Dreier et al., 2004). Furthermore, the activity of MMP-9 is tightly controlled by tissue inhibitors of metalloproteinases (TIMPs), which bind to MMP-9 and prevent its interaction with substrates. This regulation is critical for maintaining the integrity of the extracellular matrix and preventing excessive degradation (Filanti et al., 2000).

2 Role of MMP-9 in different types of peripheral neuropathic pain

According to the latest classification system by the International Association for the Study of Pain (IASP), this paper reviews several

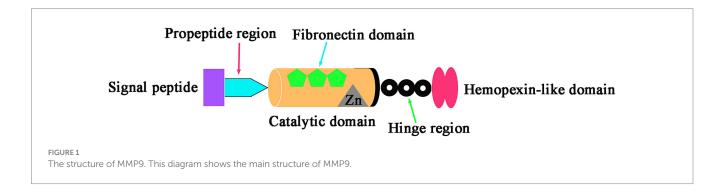
common subtypes of peripheral neuropathic pain. These include trigeminal neuralgia, chronic neuropathic pain after peripheral nerve injury, painful polyneuropathy, postherpetic neuralgia, and painful radiculopathy (Treede et al., 2019).

2.1 Trigeminal neuralgia

Trigeminal neuralgia (TN) is a well-known condition characterized by severe pain along the trigeminal nerve. It is typically categorized into three subtypes based on its cause: classical, secondary, and idiopathic (Anonymous, 2018). The main symptom of TN is sudden, sharp facial pain on one side of the face, often affecting one or more branches of the trigeminal nerve (Edlich et al., 2006). Patients often describe the pain as sharp, electric shock-like, or stabbing. Episodes can last from a few seconds to a few minutes. Pain is commonly triggered by mild stimuli such as light facial touch, chewing, or even exposure to wind. In some cases, patients may also experience involuntary facial muscle contractions or spasms (Jones et al., 2019; Bendtsen et al., 2020). Epidemiological studies report an annual incidence of TN ranging from 4.3 to 27 cases per 100,000 individuals. It is more common in people over 60 years old and more prevalent in women, with an incidence rate of 5.9 per 100,000 compared to 3.4 per 100,000 in men (Shaefer et al., 2018).

In classical trigeminal neuralgia, the vascular compression hypothesis is the most widely accepted explanation (Anonymous, 2018). Approximately 50% of cases are attributed to compression by the superior cerebellar artery, while about 25% are caused by venous compression (Hamlyn and King, 1992). Vascular compression, often due to abnormal dilation of intracranial arteries or veins, leads to demyelination of the trigeminal nerve root. This results in abnormal nerve signaling and pain (Marinković et al., 2009). Moreover, the abnormal activation of specific sodium and potassium channels on nerve membranes contributes to neuronal hyperexcitability and ectopic discharges, key mechanisms in the pathophysiology of TN (Liu et al., 2019; Ling et al., 2018). Secondary trigeminal neuralgia is usually associated with facial trauma or surgeries involving the trigeminal nerve or its branches (Renton et al., 2010). Other factors, such as genetic predisposition, infections like postherpetic neuralgia, and autoimmune disorders, can impair trigeminal nerve function and trigger or worsen pain (Fernández Rodríguez et al., 2019; Wang et al., 2023; Ferreira et al., 2020).

The main treatment for TN is pharmacological, with carbamazepine and other anticonvulsants being the first-line options. Botulinum toxin and local anesthetics have also been found to provide



quick and effective pain relief (Kowacs et al., 2015; Han et al., 2008). More recently, novel sodium channel blockers have shown promising results in clinical trials (Zakrzewska et al., 2017; Dunbar et al., 2020). For patients who do not respond to medication, surgical options such as microvascular decompression and nerve block procedures can be effective and may be considered either as first-line or secondary treatments following recurrence (Xia et al., 2014; Wang et al., 2022).

Recent studies using chronic constriction injury (CCI) models in CD-1 mice have shown that MMP-9 and MMP-2 expression increases significantly in the trigeminal ganglion at different time points. This suggests that MMP-9, in particular, may play a crucial role in the onset and progression of trigeminal neuralgia (Yin et al., 2019). In models of spinal nerve ligation (SNL) and infraorbital nerve CCI (CCI-IoN), blocking MMP-9/2 activity significantly reduces mechanical allodynia. This indicates that targeting MMP-9/2 could be a promising strategy for pain relief (Henry et al., 2015). Similarly, in a rat model of temporomandibular joint arthritis, mechanical allodynia of the trigeminal nerve was observed. This was accompanied by increased MMP-9 expression and activity in both the limbic system and trigeminal ganglion. These changes may be linked to a reduction in immunoreactivity of the voltage-gated K + channel subtype 1.4 in trigeminal ganglion neurons (Takeda et al., 2008; Nascimento et al., 2021). Low-level laser therapy (LLLT) has been shown to reduce MMP-9 levels in the trigeminal ganglion, decrease its gelatinolytic activity, and alleviate both mechanical allodynia and orofacial hyperalgesia. These results suggest that LLLT may aid in tissue repair and limit extracellular matrix degradation (Desiderá et al., 2015; Lemos et al., 2016). Taken together, these studies underline the central role of MMP-9 in the development of trigeminal neuralgia and provide a strong foundation for exploring new treatment options, with MMP-9 inhibition emerging as a particularly promising approach.

2.2 Neuropathic pain following peripheral nerve injury

Peripheral nerve injury causes both structural damage and functional impairments, often accompanied by significant neuropathic pain (Lopes et al., 2022). In response to injury, the affected nerve region rapidly triggers an inflammatory reaction. This reaction recruits immune cells, including macrophages, neutrophils, and T-cells (Balog et al., 2023; Moalem et al., 2004). These immune cells release proinflammatory mediators such as tumor necrosis factoralpha (TNF- α), interleukin-1 beta (IL-1 β), interleukin-6 (IL-6), and prostaglandins. These mediators amplify inflammation and increase nerve sensitivity. As a result, pain intensity increases and persists (Nadeau et al., 2011; Gui et al., 2016).

Recent studies have shown that MMP-9 plays a key role in the development of neuropathic pain. MMP-9 influences the function of the voltage-gated sodium channel subunit Nav1.7, which contributes to pain onset. It also interacts with N-methyl-D-aspartate receptors (NMDARs), which regulate synaptic plasticity and memory formation (Remacle et al., 2015; Wiera et al., 2017). Specifically, during neurodevelopment, MMP-9 accelerates the functional inactivation of the Nav1.7 sodium channel by hydrolyzing its external exposed region, particularly the structural region between the S5-S6 transmembrane segments. MMP-9 may also affect neuronal conduction by modulating the extracellular matrix and relevant

signaling pathways. For instance, MMP-9 might influence local neuronal conduction properties through its actions on the extracellular matrix. Additionally, MMP-9 activity may interact with inflammatory response pathways, such as the NF-kB pathway. This interaction further exacerbates plasticity changes in the nervous system, contributing to pain regulation and other neurodegenerative diseases (Hackel et al., 2012; Remacle et al., 2015; Wang et al., 2019). In various pain models, MMP-9 has been shown to influence NMDA receptors. For example, in the bone cancer pain model, the upregulation of MMP-9 activates Ephrin type-B receptor 1. This activation enhances the phosphorylation of NMDA receptor subunits NR1 and NR2B, leading to increased Ca2+ influx. This amplifies downstream signaling, promoting hyperalgesia and opioid tolerance (Liu et al., 2011). In a mouse plantar incision model, MMP-9 activity significantly increases in the spinal cord and glial cells. This activity contributes to postoperative hyperalgesia through the p38/IL-1β signaling pathway (Li et al., 2023; Jiang et al., 2020; Gu et al., 2019). Similarly, during the early stages of chronic sciatic nerve constriction injury in rats, MMP-9 activity is strongly associated with elevated levels of the chemokine C-X3-C motif ligand 1 (CX3CL1) (Zhao et al., 2020). In the sciatic nerve crush (SNC) rat model, overexpression of MMP-9 is linked to excessive activation of the TRPV1 channel, intensifying pain perception and inflammatory responses (Awad-Igbaria et al., 2023). Inhibition of MMP-9 activity or expression has been shown to effectively alleviate both postoperative and neuropathic pain (Lv et al., 2018; Liu et al., 2017). These findings emphasize the central role of MMP-9 in nerve injury and pain progression. Further research into MMP-9-mediated mechanisms may offer valuable insights into the molecular basis of pain and pave the way for developing novel therapeutic strategies.

2.3 Painful polyneuropathy

Painful polyneuropathy is generally classified into two main types: diabetic and non-diabetic (Scholz et al., 2019). Among these, painful diabetic neuropathy (PDN) is the most common, affecting about 20-30% of people with diabetes (Abbott et al., 2011; Aronson et al., 2021). Patients with PDN often report sharp, electric shock-like pain in their feet. This pain can become chronic and significantly lower their quality of life. In many cases, PDN is accompanied by psychological issues such as anxiety, depression, and sleep problems (Gylfadottir et al., 2020). Although the exact biological mechanisms behind PDN remain unclear, treatment primarily focuses on symptom relief. Common drugs include tricyclic antidepressants (TCAs), duloxetine, pregabalin, and gabapentin (Javed et al., 2015; Iqbal et al., 2018). However, these medications often cause side effects like nausea, drowsiness, and constipation. These adverse effects can reduce patient compliance and overall satisfaction with treatment (Tesfaye et al., 2022). Among non-diabetic causes of painful polyneuropathy, chemotherapy-induced peripheral neuropathy (CIPN) is especially common. It usually develops after treatment with chemotherapy agents such as paclitaxel or cisplatin. Patients with CIPN often experience persistent pain, numbness, and increased sensitivity to temperature or touch. In severe cases, these symptoms greatly impact daily functioning and quality of life (Hu et al., 2019).

PDN involves a complex interplay of multiple biological factors. Research has shown the involvement of pro-inflammatory cytokines

(such as TNF- α and IL-1 β), ion channels in sensory neurons, T-type calcium channels, as well as microglia and astrocytes (Yamakawa et al., 2011; Hangping et al., 2020; Bierhaus et al., 2012; Orestes et al., 2013; Liao et al., 2011). Additionally, the accumulation of harmful metabolic byproducts—such as reactive oxygen species (ROS), inflammatory transcription factors, and glutamate—can further drive the progression of PDN (Rivera-Aponte et al., 2015; Rendra et al., 2019). Due to the incomplete understanding of these mechanisms, no standardized treatment for PDN has been established. CIPN shares many of the same biological triggers. These include damage to intraepidermal nerve fibers (IENF), oxidative stress, ion channel activation, cytokine upregulation, and neuroimmune responses. Current treatments mainly rely on anti-inflammatory and pain-relief strategies to ease symptoms and improve patient comfort (Koskinen et al., 2011; Butturini et al., 2013; Zhang and Dougherty, 2014).

In preclinical studies, diabetic neuropathy models induced by streptozotocin in rats show heightened pain sensitivity and altered pain signaling. These models are widely used to explore PDN mechanisms. Treatments targeting oxidative-nitrosative stress, inflammatory cytokines, and MMP-9 activity have demonstrated significant reductions in neuropathic symptoms (Singh et al., 2020). More recent findings have highlighted a novel role for MMP-9 in cleaving beta-dystroglycan (β -DG). Inhibiting MMP-9 also affects the expression and localization of aquaporin-4 (AQP4) in the spinal cord's glymphatic system. This modulation may enhance the clearance of metabolic waste in the central nervous system and offer a new therapeutic pathway for PDN (Li et al., 2024). Furthermore, network pharmacology studies have identified MMP-9 as a potential molecular target of Dolastatin 16, a compound involved in the TNF signaling pathway. Dolastatin 16 is associated with both diabetic foot ulcers and PDN. As a potential MMP-9 inhibitor, it may also promote wound healing in diabetic patients (Luthfiana and Utomo, 2023). Together, these findings suggest that MMP-9 is not only involved in the pathophysiology of PDN but also represents a promising target for future treatment strategies.

Recent clinical and laboratory studies have also drawn attention to the role of MMP-9 in CIPN. In patients receiving chemotherapy, higher levels of MMP-9 and high-mobility group box 1 (HMGB1) in the blood have been linked to more severe neuropathic symptoms (Yang et al., 2023). In animal models, increased MMP-9 expression in dorsal root ganglion (DRG) neurons has been associated with pain sensitivity, oxidative damage, and inflammation. Mechanistically, MMP-9 contributes to extracellular matrix remodeling, promotes neuroinflammation, and worsens neuronal injury through pathways like the HMGB1-TLR4-PI3K/Akt axis (Gu et al., 2020). Blocking MMP-9—either through gene knockout or pharmacological inhibition—has been shown to reduce CIPN severity in animal models. This evidence points to MMP-9 as a viable therapeutic target. Several strategies to inhibit MMP-9 have shown promise. These include monoclonal antibodies targeting MMP-9 and small-molecule inhibitors like N-acetylcysteine (NAC), which have successfully reduced symptoms in preclinical models (Tonello et al., 2019). In addition, chemotherapy drugs such as cisplatin can induce cellular senescence in neurons. This senescence process is often marked by increased MMP-9 expression and the release of inflammatory molecules—a phenomenon known as the senescence-associated secretory phenotype (SASP) (Acklin et al., 2020; Saleh et al., 2024). Altogether, these results underline the central role of MMP-9 in both the onset and progression of CIPN. Targeting MMP-9 could open new avenues for improving outcomes in cancer patients affected by this challenging condition.

2.4 Painful radiculopathy

Painful radiculopathy is typically caused by lesions in the cervical, thoracic, lumbar, or sacral nerve roots. Low back pain is the most common symptom associated with this condition(Scholz et al., 2019). Intervertebral disk herniation is a leading cause of low back pain, affecting more than 70% of individuals and often worsening with age (Hoy et al., 2010). In addition to severe discomfort, low back pain can limit daily activities and lead to occupational disability. This imposes a significant burden on patients, their families, and society (Will et al., 2018). While many patients with acute low back pain recover without long-term symptoms or functional impairment through conservative treatments, such as education, pharmacological therapy, and physical rehabilitation, about 30% experience a recurrence of pain within 1 year. In some cases, this can progress to chronic low back pain (da Silva et al., 2017).

The underlying mechanisms of chronic low back pain remain debated. It is generally believed that its onset is closely linked to mechanical compression of nerve roots due to intervertebral disk herniation, accompanied by local inflammatory responses (Erbüyün et al., 2018). Chronic low back pain is often triggered by a combination of neuropathic and nociceptive pain mechanisms (Freynhagen et al., 2006; Kaki et al., 2005). Furthermore, intervertebral disk degeneration is considered a key factor contributing to low back pain (Peng, 2013). As individuals age, cellular senescence and phenotypic changes, alongside dysfunction of the extracellular matrix (ECM), lead to early degenerative changes in the intervertebral disks. These changes, in turn, trigger inflammatory responses that contribute to the progression of pain (Bermudez-Lekerika et al., 2022; Lyu et al., 2021).

Current clinical guidelines recommend maintaining physical activity and engaging in regular exercise to manage chronic low back pain. Analgesics can be used when necessary. Nonsteroidal anti-inflammatory drugs (NSAIDs) are among the most commonly prescribed treatments. Short-term use of NSAIDs has been shown to alleviate pain effectively (Piccoliori et al., 2013). However, despite their analgesic and anti-inflammatory effects, NSAIDs can cause gastrointestinal and cardiovascular side effects (Sostres et al., 2013; Wehling, 2014). Because chronic low back pain often involves both neuropathic and nociceptive components, combining different types of analgesics is a rational approach to treatment. Personalized combination therapy can improve analgesic efficacy while minimizing the risk of side effects by reducing the dosages of individual medications (Müller-Schwefe et al., 2017).

For patients with chronic low back pain due to identifiable causes, such as nerve root compression from intervertebral disk herniation or spinal stenosis, surgery may be considered, especially in cases of severe pain. Lumbar fusion surgery has been shown to significantly reduce pain and improve functional outcomes (Helm Ii et al., 2012; Trescot et al., 2007). Some studies suggest that lumbar fusion surgery may influence MMP-9 activity by altering the mechanical load on the intervertebral disks or modulating the local inflammatory environment (Omair et al., 2013; Zhang et al., 2018). However, despite various surgical options, a thorough preoperative evaluation is

essential (Chopko et al., 2014). Recent evidence suggests that long-term outcomes of surgical and non-surgical treatments are similar, highlighting the importance of carefully weighing the risks and high costs associated with spinal surgery (Mannion et al., 2013). Therefore, a comprehensive multidisciplinary rehabilitation program, combining pharmacological treatment, physical rehabilitation, psychological support, and addressing social and occupational factors, is recommended for optimal outcomes in patients with chronic low back pain (Guzmán et al., 2001).

Intervertebral disk degeneration (IDD) is characterized by cellular loss, ECM degradation, and reduced spinal flexibility (Yabe et al., 2015). Previous studies suggest that slowing ECM degradation may help delay disk degeneration (Gao et al., 2024). MMP-9, a key enzyme involved in ECM degradation, plays an important role in IDD, making it a potential therapeutic target. Research has shown that MMP-9 expression is upregulated in degenerated intervertebral disks (Wang W. J. et al., 2018). Inhibition of MMP-9 expression-via siRNAmediated gene silencing, small molecule inhibitors (e.g., SB-3CT), or modulation through upstream signaling pathways such as miR-21/ PTEN/Akt/mTOR—has been shown to reduce ECM breakdown and inflammatory responses in vitro NP cell models and in vivo rat models of IDD (Li et al., 2017; Gong et al., 2023; Shin et al., 2016). These findings suggest that controlling MMP-9 activity may slow the progression of disk degeneration. Additionally, genetic polymorphisms of MMP-9 have been identified as independent predictive factors for chronic low back pain and may influence patient recovery (Bjorland et al., 2019; Bjorland et al., 2017). MMP-9 also plays a significant role in low back pain associated with pregnancy, likely due to changes in the pelvic ligament ECM components caused by the rs17576 A > G polymorphism in the MMP-9 gene (Mahmood et al., 2018). While MMP-9 shows potential as a therapeutic target, its clinical feasibility is still under investigation. The development of selective MMP-9 inhibitors suitable for clinical use has been limited by issues such as off-target effects and poor bioavailability. Further studies using animal models and clinical samples are needed to assess the long-term efficacy and safety of these inhibitors (Kamieniak et al., 2019) (Table 1; Figure 2).

3 Discussion

Peripheral neuropathic pain places a heavy burden on both the body and mind. While current treatments can relieve some symptoms, they often fall short due to limited effectiveness, drug resistance, or side effects. As a result, there is an urgent need to uncover new molecular mechanisms and develop more targeted therapies. Among the molecules involved, matrix metalloproteinase-9 (MMP-9) has drawn increasing attention over the past decades. It is known for its key role in neuroinflammation and nerve remodeling in neuropathic pain.

MMP-9 contributes to pain progression by degrading extracellular matrix (ECM) components. This breakdown facilitates the infiltration of inflammatory cells and enhances its own expression, creating a feedback loop that worsens pain (Adams et al., 2015; Kwan et al., 2019). In addition, MMP-9 influences neuron–glia interactions and promotes nerve regeneration and axon growth, which are critical for recovery after nerve injury (Kim et al., 2012; Lu et al., 2022). It also indirectly affects how pain signals are transmitted by regulating

cytokines and chemokines. For example, MMP-9 can degrade nerve growth factor (NGF) released by glial cells, which in turn affects pain sensitivity (Osikowicz et al., 2013). Furthermore, MMP-9 increases the permeability of the blood-brain barrier, allowing inflammatory molecules to enter the central nervous system and activate central pain pathways (Wang et al., 2024). The role of MMP-9 differs across pain types. In mechanical allodynia, it activates the NLRP3 inflammasome, which promotes the release of IL-1 β and triggers a strong inflammatory response in the central nervous system (Deng et al., 2021). An imbalance between MMP-9 and its inhibitor TIMP1 has also been linked to pain. Disruption of the EZH2/TIMP1-MMP9 axis can activate the NLRP3 inflammasome and increase IL-1β secretion, further amplifying pain, especially in chronic constriction injury (CCI) models (Wan et al., 2025). MMP-9 also alters pain signaling by disrupting neuroglial interactions, breaking down neurotrophic factors such as NGF, and regulating ion channels like TRPV1 or transcription factors such as E74-like factor 1 (Li et al., 2023; Zhang et al., 2023). In thermal hyperalgesia, its involvement in nerve remodeling and pathways like PKC signaling appears more pronounced (Moon et al., 2012; Kim et al., 2014). Overall, higher levels of MMP-9 are often associated with more severe pain. Blocking MMP-9 activity has been shown to relieve symptoms in multiple experimental models, making it a promising therapeutic target (Table 2).

The function of MMP-9 varies significantly between pain models. This may be due to its presence in different cell types, such as neurons and immune cells, and its interaction with specific ECM components (Folgueras et al., 2009; Martins-Oliveira et al., 2009). These differences suggest that treatments targeting MMP-9 should be tailored to the specific form of neuropathic pain. For example, in trigeminal neuralgia, which is often caused by vascular compression, the inflammatory role of MMP-9 may be more relevant (Henry et al., 2015). In contrast, in long-term neuropathic pain, its impact on nerve remodeling and regeneration becomes more important (Tonello et al., 2019). Optimizing drug development based on MMP-9's specific function in each condition is essential. Evidence from animal studies shows that reducing MMP-9 activity can ease pain-related behaviors and hyperalgesia after nerve injury (Chattopadhyay et al., 2007; Zhang et al., 2016). Several inhibitors of MMP-9, such as BB-1101 and SB-3CT, have shown strong results in preclinical models. They reduce inflammation and protect neurons, contributing to pain relief (Demestre et al., 2004; Yin et al., 2019). Recent small-scale clinical studies echo these findings. For instance, Chu et al. (2020) found that patients with greater reductions in plasma MMP-9 levels after videoassisted thoracoscopic surgery reported less postoperative pain. This clinical observation supports the theory that MMP-9 plays a key role in pain regulation. Likewise, Ferianec et al. (2020) reported higher MMP-9 levels in patients with chronic pain, suggesting a link between circulating MMP-9 and pain severity. Taken together, both experimental and clinical data highlight the importance of MMP-9 in neuropathic pain. Targeting its activity could offer a new direction for developing effective and personalized pain therapies (Ferianec et al., 2020; Chu et al., 2020).

Recent studies have highlighted the role of MMP-9 in peripheral neuropathic pain. MMP-9 inhibitors are now under investigation as potential therapies. Several naturally derived compounds, such as β -sitosterol, resveratrol, mangiferin, and epigallocatechin-3-gallate, have shown analgesic effects in experimental models (Mohajeri et al.,

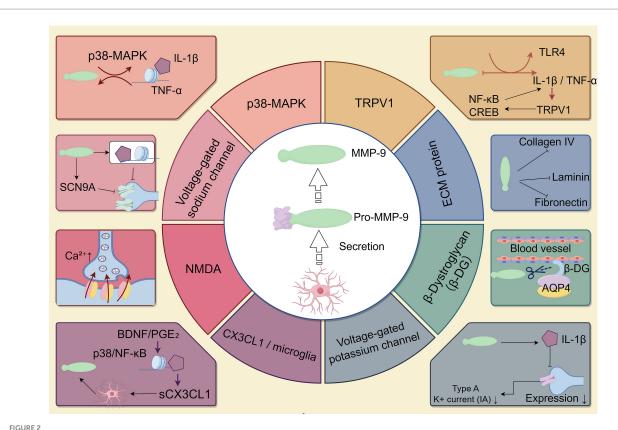
TABLE 1 The role of MMP-9 in different peripheral neuropathic pain models (preclinical studies).

Neuropathic pain model	MMP-9 expression	Mechanisms involved	Key findings	Key references
Chronic constriction injury of trigeminal nerve (mice)	MMP-9 was significantly upregulated in the trigeminal ganglion at early stages after CCI.	The TLR-4/NF-κB signaling pathway and SOCS3-mediated regulation were implicated in MMP-9/2 activation and pain sensitization.	Resveratrol pretreatment reduced MMP-9/2 activation and inflammatory cytokines	Yin et al. (2019)
Spinal nerve ligation model; chronic constriction injury of infraorbital nerve (rat)	Implicated indirectly via effect of MMP-2/–9 inhibition; not directly measured	Inhibition of MMP-2 and MMP-9 was shown to reduce mechanical allodynia	AQU-118, an MMP-2/9 inhibitor, demonstrated comparable efficacy to gabapentin	Henry et al. (2015)
Spinal nerve ligation (mice)	MMP-9 upregulated in spinal cord and DRG during neuroinflammation; Naru-3 reduces its expression	MMP-9/IL-1β signaling pathway-mediated microglia activation and inflammatory loop via p38/IL-1β axis	MMP-9 and IL-1β are key therapeutic targets of Naru-3's synergistic analgesic effects	Zhou et al. (2023)
Sciatic nerve crush (rat)	MMP-9 expression increased after sciatic nerve crush; significantly reduced by Hyperbaric oxygen therapy in DRG and spinal cord	MMP-9 involved in inflammatory response; Hyperbaric oxygen therapy reduces MMP-9, TRPV1, cytokines (TNFα, IL-6, IL-1β), and mitochondrial stress (TSPO)	Hyperbaric oxygen therapy modulates inflammation and mitochondrial function, reduces neuronal apoptosis, and prevents progression from acute to chronic neuropathic pain	Awad-Igbaria et al. (2023)
Chronic constriction injury (rat)	Upregulated rapidly (as early as 3 h post-CCI)	MMP-9 and MMP-2 mediate TNF-α activation and receptor modulation, contributing to axonal degeneration and endoneurial inflammation.	Targeting MMP-9 and MMP-2 may help prevent TNF-α-mediated neuroinflammation and pain in neuropathic conditions.	Shubayev and Myers (2000)
Chronic constriction injury (rat)	MMP-9 expression was significantly upregulated by CCI and downregulated following GDNF treatment.	GDNF mitigates neural injury by inhibiting microglial activation and inflammatory cytokines via p38 and PKC signaling pathways.	GDNF shows anti-inflammatory and neuroprotective effects, making it a potential therapeutic option for CCI-induced neuropathic pain.	Chou et al. (2014)
Partial sciatic nerve ligation (mice)	Upregulated after PSNL; suppressed by anti-HMGB1 antibody	HMGB1 upregulates MMP-9 in injured nerve; both contribute to mechanical hypersensitivity	Blocking HMGB1 or inhibiting MMP-9 reduces neuropathic pain	Zhang et al. (2016)
Painful diabetic neuropathy (rat)	MMP-9 expression was significantly increased in PDN rats	Inhibiting the expression of MMP9 can alleviate PDN	MMP-9 is a potential therapeutic target for improving PDN	Li et al. (2024)

2020; Lin et al., 2019; Tohge and Fernie, 2017; Yiemwattana et al., 2019; Subedi and Gaire, 2021). These agents also show promise in treating inflammatory and degenerative conditions. However, many of them are not selective for MMP-9. Their pain-relieving effects likely involve multiple mechanisms, including antioxidant and anti-inflammatory actions. Thus, the observed benefits cannot be attributed to MMP-9 inhibition alone.

Although MMP-9 is an attractive therapeutic target, its clinical application remains challenging. It interacts with inflammatory mediators such as IL-1 β , TNF- α , and NMDA receptors, placing it at the center of neuropathic pain regulation (Berta et al., 2012). A deeper understanding of these interactions is needed to develop effective combination strategies that reduce the drawbacks of monotherapy. Another challenge comes from the dual role of MMP-9 in nerve repair. After peripheral nerve injury, MMP-9

supports regeneration by regulating communication between neurons and glial cells (Kawasaki et al., 2008). Excessive inhibition can ease pain, but over-inhibition may block repair (LeBert et al., 2015). Therefore, strategies must strike a balance: suppressing pain without halting neural recovery. MMP-9 also influences sodium channels, such as Nav1.7, and receptors including TRPV1, which play major roles in neuropathic pain (Remacle et al., 2015; AwadIgbaria et al., 2023). E During neural repair, these interactions may be critical for pain development (Kim et al., 2012). This indicates that combining MMP-9 inhibitors with agents targeting sodium channels, NMDA receptors, or TRPV1 may improve efficacy and lower drug resistance. Still, most evidence so far comes from rodent models. Differences in expression patterns, immune responses, and pain phenotypes between species limit translation to humans. In addition, many studies rely on broad



The Role and Mechanisms of Matrix Metalloproteinase-9 in Peripheral Neuropathic Pain. MMP9 plays distinct roles in different types of peripheral neuropathic pain, particularly in conditions such as trigeminal neuralgia, neuropathic pain following peripheral nerve injury, painful polyneuropathy, and painful radiculopathy. This diagram highlights its crucial involvement in the pathophysiological processes of these disorders, underscoring its key role in their pathogenesis.

pharmacological inhibitors that carry off-target effects or trigger compensatory activation of other MMPs, such as MMP-2. These issues point to the need for selective genetic and proteomic approaches to define MMP-9's precise role.

Clinical research will also be critical. Current preclinical findings show that MMP-9 inhibitors can reduce pain in animal models (Kwan et al., 2019). However, our study has limitations. Because we used non-selective compounds and broad-spectrum inhibitors, we cannot conclude that MMP-9 alone drives the observed effects. Future studies should use more selective tools such as monoclonal antibodies, gene knockouts, or highly specific inhibitors—to clarify its exact contribution. Functional studies with these tools would also help minimize off-target concerns. Past failures of MMP inhibitors in clinical cancer trials remind us of the risks. These drugs lacked specificity, produced unacceptable side effects, and disrupted the function of MMPs with protective roles (Vandenbroucke and Libert, 2014). To avoid repeating this pattern, future success will depend on developing selective inhibitors that target disease-relevant MMPs like MMP-9, while sparing beneficial ones. Combining MMP-9 inhibition with agents targeting sodium channels, NMDA receptors, or TRPV1 may also offer synergistic benefits. In addition, advanced delivery systems, such as nanoparticles, could improve localization and reduce systemic toxicity. A clearer understanding of the context-specific functions of MMP-9 will guide the design of these new therapeutic strategies.

To move MMP-9 closer to clinical use as a treatment target, comprehensive and systematic clinical trials are needed to evaluate its efficacy, safety, and long-term effects. Additionally, given the variability among patients, precision medicine strategies based on MMP-9 could represent a major breakthrough in treating neuropathic pain. With rapid advancements in big data, artificial intelligence, and precision medicine, many studies now integrate genomics, proteomics, and clinical data to better understand MMP-9 and its pathways in various neuropathic pain types. This approach offers more precise evidence for personalized treatments (Knight et al., 2019; Osthues et al., 2020). Moreover, network pharmacology has identified new drug targets related to MMP-9, broadening the treatment options for neuropathic pain (Jo et al., 2023). However, these methods are still in early stages, and their clinical effectiveness has yet to be rigorously tested in large patient populations. The lack of standardized biomarkers for MMP-9 activity in human tissues makes it difficult to categorize patients or track therapeutic responses. Additionally, previous attempts to develop MMP inhibitors for cancer and inflammatory diseases failed in clinical trials due to poor specificity and adverse effects. These failures raise concerns about the safety and feasibility of MMP-9 inhibitors in managing chronic pain. Therefore, future clinical applications of MMP-9 inhibitors must be preceded by developing highly selective agents and robust diagnostic tools to guide targeted therapy.

In summary, MMP-9 plays a key role in the onset and progression of various types of peripheral neuropathic pain.

TABLE 2 Summary of major clinical studies.

Population	Intervention	Study design	Key findings	Limitations	Key references
14 healthy participants	Acupuncture was applied at specific points used in the treatment of spinal cord injury.	Interventional study	Acupuncture significantly decreased BDNF and MMP-9 levels in peripheral blood.	Small sample size and lack of clinical outcomes related to spinal cord injury.	Moldenhauer et al. (2010)
21 patients with Guillain- Barré syndrome	Plasma MMP-9 levels	Observational correlation study	Higher plasma MMP-9 levels were associated with demyelination and peripheral nerve dysfunction in Guillain- Barré syndrome.	Small sample size and lack of causal inference.	Sharshar et al. (2002)
86 patients with refractory diabetic dermal ulcers	Patients received either standard wound care or standard care plus topical application of autologous platelet-rich gel (APG).	Randomized controlled trial	Topical APG significantly reduced ulcer area and improved proteolytic balance by lowering MMPs and increasing TIMP-1.	Lacks long-term follow-up and detailed analysis of systemic effects.	He et al. (2012)
33 patients with chronic diabetic foot lesions (UT stage 2a)	Patients received either standard wound care or standard care plus daily application of ORC/collagen protease-inhibiting matrix.	Interventional study	The ORC/collagen matrix reduced the MMP-9/TIMP-2 ratio	Small sample size and short follow-up duration limited to 8 days	Lobmann et al. (2006)
12 patients with non- healing diabetic foot ulcers	Participants were randomized to receive non-contact low-frequency ultrasound (NCLF-US) either three times or once per week, or no ultrasound treatment.	Prospective randomized clinical trial	Thrice-weekly NCLF- US treatment significantly reduced wound area and was associated with decreased MMP-9, pro-inflammatory cytokines and improved healing.	Small sample size, limiting the generalizability of the results	Yao et al. (2014)
24 patients with diabetic foot ulcers	Topical propolis was applied to diabetic foot ulcers	Controlled Clinical Trial	Topical propolis significantly improved ulcer healing rates and reduced bacterial load and MMP-9 activity compared to controls.	Small sample size, single- center design, and use of a non-randomized control group.	Henshaw et al. (2014)

Although its clinical application is still in early stages, growing clinical evidence supports its potential as both a biomarker and a therapeutic target. Importantly, clinical studies have begun to confirm mechanisms previously identified in preclinical models. For example, clinical data show that patients who experienced better postoperative analgesia after video-assisted thoracoscopic surgery lobectomy also had lower plasma MMP-9 levels, indicating its role in pain modulation (Chu et al., 2020). Other studies have found correlations between plasma MMP-9 levels and symptom severity in patients with acute coronary syndrome, suggesting that MMP-9 could be used as a biomarker for disease progression and patient stratification (Hamed and Fattah, 2015; Kobayashi et al., 2011). Although these studies do not yet demonstrate direct therapeutic modulation of MMP-9, they highlight its importance in human disease and support further investigation in precision medicine.

Taken together, these findings highlight the complex role of MMP-9 and other MMP isoforms in chronic pain. Targeted inhibition of specific MMP isoforms-particularly MMP-2 and MMP-9—has shown promise in reducing neuropathic pain while preserving normal matrix remodeling. However, challenges remain regarding specificity, pharmacokinetics, and off-target effects. Additionally, the potential of MMPs as diagnostic and prognostic biomarkers requires further validation, particularly for distinguishing between inflammatory and neuropathic pain. MMPs also modulate glial activation and neuroimmune interactions, making them key players in central sensitization and the persistence of chronic pain. Innovative techniques such as gene editing (e.g., CRISPR/Cas9) and RNA interference (e.g., siRNA, miRNA) offer promising approaches to selectively suppress $\ensuremath{\mathsf{MMP}}$ expression in targeted neural regions. These strategies could overcome the limitations of traditional small-molecule inhibitors.

At the same time, understanding the interactions between MMPs and other pain-related pathways, such as Toll-like receptors, pro-inflammatory cytokines, and ion channels, may uncover additional targets for combination therapies. Future research should integrate these mechanistic insights with translational strategies, advancing MMP-targeted therapeutics that are not only effective but also safe and clinically viable. Ultimately, deepening our understanding of MMP biology in chronic pain will be essential in bridging the gap between bench research and clinical application.

Author contributions

SJ: Writing – original draft, Writing – review & editing. X-JT: Writing – original draft, Writing – review & editing. YS: Conceptualization, Methodology, Writing – review & editing. Y-NL: Methodology, Writing – review & editing. ZM: Methodology, Writing – review & editing. J-YL: Conceptualization, Funding acquisition, Project administration, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This research was supported by the Nanchong Science and Technology Bureau Municipal School Cooperation Project (22SXQT0125).

References

Abbott, C. A., Malik, R. A., Van Ross, E. R., Kulkarni, J., and Boulton, A. J. (2011). Prevalence and characteristics of painful diabetic neuropathy in a large community-based diabetic population in the U.K. *Diabetes Care* 34, 2220–2224. doi: 10.2337/dc11-1108

Acklin, S., Zhang, M., Du, W., Zhao, X., Plotkin, M., Chang, J., et al. (2020). Depletion of senescent-like neuronal cells alleviates cisplatin-induced peripheral neuropathy in mice. *Sci. Rep.* 10:14170. doi: 10.1038/s41598-020-71042-6

Adams, S. B., Setton, L. A., Bell, R. D., Easley, M. E., Huebner, J. L., Stabler, T., et al. (2015). Inflammatory cytokines and matrix metalloproteinases in the synovial fluid after intra-articular ankle fracture. *Foot Ankle Int.* 36, 1264–1271. doi: 10.1177/1071100715611176

Agrawal, A., Romero-Perez, D., Jacobsen, J. A., Villarreal, F. J., and Cohen, S. M. (2008). Zinc-binding groups modulate selective inhibition of MMPs. *ChemMedChem* 3, 812–820. doi: 10.1002/cmdc.200700290

Anonymous (2018). Headache classification Committee of the International Headache Society (IHS) the International Classification of Headache Disorders, 3rd edition. *Cephalalgia* 38, 1–211. doi: 10.1177/0333102417738202

Aronson, R., Chu, L., Joseph, N., and Brown, R. (2021). Prevalence and risk evaluation of diabetic complications of the foot among adults with type 1 and type 2 diabetes in a large Canadian population (PEDAL study). *Can. J. Diabetes* 45, 588–593. doi: 10.1016/j.jcjd.2020.11.011

Awad-Igbaria, Y., Ferreira, N., Keadan, A., Sakas, R., Edelman, D., Shamir, A., et al. (2023). HBO treatment enhances motor function and modulates pain development after sciatic nerve injury via protection the mitochondrial function. *J. Transl. Med.* 21:545. doi: 10.1186/s12967-023-04414-x

Balog, B. M., Sonti, A., and Zigmond, R. E. (2023). Neutrophil biology in injuries and diseases of the central and peripheral nervous systems. *Prog. Neurobiol.* 228:102488. doi: 10.1016/j.pneurobio.2023.102488

Bendtsen, L., Zakrzewska, J. M., Heinskou, T. B., Hodaie, M., Leal, P. R. L., Nurmikko, T., et al. (2020). Advances in diagnosis, classification, pathophysiology, and management of trigeminal neuralgia. *Lancet Neurol.* 19, 784–796. doi: 10.1016/S1474-4422(20)30233-7

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The authors declare that Gen AI was used in the creation of this manuscript. During the preparation of this work the authors used (ChatGPT 4.0, GPT-4, Open AI) in order to improve readability and language. After using this tool, the authors reviewed and edited the content as needed and take full responsibility for the content of the publication. The illustrations for this manuscript were created using figdraw (www.figdraw.com).

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Bermudez-Lekerika, P., Crump, K. B., Tseranidou, S., Nüesch, A., Kanelis, E., Alminnawi, A., et al. (2022). Immuno-modulatory effects of intervertebral disc cells. *Front. Cell Dev. Biol.* 10:924692. doi: 10.3389/fcell.2022.924692

Berta, T., Liu, T., Liu, Y. C., Xu, Z. Z., and Ji, R. R. (2012). Acute morphine activates satellite glial cells and up-regulates IL-1 β in dorsal root ganglia in mice via matrix metalloprotease-9. *Mol. Pain* 8:18. doi: 10.1186/1744-8069-8-18

Bierhaus, A., Fleming, T., Stoyanov, S., Leffler, A., Babes, A., Neacsu, C., et al. (2012). Methylglyoxal modification of Nav1.8 facilitates nociceptive neuron firing and causes hyperalgesia in diabetic neuropathy. *Nat. Med.* 18, 926–933. doi: 10.1038/nm.2750

Bjorland, S., Gjerstad, J., Schistad, E., Swanson, D. M., and RøE, C. (2019). Persistent lumbar radicular and low back pain; impact of genetic variability versus emotional distress. *BMC. Res. Notes* 12:547. doi: 10.1186/s13104-019-4592-6

Bjorland, S., Røe, C., Moen, A., Schistad, E., Mahmood, A., and Gjerstad, J. (2017). Genetic predictors of recovery in low back and lumbar radicular pain. *Pain* 158, 1456–1460. doi: 10.1097/j.pain.000000000000934

Bratcher, P. E., Weathington, N. M., Nick, H. J., Jackson, P. L., Snelgrove, R. J., and Gaggar, A. (2012). MMP-9 cleaves SP-D and abrogates its innate immune functions in vitro. *PLoS One* 7:e41881. doi: 10.1371/journal.pone.0041881

Butturini, E., Carcereri de Prati, A., Chiavegato, G., Rigo, A., Cavalieri, E., Darra, E., et al. (2013). Mild oxidative stress induces S-glutathionylation of STAT3 and enhances chemosensitivity of tumoural cells to chemotherapeutic drugs. *Free Radic. Biol. Med.* 65, 1322–1330. doi: 10.1016/j.freeradbiomed.2013.09.015

Cabral-Pacheco, G. A., Garza-Veloz, I., Castruita-De la Rosa, C., Ramirez-Acuña, J. M., Perez-Romero, B. A., Guerrero-Rodriguez, J. F., et al. (2020). The roles of matrix metalloproteinases and their inhibitors in human diseases. *Int. J. Mol. Sci.* 21:9739. doi: 10.3390/jims21249739

Chattopadhyay, S., Myers, R. R., Janes, J., and Shubayev, V. (2007). Cytokine regulation of MMP-9 in peripheral glia: implications for pathological processes and pain in injured nerve. *Brain Behav. Immun.* 21, 561–568. doi: 10.1016/j.bbi.2006.10.015

Chopko, B., Liu, J. C., and Khan, M. K. (2014). Anatomic surgical management of chronic low back pain. *Neuromodulation* 17, 46–51. doi: 10.1111/ner.12169

- Chou, A. K., Yang, M. C., Tsai, H. P., Chai, C. Y., Tai, M. H., Kwan, A. L., et al. (2014). Adenoviral-mediated glial cell line-derived neurotrophic factor gene transfer has a protective effect on sciatic nerve following constriction-induced spinal cord injury. *PLoS One* 9:e92264. doi: 10.1371/journal.pone.0092264
- Chu, H., Dong, H., Wang, Y., and Niu, Z. (2020). Effects of ultrasound-guided paravertebral block on MMP-9 and postoperative pain in patients undergoing VATS lobectomy: a randomized, controlled clinical trial. *BMC Anesthesiol.* 20:59. doi: 10.1186/s12871-020-00976-1
- Da Silva, T., Mills, K., Brown, B. T., Herbert, R. D., Maher, C. G., and Hancock, M. J. (2017). Risk of recurrence of low back pain: a systematic review. *J. Orthop. Sports Phys. Ther.* 47, 305–313. doi: 10.2519/jospt.2017.7415
- Demestre, M., Wells, G. M., Miller, K. M., Smith, K. J., Hughes, R. A., Gearing, A. J., et al. (2004). Characterisation of matrix metalloproteinases and the effects of a broad-spectrum inhibitor (BB-1101) in peripheral nerve regeneration. *Neuroscience* 124, 767–779. doi: 10.1016/j.neuroscience.2003.12.037
- Deng, X., Ma, P., Wu, M., Liao, H., and Song, X. J. (2021). Role of matrix metalloproteinases in myelin abnormalities and mechanical allodynia in rodents with diabetic neuropathy. *Aging Dis.* 12, 1808–1820. doi: 10.14336/AD.2021.0126
- Desiderá, A. C., Nascimento, G. C., Gerlach, R. F., and Leite-Panissi, C. R. (2015). Laser therapy reduces gelatinolytic activity in the rat trigeminal ganglion during temporomandibular joint inflammation. *Oral Dis.* 21, 652–658. doi: 10.1111/odi.12330
- Dreier, R., Grässel, S., Fuchs, S., Schaumburger, J., and Bruckner, P. (2004). Pro-MMP-9 is a specific macrophage product and is activated by osteoarthritic chondrocytes via MMP-3 or a MT1-MMP/MMP-13 cascade. *Exp. Cell Res.* 297, 303–312. doi: 10.1016/j.yexcr.2004.02.027
- Dunbar, J., Versavel, M., Zhao, Y., Tate, S., Morisset, V., Giblin, G. M. P., et al. (2020). Evaluation of the pharmacokinetic interaction between the voltage- and use-dependent Nav1.7 channel blocker vixotrigine and carbamazepine in healthy volunteers. *Clin. Pharmacol. Drug Dev.* 9, 62–73. doi: 10.1002/cpdd.739
- Duraisamy, A. J., Mishra, M., and Kowluru, R. A. (2017). Crosstalk between histone and DNA methylation in regulation of retinal matrix metalloproteinase-9 in diabetes. *Invest. Ophthalmol. Vis. Sci.* 58, 6440–6448. doi: 10.1167/iovs.17-22706
- Edlich, R. F., Winters, K. L., Britt, L., and Long, W. B. (2006). Trigeminal neuralgia. *J. Long-Term Eff. Med. Implants* 16, 185–192. doi: 10.1615/jlongtermeffmedimplants. v16.i2.80
- Erbüyün, S. C., Oval, G. Y., Özyurt, B., and Yentür, E. A. (2018). Lumbar disc herniation, the association between quantitative sensorial test and magnetic resonance imaging findings. *Pain Med.* 19, 1373–1381. doi: 10.1093/pm/pnx236
- Fan, Y. X., Hu, L., Zhu, S. H., Han, Y., Liu, W. T., Yang, Y. J., et al. (2018). Paeoniflorin attenuates postoperative pain by suppressing matrix metalloproteinase-9/2 in mice. *Eur. J. Pain* 22, 272–281. doi: 10.1002/ejp.1116
- Ferianec, V., Fülöp, M., Ježovičová, M., Radošinská, J., Husseinová, M., Feriancová, M., et al. (2020). The oak-wood extract Robuvit(**) improves recovery and oxidative stress after hysterectomy: A randomized, double-blind, placebo-controlled pilot study. *Nutrients* 12:913. doi: 10.3390/nu12040913
- Fernández Rodríguez, B., Simonet, C., Cerdán, D. M., Morollón, N., Guerrero, P., Tabernero, C., et al. (2019). Familial classic trigeminal neuralgia. *Neurologia (Engl Ed)* 34, 229–233. doi: 10.1016/j.nrl.2016.12.004
- Ferreira, A. C., Romão, T. T., Macedo, Y. S., Pupe, C., and Nascimento, O. J.Fellow of the American Academy of Neurology (FAAN) (2020). COVID-19 and herpes zoster co-infection presenting with trigeminal neuropathy. *Eur. J. Neurol.* 27, 1748–1750. doi: 10.1111/ene.14361
- Filanti, C., Dickson, G. R., Di Martino, D., Ulivi, V., Sanguineti, C., Romano, P., et al. (2000). The expression of metalloproteinase-2, –9, and –14 and of tissue inhibitors-1 and -2 is developmentally modulated during osteogenesis *in vitro*, the mature osteoblastic phenotype expressing metalloproteinase-14. *J. Bone Miner. Res.* 15, 2154–2168. doi: 10.1359/jbmr.2000.15.11.2154
- Finnerup, N. B., Kuner, R., and Jensen, T. S. (2021). Neuropathic pain: from mechanisms to treatment. *Physiol. Rev.* 101, 259–301. doi: 10.1152/physrev.00045.2019
- Folgueras, A. R., Valdés-Sánchez, T., Llano, E., Menéndez, L., Baamonde, A., Denlinger, B. L., et al. (2009). Metalloproteinase MT5-MMP is an essential modulator of neuro-immune interactions in thermal pain stimulation. *Proc. Natl. Acad. Sci. USA* 106, 16451–16456. doi: 10.1073/pnas.0908507106
- Freynhagen, R., Baron, R., Tölle, T., Stemmler, E., Gockel, U., Stevens, M., et al. (2006). Screening of neuropathic pain components in patients with chronic back pain associated with nerve root compression: a prospective observational pilot study (MIPORT). *Curr. Med. Res. Opin.* 22, 529–537. doi: 10.1185/030079906x89874
- Gao, S., Wang, C., Qi, L., Liang, S., Qu, X., Liu, W., et al. (2024). Bushen Huoxue formula inhibits IL-1β-induced apoptosis and extracellular matrix degradation in the nucleus pulposus cells and improves intervertebral disc degeneration in rats. *J. Inflamm. Res.* 17, 121−136. doi: 10.2147/jir.S431609
- Gong, Y., Qiu, J., Jiang, T., Li, Z., Zhang, W., Zheng, X., et al. (2023). Maltol ameliorates intervertebral disc degeneration through inhibiting PI3K/AKT/NF-κB pathway and regulating NLRP3 inflammasome-mediated pyroptosis. *Inflammopharmacology* 31, 369–384. doi: 10.1007/s10787-022-01098-5

- Gu, H., Wang, C., Li, J., Yang, Y., Sun, W., Jiang, C., et al. (2020). High mobility group box-1-toll-like receptor 4-phosphatidylinositol 3-kinase/protein kinase B-mediated generation of matrix metalloproteinase-9 in the dorsal root ganglion promotes chemotherapy-induced peripheral neuropathy. *Int. J. Cancer* 146, 2810–2821. doi: 10.1002/jic.32652
- Gu, H. W., Xing, F., Jiang, M. J., Wang, Y., Bai, L., Zhang, J., et al. (2019). Upregulation of matrix metalloproteinase-9/2 in the wounded tissue, dorsal root ganglia, and spinal cord is involved in the development of postoperative pain. *Brain Res.* 1718, 64–74. doi: 10.1016/j.brainres.2019.05.007
- Gui, W. S., Wei, X., Mai, C. L., Murugan, M., Wu, L. J., Xin, W. J., et al. (2016). Interleukin-1β overproduction is a common cause for neuropathic pain, memory deficit, and depression following peripheral nerve injury in rodents. *Mol. Pain* 12:6784. doi: 10.1177/1744806916646784
- Guzmán, J., Esmail, R., Karjalainen, K., Malmivaara, A., Irvin, E., and Bombardier, C. (2001). Multidisciplinary rehabilitation for chronic low back pain: systematic review. *BMJ* 322, 1511–1516. doi: 10.1136/bmj.322.7301.1511
- Gylfadottir, S. S., Christensen, D. H., Nicolaisen, S. K., Andersen, H., Callaghan, B. C., Itani, M., et al. (2020). Diabetic polyneuropathy and pain, prevalence, and patient characteristics: a cross-sectional questionnaire study of 5,514 patients with recently diagnosed type 2 diabetes. Pain 161, 574–583. doi: 10.1097/j.pain.00000000000001744
- Hackel, D., Krug, S. M., Sauer, R. S., Mousa, S. A., Böcker, A., Pflücke, D., et al. (2012). Transient opening of the perineurial barrier for analgesic drug delivery. *Proc. Natl. Acad. Sci. USA* 109, E2018–E2027. doi: 10.1073/pnas.1120800109
- Hamed, G. M., and Fattah, M. F. (2015). Clinical relevance of matrix metalloproteinase 9 in patients with acute coronary syndrome. *Clin. Appl. Thromb. Hemost.* 21, 705–711. doi: 10.1177/1076029614567309
- Hamlyn, P. J., and King, T. T. (1992). Neurovascular compression in trigeminal neuralgia: a clinical and anatomical study. *J. Neurosurg.* 76, 948–954. doi: 10.3171/jns.1992.76.6.0948
- Han, K. R., Kim, C., Chae, Y. J., and Kim, D. W. (2008). Efficacy and safety of high concentration lidocaine for trigeminal nerve block in patients with trigeminal neuralgia. *Int. J. Clin. Pract.* 62, 248–254. doi: 10.1111/j.1742-1241.2007.01568.x
- Hangping, Z., Ling, H., Lijin, J., Wenting, Z., Xiaoxia, L., Qi, Z., et al. (2020). The preventive effect of IL-1beta antagonist on diabetic peripheral neuropathy. *Endocr. Metab. Immune Disord. Drug Targets* 20, 753–759. doi: 10.2174/1871530319666191022114139
- He, L. P., Wang, C., Chen, D. W., Li, X. J., and Ran, X. W. (2012). Dynamic changes of MMP-1, MMP-9 and TIMP-1 in the refractory diabetic dermal ulcers treated by autologous platelet-rich gel. Sichuan Da Xue Xue Bao Yi Xue Ban 43, 757–761.
- Helm Ii, S., Benyamin, R. M., Chopra, P., Deer, T. R., and Justiz, R. (2012). Percutaneous adhesiolysis in the management of chronic low back pain in post lumbar surgery syndrome and spinal stenosis: a systematic review. *Pain Physician* 15, E435–E462.
- Henry, M. A., Fairchild, D. D., Patil, M. J., Hanania, T., Hain, H. S., Davis, S. F., et al. (2015). Effect of a novel, orally active matrix metalloproteinase-2 and -9 inhibitor in spinal and trigeminal rat models of neuropathic pain. *J. Oral Facial Pain Headache* 29, 286–296. doi: 10.11607/ofph.1350
- Henshaw, F. R., Bolton, T., Nube, V., Hood, A., Veldhoen, D., Pfrunder, L., et al. (2014). Topical application of the bee hive protectant propolis is well tolerated and improves human diabetic foot ulcer healing in a prospective feasibility study. *J. Diabetes Complicat.* 28, 850–857. doi: 10.1016/j.jdiacomp.2014.07.012
- Hingorani, D. V., Lippert, C. N., Crisp, J. L., Savariar, E. N., Hasselmann, J. P. C., Kuo, C., et al. (2018). Impact of MMP-2 and MMP-9 enzyme activity on wound healing, tumor growth and RACPP cleavage. *PLoS One* 13:e0198464. doi: 10.1371/journal.pone.0198464
- Hoy, D., Brooks, P., Blyth, F., and Buchbinder, R. (2010). The epidemiology of low back pain. *Best Pract. Res. Clin. Rheumatol.* 24, 769–781. doi: 10.1016/j.berh.2010.10.002
- Hu, L. Y., Mi, W. L., Wu, G. C., Wang, Y. Q., and Mao-Ying, Q. L. (2019). Prevention and treatment for chemotherapy-induced peripheral neuropathy: therapies based on CIPN mechanisms. *Curr. Neuropharmacol.* 17, 184–196. doi: 10.2174/1570159X15666170915143217
- Huang, Y., Zhang, B., Haneke, H., Haage, V., Lubas, M., Yuan, Y., et al. (2021). Glial cell line-derived neurotrophic factor increases matrix metallopeptidase 9 and 14 expression in microglia and promotes microglia-mediated glioma progression. *J. Neurosci. Res.* 99, 1048–1063. doi: 10.1002/jnr.24768
- Iqbal, Z., Azmi, S., Yadav, R., Ferdousi, M., Kumar, M., Cuthbertson, D. J., et al. (2018). Diabetic peripheral neuropathy: epidemiology, diagnosis, and pharmacotherapy. *Clin. Ther.* 40, 828–849. doi: 10.1016/j.clinthera.2018.04.001
- Javed, S., Petropoulos, I. N., Alam, U., and Malik, R. A. (2015). Treatment of painful diabetic neuropathy. *Ther. Adv. Chronic Dis.* 6, 15–28. doi: 10.1177/2040622314552071
- Jiang, M., Liu, Y., Wu, H., Ma, Z., and Gu, X. (2020). High estrogen level modifies postoperative hyperalgesia via GPR30 and MMP-9 in dorsal root ganglia neurons. *Neurochem. Res.* 45, 1661–1673. doi: 10.1007/s11064-020-03032-z

- Jo, H. G., Baek, C. Y., Kim, D., Kim, S., Han, Y., Park, C., et al. (2023). Network analysis, in vivo, and in vitro experiments identified the mechanisms by which *Piper longum* L. [Piperaceae] alleviates cartilage destruction, joint inflammation, and arthritic pain. *Front. Pharmacol.* 14:1282943. doi: 10.3389/fphar.2023.1282943
- Jones, M. R., Urits, I., Ehrhardt, K. P., Cefalu, J. N., Kendrick, J. B., Park, D. J., et al. (2019). A comprehensive review of trigeminal neuralgia. *Curr. Pain Headache Rep.* 23:74. doi: 10.1007/s11916-019-0810-0
- Kaki, A. M., El-Yaski, A. Z., and Youseif, E. (2005). Identifying neuropathic pain among patients with chronic low-back pain: use of the Leeds assessment of neuropathic symptoms and signs pain scale. *Reg. Anesth. Pain Med.* 30, 422–428. doi: 10.1016/j.rapm.2005.05.013
- Kamieniak, P., Bielewicz, J., Kurzepa, J., Daniluk, B., Kocot, J., and Trojanowski, T. (2019). The impact of changes in serum levels of metalloproteinase-2 and metalloproteinase-9 on pain perception in patients with disc herniation before and after surgery. *J. Pain Res.* 12, 1457–1464. doi: 10.2147/JPR.S201199
- Kawasaki, Y., Xu, Z. Z., Wang, X., Park, J. Y., Zhuang, Z. Y., Tan, P. H., et al. (2008). Distinct roles of matrix metalloproteases in the early- and late-phase development of neuropathic pain. *Nat. Med.* 14, 331–336. doi: 10.1038/nm1723
- Kim, M. J., Lee, S. Y., Yang, K. Y., Nam, S. H., Kim, H. J., Kim, Y. J., et al. (2014). Differential regulation of peripheral IL-1 β -induced mechanical allodynia and thermal hyperalgesia in rats. *Pain* 155, 723–732. doi: 10.1016/j.pain.2013.12.030
- Kim, Y., Remacle, A. G., Chernov, A. V., Liu, H., Shubayev, I., Lai, C., et al. (2012). The MMP-9/TIMP-1 axis controls the status of differentiation and function of myelinforming Schwann cells in nerve regeneration. *PLoS One* 7:e33664. doi: 10.1371/journal.pone.0033664
- Knight, B. E., Kozlowski, N., Havelin, J., King, T., Crocker, S. J., Young, E. E., et al. (2019). TIMP-1 attenuates the development of inflammatory pain through MMP-dependent and receptor-mediated cell signaling mechanisms. *Front. Mol. Neurosci.* 12:220. doi: 10.3389/fnmol.2019.00220
- Kobayashi, N., Hata, N., Kume, N., Yokoyama, S., Shinada, T., Tomita, K., et al. (2011). Matrix metalloproteinase-9 for the earliest stage acute coronary syndrome. *Circ. J.* 75, 2853–2861. doi: 10.1253/circj.cj-11-0640
- Koskinen, M. J., Kautio, A. L., Haanpää, M. L., Haapasalo, H. K., Kellokumpu-Lehtinen, P. L., Saarto, T., et al. (2011). Intraepidermal nerve fibre density in cancer patients receiving adjuvant chemotherapy. *Anticancer Res.* 31, 4413–4416.
- Kowacs, P. A., Utiumi, M. A., Nascimento, F. A., Piovesan, E. J., and Teive, H. A. (2015). OnabotulinumtoxinA for trigeminal neuralgia: a review of the available data. *Arq. Neuropsiquiatr.* 73, 877–884. doi: 10.1590/0004-282X20150109
- Kwan, M. Y., Choo, A., Hanania, T., Ghavami, A., Beltran, J., Shea, J., et al. (2019). Biomarker analysis of orally dosed, dual active, matrix metalloproteinase (MMP)-2 and MMP-9 inhibitor, AQU-118, in the spinal nerve ligation (SNL) rat model of neuropathic pain. *Int. J. Mol. Sci.* 20:811. doi: 10.3390/ijms20040811
- Lebert, D. C., Squirrell, J. M., Rindy, J., Broadbridge, E., Lui, Y., Zakrzewska, A., et al. (2015). Matrix metalloproteinase 9 modulates collagen matrices and wound repair. *Development* 142, 2136–2146. doi: 10.1242/dev.121160
- Lemos, G. A., Rissi, R., de Souza Pires, I. L., de Oliveira, L. P., de Aro, A. A., Pimentel, E. R., et al. (2016). Low-level laser therapy stimulates tissue repair and reduces the extracellular matrix degradation in rats with induced arthritis in the temporomandibular joint. *Lasers Med. Sci.* 31, 1051–1059. doi: 10.1007/s10103-016-1946-3
- Li, J., Jia, S., Song, Y., Xu, W., and Lin, J. (2024). Ginkgolide B can alleviate spinal cord glymphatic system dysfunction and provide neuroprotection in painful diabetic neuropathy rats by inhibiting matrix metalloproteinase-9. *Neuropharmacology* 250:109907. doi: 10.1016/j.neuropharm.2024.109907
- Li, Z., Li, X., Chen, C., MTV, C., WKK, W., and Shen, J. (2017). Melatonin inhibits nucleus pulposus (NP) cell proliferation and extracellular matrix (ECM) remodeling via the melatonin membrane receptors mediated PI3K-Akt pathway. *J. Pineal Res.* 63:2435. doi: 10.1111/jpi.12435
- Li, J., Ruan, S., Jia, J., Li, Q., Jia, R., Wan, L., et al. (2023). Hydrogen attenuates postoperative pain through Trx1/ASK1/MMP9 signaling pathway. *J. Neuroinflammation* 20:22. doi: 10.1186/s12974-022-02670-0
- Li, S., Ung, T. T., Nguyen, T. T., Sah, D. K., Park, S. Y., and Jung, Y. D. (2020). Cholic acid stimulates MMP-9 in human Colon Cancer cells via activation of MAPK, AP-1, and NF- κ B activity. *Int. J. Mol. Sci.* 21:3420. doi: 10.3390/ijms21103420
- Liao, Y. H., Zhang, G. H., Jia, D., Wang, P., Qian, N. S., He, F., et al. (2011). Spinal astrocytic activation contributes to mechanical allodynia in a mouse model of type 2 diabetes. *Brain Res.* 1368, 324–335. doi: 10.1016/j.brainres.2010.10.044
- Lin, F. L., Cheng, Y. W., Yu, M., Ho, J. D., Kuo, Y. C., Chiou, G. C. Y., et al. (2019). The fungus-derived retinoprotectant theissenolactone C improves glaucoma-like injury mediated by MMP-9 inhibition. *Phytomedicine* 56, 207–214. doi: 10.1016/j.phymed.2018.11.002
- Ling, J., Erol, F., and Gu, J. G. (2018). Role of KCNQ2 channels in orofacial cold sensitivity: KCNQ2 upregulation in trigeminal ganglion neurons after infraorbital nerve chronic constrictive injury. *Neurosci. Lett.* 664, 84–90. doi: 10.1016/j.neulet.2017.11.026
- $\label{eq:Liu, S., Liu, W. T., Liu, Y. P., Dong, H. L., Henkemeyer, M., Xiong, L. Z., et al. (2011). \\ Blocking EphB1 receptor forward signaling in spinal cord relieves bone cancer pain and \\$

rescues analgesic effect of morphine treatment in rodents. Cancer Res. 71, 4392–4402. doi: 10.1158/0008-5472.CAN-10-3870

- Liu, Y., Ni, Y., Zhang, W., Sun, Y. E., Ma, Z., and Gu, X. (2017). N-acetyl-cysteine attenuates remifentanil-induced postoperative hyperalgesia via inhibiting matrix metalloproteinase-9 in dorsal root ganglia. *Oncotarget* 8, 16988–17001. doi: 10.18632/oncotarget.15217
- Liu, M., Zhong, J., Xia, L., Dou, N., and Li, S. (2019). The expression of voltage-gated sodium channels in trigeminal nerve following chronic constriction injury in rats. *Int. J. Neurosci.* 129, 955–962. doi: 10.1080/00207454.2019.1595616
- Lobmann, R., Zemlin, C., Motzkau, M., Reschke, K., and Lehnert, H. (2006). Expression of matrix metalloproteinases and growth factors in diabetic foot wounds treated with a protease absorbent dressing. *J. Diabetes Complicat.* 20, 329–335. doi: 10.1016/j.jdiacomp.2005.08.007
- Lopes, B., Sousa, P., Alvites, R., Branquinho, M., Sousa, A. C., Mendonça, C., et al. (2022). Peripheral nerve injury treatments and advances: one health perspective. *Int. J. Mol. Sci.* 23:918. doi: 10.3390/ijms23020918
- Lu, P., Wang, G., Lu, X., Qiao, P., Jin, Y., Yu, J., et al. (2022). Elevated matrix metalloproteinase 9 supports peripheral nerve regeneration via promoting Schwann cell migration. *Exp. Neurol.* 352:114020. doi: 10.1016/j.expneurol.2022.114020
- Luthfiana, D., and Utomo, D. H. (2023). Network pharmacology reveals the potential of Dolastatin 16 as a diabetic wound healing agent. *In Silico Pharmacol.* 11:23. doi: 10.1007/s40203-023-00161-5
- Lv, C. C., Xia, M. L., Shu, S. J., Chen, F., and Jiang, L. S. (2018). Attenuation of remifentanil-induced hyperalgesia by betulinic acid associates with inhibiting oxidative stress and inflammation in spinal dorsal horn. *Pharmacology* 102, 300–306. doi: 10.1159/000493144
- Lyu, F. J., Cui, H., Pan, H., Mc Cheung, K., Cao, X., Iatridis, J. C., et al. (2021). Painful intervertebral disc degeneration and inflammation: from laboratory evidence to clinical interventions. *Bone Res* 9:7. doi: 10.1038/s41413-020-00125-x
- Mahmood, A. K., Moen, A., Stafne, S. N., Robinson, H. S., Vøllestad, N. K., Salvesen, K. Å., et al. (2018). The MMP9 rs17576 A>G polymorphism is associated with increased lumbopelvic pain-intensity in pregnant women. *Scand J Pain* 18, 93–98. doi: 10.1515/sjpain-2017-0168
- Malemud, C. J. (2006). Matrix metalloproteinases: role in skeletal development and growth plate disorders. *Front. Biosci.* 11, 1702–1715. doi: 10.2741/1916
- Mannion, A. F., Brox, J. I., and Fairbank, J. C. (2013). Comparison of spinal fusion and nonoperative treatment in patients with chronic low back pain: long-term follow-up of three randomized controlled trials. *Spine J.* 13, 1438–1448. doi: 10.1016/j.spinee.2013.06.101
- Marinković, S., Gibo, H., Todorović, V., Antić, B., Kovacević, D., Milisavljević, M., et al. (2009). Ultrastructure and immunohistochemistry of the trigeminal peripheral myelinated axons in patients with neuralgia. *Clin. Neurol. Neurosurg.* 111, 795–800. doi: 10.1016/j.clineuro.2009.07.020
- Martins-Oliveira, A., Speciali, J. G., Dach, F., Marcaccini, A. M., Gonçalves, F. M., Gerlach, R. F., et al. (2009). Different circulating metalloproteinases profiles in women with migraine with and without aura. *Clin. Chim. Acta* 408, 60–64. doi: 10.1016/j.cca.2009.07.008
- Mcclellan, S. A., Huang, X., Barrett, R. P., Lighvani, S., Zhang, Y., Richiert, D., et al. (2006). Matrix metalloproteinase-9 amplifies the immune response to *Pseudomonas aeruginosa* corneal infection. *Invest. Ophthalmol. Vis. Sci.* 47, 256–264. doi: 10.1167/iovs.05-1050
- Meacham, K., Shepherd, A., Mohapatra, D. P., and Haroutounian, S. (2017). Neuropathic pain: central vs. peripheral mechanisms. *Curr. Pain Headache Rep.* 21:28. doi: 10.1007/s11916-017-0629-5
- Moalem, G., Xu, K., and Yu, L. (2004). T lymphocytes play a role in neuropathic pain following peripheral nerve injury in rats. *Neuroscience* 129, 767–777. doi: 10.1016/j.neuroscience.2004.08.035
- Mohajeri, M., Bianconi, V., Ávila-Rodriguez, M. F., Barreto, G. E., Jamialahmadi, T., Pirro, M., et al. (2020). Curcumin: a phytochemical modulator of estrogens and androgens in tumors of the reproductive system. *Pharmacol. Res.* 156:104765. doi: 10.1016/j.phrs.2020.104765
- Moldenhauer, S., Burgauner, M., Hellweg, R., Lun, A., Hohenböken, M., Dietz, E., et al. (2010). Mobilization of CD133(+)CD34(-) cells in healthy individuals following whole-body acupuncture for spinal cord injuries. *J. Neurosci. Res.* 88, 1645–1650. doi: 10.1002/jnr.22329
- Moon, J. Y., Song, S., Yoon, S. Y., Roh, D. H., Kang, S. Y., Park, J. H., et al. (2012). The differential effect of intrathecal Nav1.8 blockers on the induction and maintenance of capsaicin- and peripheral ischemia-induced mechanical allodynia and thermal hyperalgesia. *Anesth. Analg.* 114, 215–223. doi: 10.1213/ANE.0b013e318238002e
- Müller-Schwefe, G., Morlion, B., Ahlbeck, K., Alon, E., Coaccioli, S., Coluzzi, F., et al. (2017). Treatment for chronic low back pain: the focus should change to multimodal management that reflects the underlying pain mechanisms. *Curr. Med. Res. Opin.* 33, 1199–1210. doi: 10.1080/03007995.2017.1298521
- Murthy, S., Ryan, A., He, C., Mallampalli, R. K., and Carter, A. B. (2010). Rac1-mediated mitochondrial $\rm H_2O_2$ generation regulates MMP-9 gene expression in

macrophages via inhibition of SP-1 and AP-1. *J. Biol. Chem.* 285, 25062–25073. doi: 10.1074/jbc.M109.099655

- Nadeau, S., Filali, M., Zhang, J., Kerr, B. J., Rivest, S., Soulet, D., et al. (2011). Functional recovery after peripheral nerve injury is dependent on the pro-inflammatory cytokines IL-1 β and TNF: implications for neuropathic pain. *J. Neurosci.* 31, 12533–12542. doi: 10.1523/JNEUROSCI.2840-11.2011
- Nagase, H., Visse, R., and Murphy, G. (2006). Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc. Res.* 69, 562–573. doi: 10.1016/j.cardiores.2005.12.002
- Nascimento, G. C., De Paula, B. B., Gerlach, R. F., and CRA, L.-P. (2021). Temporomandibular inflammation regulates the matrix metalloproteinases MMP-2 and MMP-9 in limbic structures. *J. Cell. Physiol.* 236, 6571–6580. doi: 10.1002/jcp.30341
- Omair, A., Holden, M., Lie, B. A., Reikeras, O., and Brox, J. I. (2013). Treatment outcome of chronic low back pain and radiographic lumbar disc degeneration are associated with inflammatory and matrix degrading gene variants: a prospective genetic association study. *BMC Musculoskelet. Disord.* 14:105. doi: 10.1186/1471-2474-14-105
- Orestes, P., Osuru, H. P., Mcintire, W. E., Jacus, M. O., Salajegheh, R., Jagodic, M. M., et al. (2013). Reversal of neuropathic pain in diabetes by targeting glycosylation of ca(V)3.2 T-type calcium channels. *Diabetes* 62, 3828–3838. doi: 10.2337/db13-0813
- Osikowicz, M., Longo, G., Allard, S., Cuello, A. C., and Ribeiro-da-Silva, A. (2013). Inhibition of endogenous NGF degradation induces mechanical allodynia and thermal hyperalgesia in rats. *Mol. Pain* 9:37. doi: 10.1186/1744-8069-9-37
- Osthues, T., Zimmer, B., Rimola, V., Klann, K., Schilling, K., Mathoor, P., et al. (2020). The lipid receptor G2A (GPR132) mediates macrophage migration in nerve injury-induced neuropathic pain. *Cells* 9:1740. doi: 10.3390/cells9071740
- Pan, C., Wang, C., Zhang, L., Song, L., Chen, Y., Liu, B., et al. (2018). Procyanidins attenuate neuropathic pain by suppressing matrix metalloproteinase-9/2. *J. Neuroinflammation* 15:187. doi: 10.1186/s12974-018-1182-9
- Peng, B. G. (2013). Pathophysiology, diagnosis, and treatment of discogenic low back pain. *World J. Orthop.* 4, 42–52. doi: 10.5312/wjo.v4.i2.42
- Piccoliori, G., Engl, A., Gatterer, D., Sessa, E., in der Schmitten, J., and Abholz, H. H. (2013). Management of low back pain in general practice is it of acceptable quality: an observational study among 25 general practices in South Tyrol (Italy). *BMC Fam. Pract.* 14:148. doi: 10.1186/1471-2296-14-148
- Qorri, B., Kalaydina, R. V., Velickovic, A., Kaplya, Y., Decarlo, A., and Szewczuk, M. R. (2018). Agonist-biased Signaling via matrix metalloproteinase-9 promotes extracellular matrix remodeling. *Cells* 7:117. doi: 10.3390/cells7090117
- Remacle, A. G., Kumar, S., Motamedchaboki, K., Cieplak, P., Hullugundi, S., Dolkas, J., et al. (2015). Matrix metalloproteinase (MMP) proteolysis of the extracellular loop of voltage-gated sodium channels and potential alterations in pain signaling. *J. Biol. Chem.* 290, 22939–22944. doi: 10.1074/jbc.C115.671107
- Rendra, E., Riabov, V., Mossel, D. M., Sevastyanova, T., Harmsen, M. C., and Kzhyshkowska, J. (2019). Reactive oxygen species (ROS) in macrophage activation and function in diabetes. *Immunobiology* 224, 242–253. doi: 10.1016/j.imbio.2018.11.010
- Renton, T., Adey-Viscuso, D., Meechan, J. G., and Yilmaz, Z. (2010). Trigeminal nerve injuries in relation to the local anaesthesia in mandibular injections. *Br. Dent. J.* 209:E15. doi: 10.1038/sj.bdj.2010.978
- Rivera-Aponte, D. E., Méndez-González, M. P., Rivera-Pagán, A. F., Kucheryavykh, Y. V., Kucheryavykh, L. Y., Skatchkov, S. N., et al. (2015). Hyperglycemia reduces functional expression of astrocytic Kir4.1 channels and glial glutamate uptake. *Neuroscience* 310, 216–223. doi: 10.1016/j.neuroscience.2015.09.044
- Saleh, T., Naffa, R., Barakat, N. A., Ismail, M. A., Alotaibi, M. R., and Alsalem, M. (2024). Cisplatin provokes peripheral nociception and neuronal features of therapy-induced senescence and calcium dysregulation in rats. *Neurotox. Res.* 42:10. doi: 10.1007/s12640-024-00690-7
- Scholz, J., Finnerup, N. B., Attal, N., Aziz, Q., Baron, R., Bennett, M. I., et al. (2019). The IASP classification of chronic pain for ICD-11: chronic neuropathic pain. *Pain* 160, 53–59. doi: 10.1097/j.pain.000000000001365
- Shaefer, J. R., Khawaja, S. N., and Bavia, P. F. (2018). Sex, gender, and orofacial pain. Dent. Clin. N. Am. 62, 665–682. doi: 10.1016/j.cden.2018.06.001
- Sharshar, T., Durand, M. C., Lefaucheur, J. P., Lofaso, F., Raphaël, J. C., Gherardi, R. K., et al. (2002). MMP-9 correlates with electrophysiologic abnormalities in Guillain-Barré syndrome. *Neurology* 59, 1649–1651. doi: 10.1212/01.wnl.0000033799.58866.7a
- Shin, J. A., Kim, H. S., Vargas, A., Yu, W. Q., Eom, Y. S., Craft, C. M., et al. (2016). Inhibition of matrix metalloproteinase 9 enhances rod survival in the S334ter-line3 retinitis pigmentosa model. *PLoS One* 11:e0167102. doi: 10.1371/journal.pone.0167102
- Shubayev, V. I., and Myers, R. R. (2000). Upregulation and interaction of TNFalpha and gelatinases A and B in painful peripheral nerve injury. *Brain Res.* 855, 83–89. doi: 10.1016/s0006-8993(99)02321-5
- Singh, P., Bansal, S., Kuhad, A., Kumar, A., and Chopra, K. (2020). Naringenin ameliorates diabetic neuropathic pain by modulation of oxidative-nitrosative stress, cytokines and MMP-9 levels. *Food Funct.* 11, 4548–4560. doi: 10.1039/c9fo00881k
- Smith, B. H., and Torrance, N. (2012). Epidemiology of neuropathic pain and its impact on quality of life. *Curr. Pain Headache Rep.* 16, 191–198. doi: 10.1007/s11916-012-0256-0

- Sostres, C., Gargallo, C. J., and Lanas, A. (2013). Nonsteroidal anti-inflammatory drugs and upper and lower gastrointestinal mucosal damage. *Arthritis Res. Ther.* 15:S3. doi: 10.1186/ar4175
- Subedi, L., and Gaire, B. P. (2021). Tanshinone IIA: a phytochemical as a promising drug candidate for neurodegenerative diseases. *Pharmacol. Res.* 169:105661. doi: 10.1016/j.phrs.2021.105661
- Takeda, M., Tanimoto, T., Nasu, M., and Matsumoto, S. (2008). Temporomandibular joint inflammation decreases the voltage-gated K+ channel subtype 1.4-immunoreactivity of trigeminal ganglion neurons in rats. *Eur. J. Pain* 12, 189–195. doi: 10.1016/j.ejpain.2007.04.005
- Tesfaye, S., Sloan, G., Petrie, J., White, D., Bradburn, M., Young, T., et al. (2022). Optimal pharmacotherapy pathway in adults with diabetic peripheral neuropathic pain: the OPTION-DM RCT. *Health Technol. Assess.* 26, 1–100. doi: 10.3310/RXUO6757
- Tohge, T., and Fernie, A. R. (2017). Leveraging natural variance towards enhanced understanding of phytochemical sunscreens. *Trends Plant Sci.* 22, 308–315. doi: 10.1016/j.tplants.2017.01.003
- Tonello, R., Lee, S. H., and Berta, T. (2019). Monoclonal antibody targeting the matrix metalloproteinase 9 prevents and reverses paclitaxel-induced peripheral neuropathy in mice. *J. Pain* 20, 515–527. doi: 10.1016/j.jpain.2018.11.003
- Treede, R. D., Rief, W., Barke, A., Aziz, Q., Bennett, M. I., Benoliel, R., et al. (2019). Chronic pain as a symptom or a disease: the IASP classification of chronic pain for the international classification of diseases (ICD-11). *Pain* 160, 19–27. doi: 10.1097/j.pain.000000000001384
- Trescot, A. M., Chopra, P., Abdi, S., Datta, S., and Schultz, D. M. (2007). Systematic review of effectiveness and complications of adhesiolysis in the management of chronic spinal pain: an update. *Pain Physician* 10, 129–146.
- Umbricht, D. (2022). Matrix metalloproteinase 9 levels and parvalbumin positive interneuron dysfunction. *Neuropsychopharmacology* 47:429. doi: 10.1038/s41386-021-01048-9
- Van Hecke, O., Austin, S. K., Khan, R. A., Smith, B. H., and Torrance, N. (2014). Neuropathic pain in the general population: a systematic review of epidemiological studies. *Pain* 155, 654–662. doi: 10.1016/j.pain.2013.11.013
- Vandenbroucke, R. E., and Libert, C. (2014). Is there new hope for the rapeutic matrix metalloproteinase inhibition? *Nat. Rev. Drug Discov.* 13, 904–927. doi: 10.1038/nrd4390
- Wan, L., Guo, H., Hu, F., Pan, Y., Yang, S., Jiang, C. Y., et al. (2025). EZH2-mediated suppression of TIMP1 in spinal GABAergic interneurons drives microglial activation via MMP-9-TLR2/4-NLRP3 signaling in neuropathic pain. *Brain Behav. Immun.* 128, 234–255. doi: 10.1016/j.bbi.2025.04.007
- Wang, T., Jin, X., Liao, Y., Sun, Q., Luo, C., Wang, G., et al. (2018). Association of NF- κ B and AP-1 with MMP-9 overexpression in 2-chloroethanol exposed rat astrocytes. Cells 7:96. doi: 10.3390/cells7080096
- Wang, J., Lu, Z., Wu, C., Li, Y., Kong, Y., Zhou, R., et al. (2019). Evaluation of the anticancer and anti-metastasis effects of novel synthetic sodium channel blockers in prostate cancer cells *in vitro* and *in vivo*. *Prostate* 79, 62–72. doi: 10.1002/pros.23711
- Wang, A., Song, Z., Zhang, X., Xiao, L., Feng, Y., Qi, C., et al. (2023). MARS1 mutations linked to familial trigeminal neuralgia via the integrated stress response. *J. Headache Pain* 24:4. doi: 10.1186/s10194-022-01537-2
- Wang, Y., Wang, C., Yang, X., Ni, K., Jiang, L., Xu, L., et al. (2024). Inhibition of cyclophilin A-metalloproteinase-9 pathway alleviates the development of neuropathic pain by promoting repair of the blood-spinal cord barrier. *Anesth. Analg.* 138, 1313–1323. doi: 10.1213/ANE.00000000000006705
- Wang, W. J., Yang, W., Ouyang, Z. H., Xue, J. B., Li, X. L., Zhang, J., et al. (2018). MiR-21 promotes ECM degradation through inhibiting autophagy via the PTEN/akt/mTOR signaling pathway in human degenerated NP cells. *Biomed. Pharmacother.* 99, 725–734. doi: 10.1016/j.biopha.2018.01.154
- Wang, B., Zhang, L., and Yu, Y. (2022). Treatment of redo-microvascular decompression or internal neurolysis plus microvascular decompression for recurrent trigeminal neuralgia: a review of long-term effectiveness and safety. *J. Int. Med. Res.* 50:3000605221080721. doi: 10.1177/03000605221080721
- Wehling, M. (2014). Non-steroidal anti-inflammatory drug use in chronic pain conditions with special emphasis on the elderly and patients with relevant comorbidities: management and mitigation of risks and adverse effects. *Eur. J. Clin. Pharmacol.* 70, 1159–1172. doi: 10.1007/s00228-014-1734-6
- Wiera, G., Nowak, D., Van Hove, I., Dziegiel, P., Moons, L., and Mozrzymas, J. W. (2017). Mechanisms of NMDA receptor- and voltage-gated L-type calcium channel-dependent hippocampal LTP critically rely on proteolysis that is mediated by distinct metalloproteinases. *J. Neurosci.* 37, 1240–1256. doi: 10.1523/JNEUROSCI.2170-16.2016
- Will, J. S., Bury, D. C., and Miller, J. A. (2018). Mechanical low Back pain. $Am.\ Fam.\ Physician\ 98,\ 421-428.$
- Xia, L., Zhong, J., Zhu, J., Wang, Y. N., Dou, N. N., Liu, M. X., et al. (2014). Effectiveness and safety of microvascular decompression surgery for treatment of trigeminal neuralgia: a systematic review. *J. Craniofac. Surg.* 25, 1413–1417. doi: 10.1097/SCS.0000000000000984
- Yabe, Y., Hagiwara, Y., Ando, A., Tsuchiya, M., Minowa, T., Takemura, T., et al. (2015). Chondrogenic and fibrotic process in the ligamentum flavum of patients with lumbar process. The such that the sum of the process is the process of the proc

spinal canal stenosis. *Spine (Phila Pa 1976)* 40, 429–435. doi: 10.1097/BR\$.0000000000000795

Yamakawa, I., Kojima, H., Terashima, T., Katagi, M., Oi, J., Urabe, H., et al. (2011). Inactivation of TNF- α ameliorates diabetic neuropathy in mice. *Am. J. Physiol. Endocrinol. Metab.* 301:E844-52. doi: 10.1152/ajpendo.00029.2011

Yang, X., Jia, R., Hu, F., Fan, W., Lin, T., Zhang, X., et al. (2023). Promoting AMPK/ SR-A1-mediated clearance of HMGB1 attenuates chemotherapy-induced peripheral neuropathy. *Cell Commun. Signal* 21:99. doi: 10.1186/s12964-023-01100-9

Yao, M., Hasturk, H., Kantarci, A., Gu, G., Garcia-Lavin, S., Fabbi, M., et al. (2014). A pilot study evaluating non-contact low-frequency ultrasound and underlying molecular mechanism on diabetic foot ulcers. *Int. Wound J.* 11, 586–593. doi: 10.1111/iwj.12005

Yeh, C. M., Lin, C. W., Yang, J. S., Yang, W. E., Su, S. C., and Yang, S. F. (2016). Melatonin inhibits TPA-induced oral cancer cell migration by suppressing matrix metalloproteinase-9 activation through the histone acetylation. *Oncotarget* 7, 21952–21967. doi: 10.18632/oncotarget.8009

Yiemwattana, I., Kaomongkolgit, R., Wirojchanasak, S., and Chaisomboon, N. (2019). *Morus alba* stem extract suppresses matrix metalloproteinases (MMP)-1, MMP-9, and tissue inhibitors of metalloproteinase (TIMP)-1 expression via inhibition of IκΒα degradation induced by *Porphyromonas gingivalis* LPS signal in THP-1 cells. *Eur. J. Dent.* 13, 229–234. doi: 10.1055/s-0039-1694314

Yin, Y., Guo, R., Shao, Y., Ge, M., Miao, C., Cao, L., et al. (2019). Pretreatment with resveratrol ameliorate trigeminal neuralgia by suppressing matrix metalloproteinase-9/2 in trigeminal ganglion. *Int. Immunopharmacol.* 72, 339–347. doi: 10.1016/j.intimp.2019.04.014

Zakrzewska, J. M., Palmer, J., Morisset, V., Giblin, G. M., Obermann, M., Ettlin, D. A., et al. (2017). Safety and efficacy of a Nav1.7 selective sodium channel blocker in patients

with trigeminal neuralgia: a double-blind, placebo-controlled, randomised with drawal phase 2a trial. $Lancet\ Neurol.\ 16, 291-300.\ doi: 10.1016/S1474-4422(17)30005-4$

Zhang, H., and Dougherty, P. M. (2014). Enhanced excitability of primary sensory neurons and altered gene expression of neuronal ion channels in dorsal root ganglion in paclitaxel-induced peripheral neuropathy. *Anesthesiology* 120, 1463–1475. doi: 10.1097/ALN.000000000000176

Zhang, L., Li, X., Feng, X., Berkman, T., Ma, R., Du, S., et al. (2023). E74-like factor 1 contributes to nerve trauma-induced nociceptive hypersensitivity through transcriptionally activating matrix metalloprotein-9 in dorsal root ganglion neurons. *Pain* 164, 119–131. doi: 10.1097/j.pain.0000000000002673

Zhang, F. F., Morioka, N., Harano, S., Nakamura, Y., Liu, K., Nishibori, M., et al. (2016). Perineural expression of high-mobility group box-1 contributes to long-lasting mechanical hypersensitivity via matrix metalloprotease-9 up-regulation in mice with painful peripheral neuropathy. *J. Neurochem.* 136, 837–850. doi: 10.1111/inc.13434

Zhang, G., Zhang, W., Hou, Y., Chen, Y., Song, J., and Ding, L. (2018). Detection of miR-29a in plasma of patients with lumbar spinal stenosis and the clinical significance. *Mol. Med. Rep.* 18, 223–229. doi: 10.3892/mmr.2018.8956

Zhao, L., Song, C., Huang, Y., Lei, W., and Sun, J. (2020). MMP-9 regulates CX3CL1/CX3CR1 in the early phase of neuropathic pain in chronic sciatic nerve constriction injury (CCI) rats. *Ann. Palliat. Med.* 9, 2020–2027. doi: 10.21037/apm-20-1078

Zhou, F. T., Zong, Y., Li, Y. B., Cao, R. L., Hou, W. Q., Yang, F., et al. (2023). [Mechanism of Mongolian drug Naru-3 in initiation of neuroinflammation of neuropathic pain from MMP9/IL-1 β signaling pathway]. *Zhongguo Zhong Yao Za Zhi*, 48, 4173–4186.

\frown	C	c	2	K	1
u			а	1 \	,
	_	_			
	O	OS	OSS	ossa	ossary

AP-1 - Activator protein 1

AQP4 - Aquaporin 4

 $\beta\text{-}\mathbf{DG}$ - Beta-dystroglycan

CX3CL1 - C-X3-C motif chemokine ligand 1

CCI - Chronic constriction injury

CCI-IoN - Chronic constriction injury of the infraorbital nerve

CIPN - Chemotherapy-induced peripheral neuropathy

DRG - Dorsal root ganglion

ECM - Extracellular matrix

HMGB1 - High-mobility group box 1

IASP - International Association for the Study of Pain

IDD - Intervertebral disk degeneration

IENF - Intraepidermal nerve fibers

IL-1β - Interleukin-1 beta

IL-6 - Interleukin-6

LLLT - Low-level laser therapy

MMP-9 - Matrix metalloproteinase-9

MMPs - Matrix metalloproteinases

NGF - Nerve growth factor

NMDAR - N-methyl-D-aspartate receptors

NR1 - N-methyl-D-aspartate receptor subunit 1

NR2B - N-methyl-D-aspartate receptor subunit 2B

NSAIDs - Non-steroidal anti-inflammatory drugs

NF-κB - Nuclear factor kappa-light-chain-enhancer of activated B cells

PDN - Painful diabetic neuropathy

ROS - Reactive oxygen species

SASP - Senescence-associated secretory phenotype

SNC - Sciatic nerve crush

SNL - Spinal nerve ligation

TCAs - Tricyclic antidepressants

TIMPs - Tissue inhibitor of metalloproteinases

TN - Trigeminal neuralgia

TNF- α - Tumor necrosis factor-alpha

TRPV1 - Transient receptor potential vanilloid 1

Nav1.7 - Voltage-gated sodium channel subunit alpha-7