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Immunometabolism and male reproductive function: linking inflammation, oxidative stress, and declining fertility

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Background: Male infertility accounts for approximately 50% of all infertility cases, and its pathogenesis is highly complex. Beyond traditional factors such as genetics, endocrine disorders, and infections, growing evidence indicates that dysregulation of immunometabolism plays a pivotal role in the onset and progression of male reproductive dysfunction.

Objective: This review aims to systematically elucidate the role of immunometabolism in male reproductive health, focusing on the complex interplay among inflammation, oxidative stress, and metabolic imbalance. Additionally, it seeks to summarize potential therapeutic targets and outline future research directions.

Methods: A narrative review was conducted in accordance with the SANRA (Scale for the Assessment of Narrative Review Articles) guidelines. Relevant studies published between January 2010 to March 2025 were retrieved from PubMed, Embase, and Web of Science using keywords such as “immunometabolism,” “testis,” “male infertility,” and “oxidative stress.”

Results: Testicular immune homeostasis depends on the metabolic coordination among Sertoli cells, Leydig cells, and local immune cells. Aberrant immunometabolism disrupts the blood–testis barrier and endocrine balance by enhancing glycolysis, suppressing oxidative phosphorylation, and promoting the accumulation of reactive oxygen species (ROS), thereby impairing spermatogenesis and testosterone synthesis. Systemic metabolic inflammation induced by obesity, diabetes, and gut microbiota dysbiosis further exacerbates testicular dysfunction through the mTOR/HIF-1 α signaling axis and the “gut–immune–gonadal axis.” Pharmacological modulation of key immunometabolic regulators, including AMPK, SIRT1, and PPAR γ , has been shown to improve sperm quality and hormone levels in experimental models.

Conclusion: Immunometabolism serves as a crucial mechanistic bridge linking inflammation, oxidative stress, and the decline of male fertility. Future studies integrating multi-omics and spatial analysis technologies are expected to delineate immunometabolic phenotypes associated with male infertility, paving the way for precision diagnosis and personalized therapeutic interventions.

KEYWORDS

male infertility, immunometabolism, testicular immunity, oxidative stress, gut microbiota, metabolic signaling

1 Introduction

Male infertility accounts for approximately 50% of all infertility cases, and its pathogenesis is highly complex, being influenced by multiple factors including genetics, endocrine regulation, infections, environmental exposure, and immune dysfunction (1). In recent years, research has progressively shifted beyond the traditional focus on spermatogenic failure, hormonal abnormalities, and reproductive tract infections toward recognizing the interplay between the immune and metabolic systems—referred to as immunometabolism—as a key determinant of male reproductive health (2).

As an emerging interdisciplinary field, immunometabolism highlights the intimate connection between immune cell function and metabolic state, offering new insights into immune regulation and holding promise for the development of novel therapeutic strategies (3, 4). Metabolic pathways not only provide energy and biosynthetic precursors for immune cells but also directly dictate their activation, differentiation, and effector functions (5). Metabolic imbalances can drive the pro-inflammatory polarization of immune cells, leading to chronic inflammation and subsequent dysfunction of multiple organ systems, including the reproductive system (6).

Within the male reproductive system, the testis exhibits a unique immune privilege that prevents autoimmune responses against sperm antigens. The maintenance of testicular immune tolerance depends on the precise coordination of energy metabolism, redox balance, and lipid homeostasis. These interconnected metabolic networks shape immune cell phenotypes and are fundamental to sustaining reproductive immune stability (7) evidence suggests that disruption of these immunometabolic pathways impairs the testicular microenvironment, resulting in defective spermatogenesis, hormonal dysregulation, and compromised fertility (8).

This review systematically summarizes the mechanistic roles of immunometabolism in male reproductive health, focusing on the interplay between inflammation, oxidative stress, and metabolic imbalance. It further discusses potential therapeutic targets and future research directions.

2 Methods literature search strategy

This narrative review was conducted in accordance with the Scale for the Assessment of Narrative Review Articles (SANRA) guidelines to ensure methodological rigor. A systematic literature search identified 806 records. After a multi-stage screening process, 90 studies were included for qualitative synthesis (Figure 1).

A systematic literature search was performed to identify relevant publications spanning from January 2010 to March 2025. The primary databases interrogated were PubMed, Web of Science, and Embase. The search strategy employed a combination of the following keywords and MeSH terms: (“immunometabolism” OR “immune metabolism”) AND (“testis” OR “testicular” OR “male infertility” OR “spermatogenesis” OR “semen quality”) AND (“oxidative stress” OR “inflammation” OR “cytokine”) AND

(“metabolic pathway” OR “glycolysis” OR “OXPHOS” OR “mTOR” OR “AMPK” OR “SIRT1”).

Inclusion criteria encompassed: (1) original research articles (*in vivo*, *in vitro*) and high-quality reviews; (2) studies focusing on the interplay between immunometabolism and male reproductive function; (3) full-text articles available in English or Chinese.

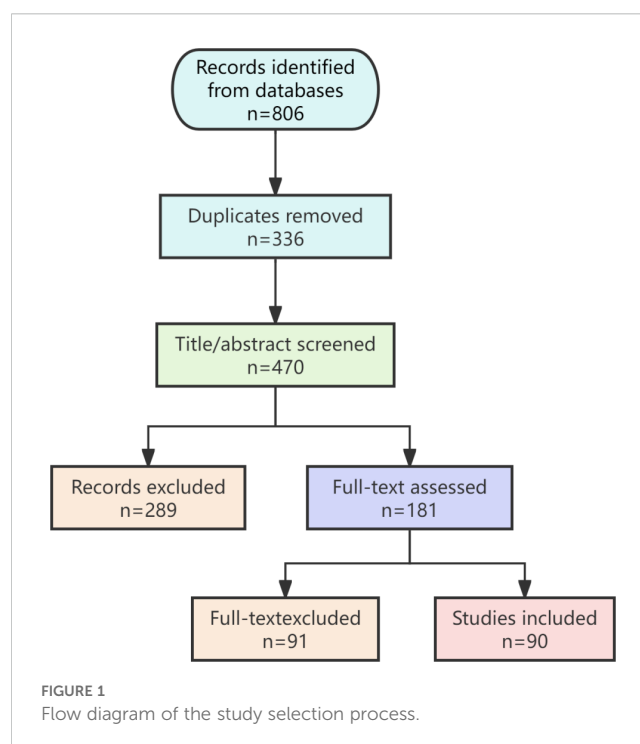
Exclusion criteria were: (1) conference abstracts, editorials, and non-peer-reviewed publications; (2) studies not primarily related to immunometabolic mechanisms; (3) case reports with limited generalizability.

The figure summarizes the literature identification, screening, eligibility, and inclusion steps according to the PRISMA guidelines, culminating in the 90 studies included for qualitative synthesis in this review.

3 Fundamental concepts and regulatory mechanisms of immunometabolism

3.1 Definition and research background

Immunometabolism is a rapidly evolving discipline at the intersection of immunology and metabolism, centered on the bidirectional regulatory relationship between metabolic pathways and immune cell function (9) immunology emphasizes cytokines, receptor signaling, and transcription factors as the primary regulators of immune cell activation, differentiation, and effector activity. However, recent evidence indicates that the metabolic programs of immune cells are not merely passive reflections of their functional states but active drivers of immune responses and cell fate decisions (10, 11).



3.1.1 Metabolic reprogramming in immune activation

Upon external stimulation, immune cells undergo metabolic reprogramming to rapidly adapt to diverse immune demands. This process not only provides energy and biosynthetic substrates but also influences immune cell fate through metabolic intermediates and signaling pathways (12).

3.1.2 Glycolytic dependency of pro-inflammatory cells

Pro-inflammatory cells (e.g. M1 macrophages, activated Th1/Th17 cells) predominantly rely on enhanced glycolysis to meet their high energy demands and generate metabolic intermediates, such as succinate, which support the synthesis of inflammatory mediators and the production of reactive oxygen species (ROS) (13).

3.1.3 Oxidative metabolism in anti-inflammatory and regulatory cells

In contrast, anti-inflammatory or immunosuppressive cells (e.g. M2 macrophages, Tregs) rely on oxidative phosphorylation (OXPHOS) and fatty acid oxidation (FAO) to maintain long-term energy stability and immune tolerance (14).

This metabolic reprogramming not only reflects but also directs the intensity and direction of immune responses. Under chronic inflammatory or metabolic disorder conditions, such as obesity and diabetes, excessive glycolytic signaling sustains low-grade inflammation, disrupting endocrine balance and reproductive function.

3.2 Key metabolic pathways

The regulation of immune cell function depends on the coordinated interaction of multiple energy and substrate metabolic pathways, which together form a complex “metabolic map” underlying immune activation, homeostasis, and disease defense (15).

3.2.1 Glycolysis

Although glycolysis is less energy-efficient, it enables rapid ATP generation under hypoxic or stress conditions, providing essential energy for short-term immune activation (16). In pro-inflammatory cells, upregulated glycolysis and succinate accumulation stabilize HIF-1 α , which promotes IL-1 β expression and amplifies inflammatory signaling—a central metabolic-inflammatory mechanism (17).

3.2.2 Tricarboxylic acid cycle and oxidative phosphorylation

Under anti-inflammatory or homeostatic conditions, immune cells maintain efficient coupling between the TCA cycle and OXPHOS to sustain cellular energy and support tissue repair (18). M2 macrophages and regulatory T (Treg) cells preserve intact mitochondrial metabolism, enhancing the secretion of IL-10 and other anti-inflammatory mediators to restrain inflammation (19).

3.2.3 Fatty acid metabolism (FAO/FAS)

Fatty acid oxidation (FAO) provides a long-term, sustained energy supply, particularly in anti-inflammatory or memory-type immune cells. In M2 macrophages, FAO supports tissue remodeling, whereas excessive fatty acid synthesis (FAS) promotes membrane remodeling and persistent immune activation (20, 21).

3.2.4 Amino acid metabolism

Glutamine serves as a crucial source of carbon and nitrogen, fueling the TCA cycle and nucleotide biosynthesis. Its metabolite, α -ketoglutarate, acts as a cofactor for epigenetic enzymes, regulating histone and DNA methylation, thereby influencing immune polarization and gene expression (22). Moreover, arginine and tryptophan metabolism modulate immune tolerance and inflammatory balance through substrate depletion and the generation of bioactive metabolites (e.g., kynurenines from tryptophan). By controlling the availability of these critical amino acids, these pathways act as essential ‘metabolic checkpoints’ that can decisively shift immune cell fate towards pro-inflammatory or tolerogenic states, thereby maintaining immune homeostasis (23).

3.3 Major regulatory molecules and signaling pathways

Dynamic immunometabolic regulation depends on several energy-sensing and signaling molecules.

3.3.1 mTOR

The mechanistic target of rapamycin (mTOR) is a critical serine/threonine kinase that functions as a central integrator of cellular energy and nutrient status. In immune cells, activation of the mTOR complex 1 (mTORC1) promotes glycolysis, lipid synthesis, and protein translation, thereby driving the activation, proliferation, and effector functions of pro-inflammatory cells such as M1 macrophages and Th17 cells (24, 25). Consequently, mTOR serves as a key molecular link connecting metabolic signaling to inflammatory activation.

3.3.2 AMPK

AMP-activated protein kinase (AMPK) functions as a cellular energy sensor, activated under conditions of low ATP availability. It restores energy homeostasis by promoting fatty acid oxidation (FAO), mitochondrial biogenesis, and autophagy, while simultaneously suppressing anabolic pathways. In the context of immunometabolism, AMPK activation favors the induction of anti-inflammatory phenotypes, such as M2 macrophages and regulatory T cells (Tregs), thereby contributing to the restoration of immune balance (26, 27).

3.3.3 HIF-1 α

Stabilized under hypoxic or inflammatory microenvironments, HIF-1 α enhances glycolysis and the production of pro-

inflammatory cytokines, creating a self-reinforcing metabolic-inflammatory loop (28).

3.3.4 PPAR γ and SIRT1

PPAR γ , a nuclear receptor, promotes anti-inflammatory phenotypes by activating fatty acid oxidation (FAO) and inhibiting the expression of pro-inflammatory genes (29). SIRT1, an NAD⁺-dependent deacetylase, regulates the acetylation of metabolic enzymes and transcription factors, thereby preserving mitochondrial function and enhancing antioxidant capacity. Together, they act synergistically to maintain immunometabolic balance (30).

4 Testicular immune microenvironment and metabolic homeostasis

The testis maintains immune tolerance and metabolic equilibrium through the blood–testis barrier (BTB), an immunosuppressive microenvironment, and the coordinated regulation of multiple cell types (31). This unique immune privilege protects haploid germ cells from autoimmune attack while ensuring efficient spermatogenesis. The maintenance of this privilege is closely linked to metabolic homeostasis, involving the coordinated functions of Sertoli cells, Leydig cells, germ cells, and resident immune cells (32).

4.1 Formation of immune privilege

To clarify the terminology used throughout this review, we define the following key concepts:

4.1.1 Testicular immune privilege

This refers to the anatomical and physiological characteristics of the testis that allow it to tolerate auto-antigenic germ cells without eliciting a destructive immune response. It is a property of the organ itself, established by physical barriers (e.g., the blood–testis barrier) and local immunosuppressive mechanisms (33, 34).

4.1.2 Immune tolerance

This describes the functional state of the immune system, specifically its acquired non-reactivity to specific antigens (in this case, sperm antigens) present within the privileged site (35).

4.1.3 Immunosuppressive environment/immune cold environment

These terms are used interchangeably in this text to describe the resultant local microenvironment within the testis, which is rich in anti-inflammatory cytokines (e.g., IL-10, TGF- β) and regulatory

immune cells (e.g., M2 macrophages, Tregs) that actively suppress effector immune responses (36, 37).

The establishment and maintenance of testicular immune privilege depend on the dual regulation of structural barriers and immune cell networks. The BTB, composed of tight junctions, adherens, and gap junctions between adjacent Sertoli cells, forms a specialized physical barrier that isolates developing germ cells from systemic immune surveillance (33, 34). Beyond its physical barrier function, local M2 macrophages, regulatory T (Treg) cells, dendritic cells (DCs), and a small number of natural killer (NK) cells secrete anti-inflammatory cytokines such as IL-10 and TGF- β , as well as metabolic regulatory mediators, to cooperatively sustain immune tolerance and local immune equilibrium (35).

4.2 Metabolic characteristics and functional differentiation of testicular cells

Different cell types within the testis exhibit distinct metabolic specializations that directly support their functional differentiation. Sertoli cells rely predominantly on glycolysis, even under normoxic conditions, preferentially metabolizing glucose into lactate to provide energy substrates for developing germ cells (38). Recent studies have shown that lactate acts not only as an energy source for developing germ cells but also as an intra-Sertoli cell signaling molecule that promotes cell survival and regulates oxidative stress responses (39). Leydig cells, in contrast, are metabolically centered on the mitochondrial conversion of cholesterol into testosterone, depending on oxidative phosphorylation (OXPHOS) and fatty acid β -oxidation to sustain energy and substrate supply (40). Cells such as macrophages, dendritic cells, and Treg cells maintain anti-inflammatory function and tissue homeostasis through a metabolic pattern characterized by low glycolysis, high fatty acid oxidation, and enhanced OXPHOS (41). This metabolic network collectively maintains the testicular microenvironment in a low-inflammatory and high-antioxidant state, facilitating continuous spermatogenesis and hormone synthesis. As summarized in Figure 2.

4.3 Interdependence between metabolic homeostasis and immune tolerance

Metabolic homeostasis is fundamental to testicular function and plays a crucial role in mediating immune tolerance. Disturbances in energy metabolism—such as oxidative stress, high-fat diets, and metabolic syndrome—can induce metabolic reprogramming of testicular immune cells, promoting their polarization from the M2 to the M1 phenotype. This repolarization is driven by specific signaling cues; for instance, pro-inflammatory signals like IFN- γ and LPS (via TLR4) promote the M1 state, while anti-inflammatory signals like IL-4 and IL-13 drive the M2 state, with

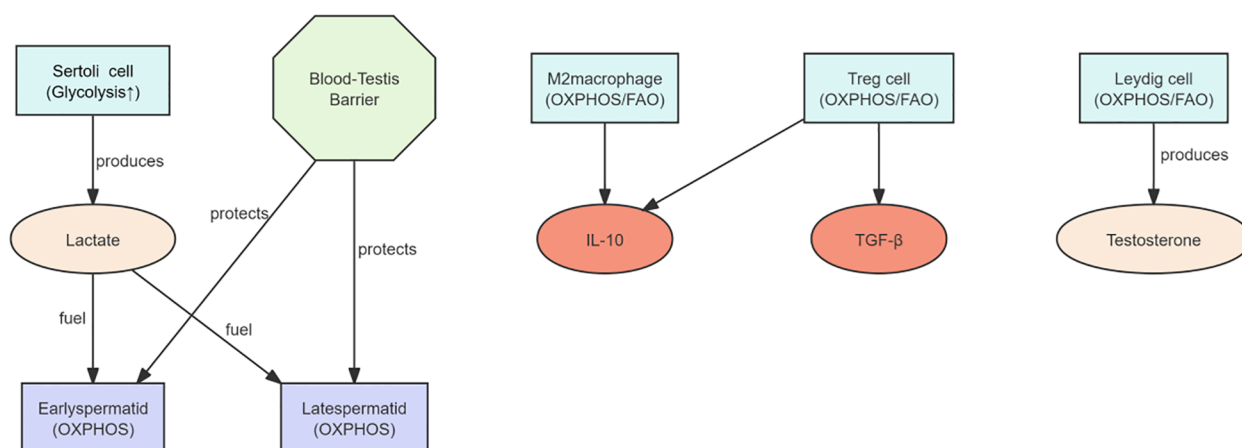


FIGURE 2

Cooperative interactions between anti-inflammatory immune cells and metabolic processes within the testicular immune microenvironment. In the testicular immune microenvironment, M2 macrophages and regulatory T cells secrete anti-inflammatory cytokines such as IL-10 and TGF- β to maintain local immune tolerance. This anti-inflammatory state is closely linked to specific metabolic patterns, including the high glycolytic activity of Sertoli cells, which produce lactate as an energy substrate for spermatogenesis and support testosterone synthesis in Leydig cells, thereby ensuring normal reproductive function.

each state being underpinned by distinct metabolic programs (glycolysis for M1, OXPHOS/FAO for M2) (42).

Inflammatory activation suppresses lactate production by Sertoli cells and testosterone synthesis by Leydig cells through multiple mechanisms, disrupting the positive feedback loop between metabolism and immunity. This disruption can contribute to impaired spermatogenesis and hormone deficiency, as observed in models of metabolic syndrome (43). The reciprocal promotion between metabolic imbalance and immune dysregulation has been identified as a central mechanism in the pathogenesis and progression of male infertility and varicocele (44).

Preclinical evidence suggests that strategies aimed at restoring local metabolic homeostasis—such as activating AMPK pathways, promoting fatty acid oxidation (FAO), or employing antioxidant therapies—represent promising avenues worthy of further investigation to improve male reproductive function (45).

5 Immune-metabolic imbalance and male infertility

Immune-metabolic imbalance serves as a common molecular basis for various male reproductive disorders. Recent studies indicate that metabolic abnormalities exacerbate inflammatory damage via immune pathways, forming the central pathological axis of “metabolism–immunity–reproduction” (46).

5.1 Inflammatory activation and metabolic reprogramming

Infections, environmental toxins, metabolic syndrome, and high-fat diets can induce metabolic reprogramming of immune

cells, shifting their metabolism from oxidative phosphorylation-dominant steady-state to glycolysis-dependent pro-inflammatory pathways. This shift exacerbates inflammatory responses and tissue damage by upregulating key signaling pathways, including HIF-1 α , mTOR (see Section 3.3.1), and NF- κ B. Activation of mTOR, as detailed in Section 3.3.1, promotes a glycolytic metabolic profile in testicular immune cells, sustaining the release of pro-inflammatory cytokines such as IL-6, TNF- α , and IL-1 β . This process establishes a chronic low-grade inflammatory state within the testis (47, 48). Excessive accumulation of lactate and reactive oxygen species (ROS) disrupts the blood-testis barrier through multiple signaling pathways, increasing immune cell infiltration and antigen exposure (49). Persistent immune activation not only damages germ cells but may also alter sperm DNA methylation patterns, leading to epigenetic reproductive harm.

5.2 Oxidative stress and mitochondrial damage

Oxidative stress serves as a critical link between metabolic disorders and reproductive dysfunction. Collectively, these events outline a core stress pathway in male infertility: metabolic disturbances (e.g., obesity and diabetes) and inflammation increase reactive oxygen species (ROS) generation, which directly damages sperm membranes, mitochondria, and DNA. At the same time, ROS activate stress-sensitive signaling pathways, such as p38 MAPK and NF- κ B, which further propagate cellular damage and inflammation, creating a vicious cycle that impairs reproductive function (50, 51). Mitochondrial damage activates caspase-9/3-dependent apoptosis, directly causing germ cell death and structural damage to the seminiferous tubules (52). Moreover, oxidative stress suppresses the expression of steroidogenic

enzymes in Leydig cells, further reducing testosterone levels and exacerbating endocrine imbalance (53).

5.3 Endocrine–immune–metabolic crosstalk

Male reproductive function depends heavily on precise regulation by the hypothalamic–pituitary–gonadal (HPG) axis, with metabolic hormones and immune signals forming a dynamic, multilayered network essential for reproductive health and systemic homeostasis (54). Insulin and leptin not only regulate energy metabolism but also modulate immune cell activation, differentiation, and function via the PI3K/Akt/mTOR signaling pathway (55). Insulin resistance induces M1 macrophage polarization, promoting chronic low-grade inflammation and the secretion of pro-inflammatory cytokines, thereby establishing a metabolism–immune positive feedback loop (56). Concurrently, inflammatory cytokines (e.g., IL-1 β , TNF- α) suppress GnRH and LH secretion, impair Leydig cell responsiveness, and cause hypogonadism (57). Low testosterone levels further inhibit Sertoli cell function and metabolic activity, disrupting the spermatogenic microenvironment and exacerbating male infertility (58). Thus, immunity, metabolism, and endocrine function form a mutually reinforcing pathological loop. Therefore, interventions aimed at breaking this vicious cycle—such as anti-inflammatory nutrition, AMPK activators, or metabolic remodeling drugs—are being investigated as potential strategies for treating male infertility.

6 Systemic immune-metabolic dysregulation and reproductive dysfunction

6.1 Bridging systemic immunometabolism and testicular dysfunction: convergent mechanisms of action

Systemic metabolic disorders, including obesity, type 2 diabetes, and gut microbiota dysbiosis, do not occur in isolation but collectively contribute to testicular dysfunction through shared immunometabolic pathways (59). This mechanistic connection is often referred to as the “systemic inflammation–metabolic disturbance–reproductive impairment axis,” which centers on the transmission of pro-inflammatory signals and metabolic intermediates from peripheral tissues to the testicular microenvironment (60).

6.1.1 Initiation of systemic inflammatory signaling

As illustrated in [Figure 3](#), this cascade involves the following key steps:

Initiation of Inflammatory Signaling: Obesity-induced adipose tissue inflammation or dysbiosis-driven gut inflammation increases circulating levels of pro-inflammatory cytokines (e.g., TNF- α , IL-6, IL-1 β) and bacterial components such as lipopolysaccharide (LPS).

6.1.2 Testicular signal transduction

These systemic factors impair the relative immune privilege of the testis through mechanisms such as compromising the integrity of the blood–testis barrier and activating testicular-resident immune cells (e.g., testicular macrophages) (61).

6.1.3 Intratesticular metabolic reprogramming

Nutrient-sensing pathways activated by hyperglycemia and hyperlipidemia, together with the local inflammatory environment, drive a metabolic shift within testicular cells (62).

Key regulatory components include:

Activation of the mTOR/HIF-1 α axis: Inflammatory cytokines and nutrients activate the mTOR and HIF-1 α pathways (for their fundamental functions, see Sections 3.3.1 and 3.3.3), promoting a metabolic shift toward glycolysis in both immune and somatic cells, thereby sustaining pro-inflammatory responses.

Suppression of AMPK/SIRT1 activity: The same inflammatory and metabolic stressors often inhibit AMPK and SIRT1 (for their fundamental functions, see Sections 3.3.2 and 3.3.4), reducing their anti-inflammatory, pro-oxidative phosphorylation, and antioxidant effects.

The following sections will detail how specific systemic disorders—obesity (Section 6.2), diabetes (Section 6.3), and gut microbiota dysbiosis (Section 6.4)—converge through this pathway to disrupt male reproductive function.

6.2 Obesity impairs spermatogenesis through a multi-faceted immunometabolic cascade

The expansion of adipose tissue, particularly visceral fat, leads to increased infiltration of pro-inflammatory M1 macrophages and the systemic release of cytokines such as TNF- α and IL-6 (63, 64). This chronic, low-grade inflammatory state disrupts the hypothalamic–pituitary–gonadal (HPG) axis and directly affects the testes. Within the testicular microenvironment, these inflammatory signals, combined with nutrient overload (e.g., high glucose and free fatty acids), promote a metabolic shift. This includes aberrant activation of the mTOR and NF- κ B pathways, which drive glycolysis and further pro-inflammatory responses in testicular cells (65). Concurrently, energy-sensing pathways such as AMPK are often suppressed, diminishing their anti-inflammatory and antioxidative effects. This metabolic reprogramming disrupts the function of Sertoli cells (compromising the blood–testis barrier) and Leydig cells (reducing testosterone synthesis), and directly induces oxidative stress and apoptosis in germ cells, ultimately leading to reduced sperm count and quality (66).

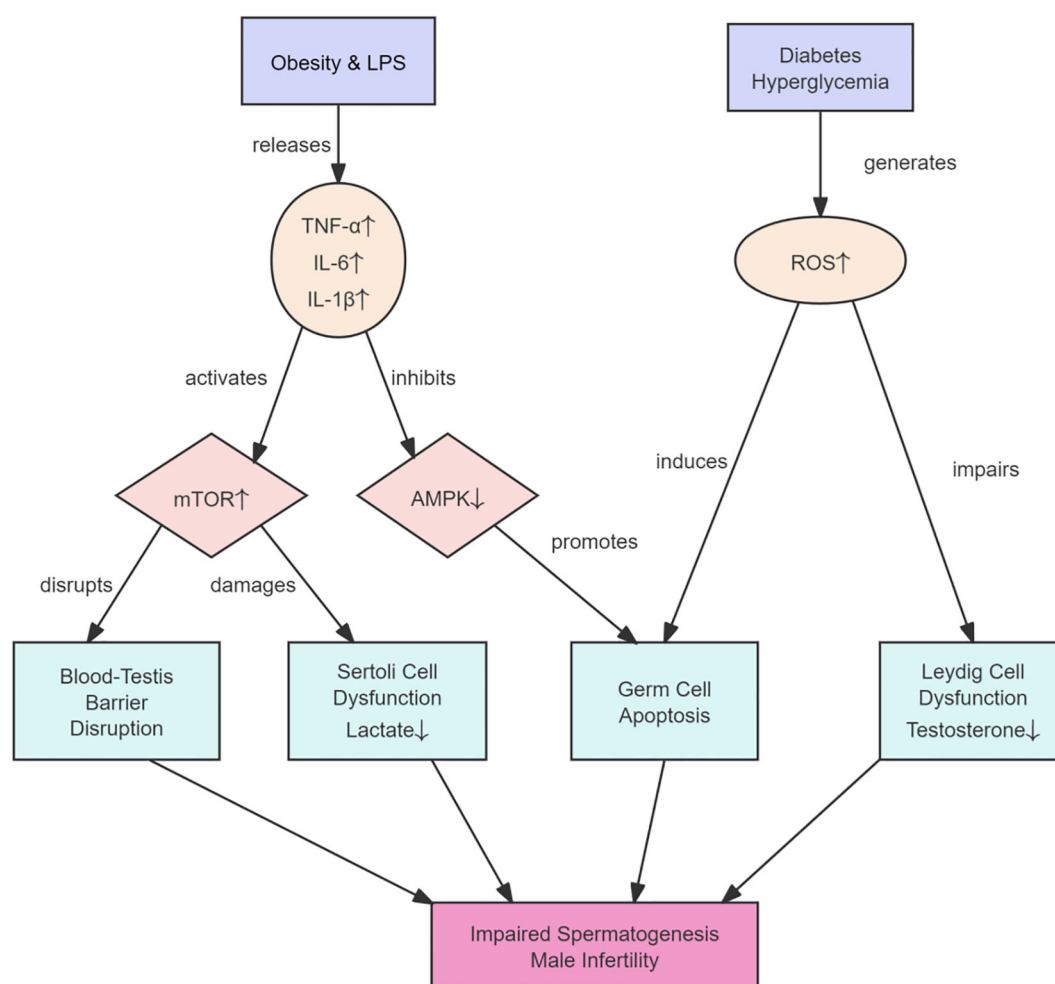


FIGURE 3

Mechanisms by which systemic metabolic disorders impair male reproductive function through inflammation and oxidative stress. Obesity, diabetes, and other systemic metabolic disorders cause adipose tissue to release pro-inflammatory cytokines such as IL-6 and TNF- α , initiating chronic low-grade systemic inflammation. These cytokines, along with hyperglycemia and lipid accumulation, increase ROS production, which impairs the function of Sertoli and Leydig cells and damages the integrity of the blood-testis barrier. This cascade ultimately reduces sperm quality and androgen synthesis, thereby compromising fertility.

6.3 Diabetes and reproductive function

Diabetes represents a typical state of combined metabolic and immune imbalance. Chronic hyperglycemia induces excessive ROS production, resulting in increased sperm DNA fragmentation and loss of mitochondrial membrane potential (67). Diabetes reprograms T cells and macrophages in the testis, promoting a glycolysis-dependent pro-inflammatory phenotype that secretes interleukin-1 β (IL-1 β) and interferon-gamma (IFN- γ), thereby exacerbating oxidative stress and apoptosis (68). Moreover, insulin/IGF-1 signaling critically regulates the expression of steroidogenic enzymes in Leydig cells and testosterone production; disruption of this signaling leads to reduced enzyme expression and decreased testosterone secretion (69). Animal studies confirm that diabetic metabolic abnormalities, through the coupling of immune inflammation and energy metabolism disorders, impair testicular mitochondrial function and

spermatogonial differentiation, directly affecting spermatogenesis (70).

6.4 Gut microbiota-immune-gonadal axis

The gut microbiome has recently emerged as a central hub linking metabolism and reproductive systems. Microbial metabolites, such as short-chain fatty acids and bile acids, regulate immune cell metabolism and function, potentially influencing hormonal regulation and reproductive health (71). A healthy microbiota composition promotes Treg and anti-inflammatory cytokine production, maintaining testicular immune homeostasis (72). Conversely, dysbiosis increases gut permeability, allowing lipopolysaccharides (LPS) and other inflammatory signals to enter the circulation, triggering immune responses and systemic inflammation (73). Clinical observations

reveal that obese and diabetic patients exhibit reduced gut microbial diversity, which correlates with decreased serum testosterone levels and abnormal semen parameters. This “gut-immune-gonadal axis” illustrates how peripheral metabolic abnormalities indirectly affect male reproductive function through immune pathways.

7 Potential therapeutic targets and interventions

7.1 Targeting signaling pathways

Several metabolic signaling pathways play a crucial role in maintaining testicular immune homeostasis, as illustrated in Figure 4.

7.1.1 mTOR pathway

Given its central role in driving pro-inflammatory metabolic programs, the overactive mTOR pathway observed in conditions of metabolic stress represents a rational therapeutic target (74). Inhibition of mTOR with rapamycin has been shown to reduce testicular oxidative stress and restore anti-inflammatory phenotypes in preclinical models (75). However, it should be noted that, as a potent immunosuppressant, prolonged or high-

dose administration of rapamycin may impair spermatogenesis. Multiple studies have demonstrated that rapamycin can induce spermatogenic arrest and disrupt the hypothalamic-pituitary-gonadal (HPG) axis. Therefore, its therapeutic window requires careful evaluation in the context of fertility treatment (76).

7.1.2 AMPK activation as a therapeutic strategy

As a critical energy sensor and promoter of oxidative metabolism (see Section 3.3.2), pharmacological activation of AMPK (e.g., by metformin) can counteract inflammation, enhance the cellular energy supply for spermatogenesis, and support the health of reproductive cells (77). Specifically, in the context of male infertility, AMPK activation is proposed to improve sperm quality through the following mechanisms: (1) reducing the production of pro-inflammatory cytokines in testicular macrophages; (2) alleviating oxidative stress in spermatogenic cells; (3) supporting the integrity of the blood-testis barrier; and (4) improving steroidogenesis in Leydig cells (45).

7.1.3 SIRT1 and PPAR γ

These molecules coordinate oxidative stress, mitochondrial homeostasis, and metabolism. Activation of SIRT1 enhances antioxidant defenses and mitochondrial function, while PPAR γ primarily regulates lipid metabolism and insulin sensitivity. Their

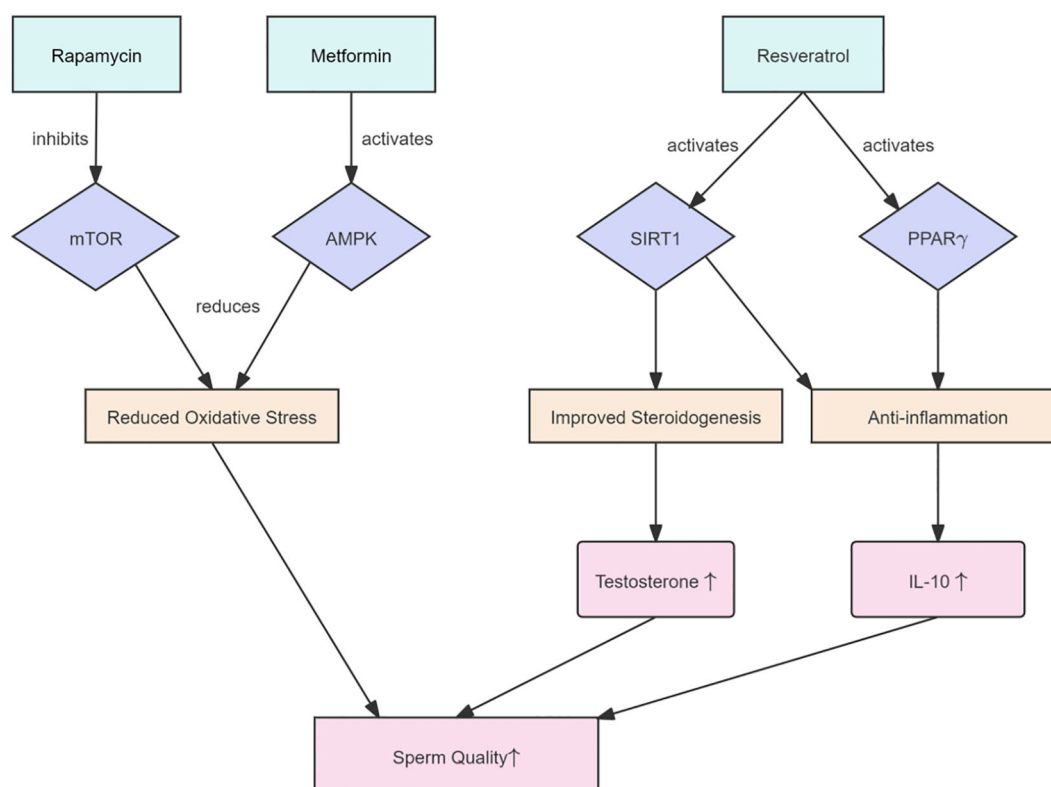


FIGURE 4

Intervention strategies targeting key immunometabolic signaling pathways to enhance male reproductive function. mTOR, AMPK, and SIRT1 are key regulators of immune-metabolic balance. Rapamycin inhibits overactive mTOR, thereby reducing inflammation; metformin, an AMPK activator, improves glucose and lipid metabolism while decreasing oxidative stress; and resveratrol activates SIRT1, enhancing antioxidant defenses and mitochondrial function. Targeting these pathways presents promising strategies to improve sperm quality and restore hormonal balance.

interplay provides a theoretical foundation for therapeutic targeting. Consequently, SIRT1 and PPAR γ have emerged as promising therapeutic targets for managing male infertility associated with obesity and metabolic syndrome (78, 79).

7.2 Pharmacological and nutritional interventions

This section discusses pharmacological and nutritional interventions aimed at correcting immunometabolic abnormalities, emphasizing their potential to enhance semen parameters, hormonal profiles, and the testicular microenvironment in cases of male infertility. Pharmacological interventions targeting immunometabolic dysfunction have demonstrated improvements in testicular inflammation and sperm parameters in animal models of obesity and diabetes. Metformin, an AMPK activator, has been demonstrated to reduce ROS, improve glucose and lipid metabolism, and support testosterone synthesis (80). However, existing research findings are contradictory; some clinical studies suggest that metformin may impair sperm mitochondrial function, leading to reduced sperm quality in certain male individuals. This underscores the importance of patient stratification and dose optimization in treating male infertility (81). Similarly, rapamycin can suppress mTOR and the release of pro-inflammatory cytokines (82), though its potential adverse effects on spermatogenesis warrant caution (as discussed in Section 7.1.1). Resveratrol, a natural polyphenol, has been shown to activate SIRT1 and exert antioxidant effects and has been shown to ameliorate immunometabolic dysregulation in preclinical studies (83). Furthermore, evidence supports that combining antioxidant supplementation with healthy lifestyle interventions—such as a balanced diet, moderate exercise, and probiotics—can synergistically reduce oxidative stress and systemic inflammation, which are known to be associated with improved sperm DNA integrity in some clinical studies (84).

7.3 Emerging research directions

Multi-omics approaches are poised to revolutionize our understanding of testicular immunometabolism. Single-cell RNA sequencing (scRNA-seq) can dissect the heterogeneity of testicular cells and precisely define the metabolic gene expression programs of immune, somatic, and germ cells in healthy and diseased states. Metabolomics can identify and quantify key metabolites (e.g., lactate, succinate, TCA cycle intermediates) within the testis, providing a functional readout of pathway activity and revealing potential diagnostic biomarkers (85, 86). When combined with spatial transcriptomics or proteomics, these techniques can map the precise location of these metabolic and immune states within the tissue architecture, thus revealing how cellular crosstalk within specific niches (e.g., seminiferous tubules) is governed by metabolism (85). Integrating these multi-omics datasets will enable the construction of comprehensive network models of testicular immunometabolism,

facilitating the development of precise diagnostic subtyping and personalized intervention strategies for male infertility.

8 Future perspectives

Research on immunometabolism provides a novel framework for understanding male reproductive disorders. Future work should transition from association to causation and clinical translation. Addressing these key areas is essential to bridge the gap between current observational knowledge and future clinically actionable insights.

8.1 Defining cell-type-specific metabolic vulnerabilities

Future studies must delineate how key pathways like mTOR, AMPK, and SIRT1 are differentially regulated in specific testicular cell types (e.g., Sertoli vs. Leydig vs. tissue-resident macrophages) under various infertile conditions. Utilizing cell-specific knockout models will be crucial to establish causal relationships and identify the most therapeutically relevant cellular targets (87).

8.2 High-resolution mapping of the testicular niche

The integration of single-cell and spatial multi-omics technologies (transcriptomics, metabolomics) is needed to create a high-resolution map of the testicular immunometabolic landscape. This will reveal novel cellular interactions, identify dysregulated metabolic checkpoints in patient subpopulations, and uncover biomarkers for infertility subtyping (88).

8.3 Translating metabolic reprogramming into therapies

While preclinical studies are promising, the safety, efficacy, and long-term reproductive outcomes of AMPK activators, SIRT1 modulators, and other metabolic drugs require rigorous evaluation in well-designed clinical trials. A critical future direction is to determine whether these interventions can reverse infertility in specific patient subgroups defined by their immunometabolic profile (89).

8.4 Exploring the gut–immune–gonadal axis

The role of the gut microbiota and its metabolites (e.g., short-chain fatty acids, bile acids) in regulating testicular immunity and

metabolism represents a frontier area. Research should focus on how specific microbial metabolites influence testicular function and whether interventions like probiotics or prebiotics can be developed as adjunct therapies for infertility (90).

9 Conclusion

Immune metabolism serves as a central link connecting inflammation, oxidative stress, and male reproductive function. The testis, as an immune-privileged organ, depends on coordinated energy metabolism and immune regulation. When metabolic imbalance and excessive immune activation create a positive feedback loop, oxidative damage, apoptosis, and endocrine disruption collectively impair spermatogenesis and hormone synthesis.

Preclinical evidence indicates that modulating immune-metabolic pathways—inhibiting mTOR, activating AMPK and SIRT1, and promoting fatty acid oxidation—effectively ameliorates inflammation-related and metabolic reproductive disorders in animal models. Integrating multi-omics data to define immune-metabolic subtypes of male infertility is expected to pave the way for more precise diagnostic and therapeutic interventions.

In summary, immune metabolism is a crucial focal point for elucidating the mechanisms of male infertility and a rapidly evolving frontier for fertility-preservation and precision-medicine strategies.

Author contributions

JZ: Methodology, Conceptualization, Writing – review & editing, Writing – original draft. SH: Software, Validation, Writing – review & editing. YO: Supervision, Writing – review & editing, Methodology. YK: Writing – review & editing,

Methodology, Resources. ML: Software, Conceptualization, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Kamiński P, Baszyński J, Jerzak I, Kavanagh BP, Nowacka-Chiari E, Polanin M, et al. External and genetic conditions determining male infertility. *Int J Mol Sci.* (2020) 21:5274. doi: 10.3390/ijms21155274
- Ye L, Huang W, Liu S, Cai S, Hong L, Xiao W, et al. Impacts of immunometabolism on male reproduction. *Front Immunol.* (2021) 12. doi: 10.3389/fimmu.2021.658432
- Makowski L, Chaib M, Rathmell J. Immunometabolism: From basic mechanisms to translation. *Immunol Rev.* (2020) 295:14–5. doi: 10.1111/imr.12858
- Patel CH, Leone RD, Horton MR, Powell JD. Targeting metabolism to regulate immune responses in autoimmunity and cancer. *Nat Rev Drug Discov.* (2019) 18:669–88. doi: 10.1038/s41573-019-0032-5
- Ma S, Wang X, Wang Y, Chen J, Li T, Wang F, et al. Cellular metabolism regulates the differentiation and function of T-cell subsets. *Cell Mol Immunol.* (2024) 21:419–35. doi: 10.1038/s41423-024-01148-8
- Deng H, Zhang Y, Li X, Chen L, Wang J, Liu Y, et al. Systematic low-grade chronic inflammation and intrinsic mechanisms in polycystic ovary syndrome. *Front Immunol.* (2024) 15. doi: 10.3389/fimmu.2024.1470283
- Christofides A, Konstantinidou E, Jani C, Boussiotis VA. The role of Peroxisome Proliferator-Activated Receptors (PPAR) in immune responses. *Metabol: Clin Exp.* (2020) 92:154338. doi: 10.1016/j.metabol.2020.154338
- Zhang M, Li Y, Zhou J, Wang H, Chen T, Liu K, et al. Transcription factor Dmrt1 triggers the SPRY1-NF- κ B pathway to maintain testicular immune homeostasis and male fertility. *Zool Res.* (2023) 44:505–21. doi: 10.24272/j.issn.2095-8137.2022.440
- O'Neill L, Kishton R, Rathmell J. A guide to immunometabolism for immunologists. *Nat Rev Immunol.* (2016) 16:553–65. doi: 10.1038/nri.2016.70
- Sun L, Fu W, Liu Y, Liu Y, Zhang C, Jiang Y, et al. Metabolic reprogramming in immune response and tissue inflammation. *Arteriosclerosis Thromb Vasc Biol.* (2020) 40:1990–2001. doi: 10.1161/ATVBAHA.120.314037
- Kelly B, O'Neill L. Metabolic reprogramming in macrophages and dendritic cells in innate immunity. *Cell Res.* (2015) 25:771–84. doi: 10.1038/cr.2015.68
- Chapman N, Chi H. Metabolic adaptation of lymphocytes in immunity and disease. *Immunity.* (2022) 55 1:14–30. doi: 10.1016/j.immuni.2021.12.012
- Soto-Hereder G, de Las Heras MM, Gabandé-Rodríguez E, Oller J, Mittelbrunn M. Glycolysis – a key player in the inflammatory response. *FEBS J.* (2020) 287:3350–69. doi: 10.1111/febs.15390
- Viola A, Munari F, Sánchez-Rodríguez R, Scolari T, Castegna A. The metabolic signature of macrophage responses. *Front Immunol.* (2019) 10. doi: 10.3389/fimmu.2019.01462

15. Ye L, Jiang Y-C, Zhang M. Crosstalk between glucose metabolism, lactate production and immune response modulation. *Cytokine Growth factor Rev.* (2022) 66:44–57. doi: 10.1016/j.cytogfr.2022.11.001
16. Kierans S, Taylor C. Glycolysis: A multifaceted metabolic pathway and signaling hub. *J Biol Chem.* (2024) 300:105656. doi: 10.1016/j.jbc.2024.107906
17. Lampropoulou V, Sergushichev A, Bambouskova M, Nair S, Vincent EE, Loginicheva E, et al. Itaconate links inhibition of succinate dehydrogenase with macrophage metabolic remodeling and regulation of inflammation. *Cell Metab.* (2016) 24:158–66. doi: 10.1016/j.cmet.2016.06.004
18. Martínez-Reyes I, Chandel N. Mitochondrial TCA cycle metabolites control physiology and disease. *Nat Commun.* (2020) 11:102. doi: 10.1038/s41467-019-13668-3
19. Ip WKE, Hoshi N, Shouval DS, Snapper S, Medzhitov R. Anti-inflammatory effect of IL-10 mediated by metabolic reprogramming of macrophages. *Science.* (2017) 356:513–9. doi: 10.1126/science.aal3535
20. Namgaladze D, Brüne B. Macrophage fatty acid oxidation and its roles in macrophage polarization and fatty acid-induced inflammation. *Biochim Biophys Acta.* (2016) 1861:1796–807. doi: 10.1016/j.bbalip.2016.09.002
21. Mills E, O'Neill L. Reprogramming mitochondrial metabolism in macrophages as an anti-inflammatory signal. *Eur J Immunol.* (2016) 46:102. doi: 10.1002/eji.201445427
22. Liu P-S, Wang H, Li X, Chao T, Christen S, Di Conza G, et al. α -ketoglutarate orchestrates macrophage activation through metabolic and epigenetic reprogramming. *Nat Immunol.* (2017) 18:985–94. doi: 10.1038/ni.3796
23. Solvay M, Ficht X, Lüscke J, Khaminejad S, Kral-O'Brien K, Gressier E, et al. Tryptophan depletion sensitizes the AHR pathway by increasing AHR expression and GCN2/LAT1-mediated kynurenine uptake, and potentiates induction of regulatory T lymphocytes. *J Immunother Cancer.* (2023) 11:e006728. doi: 10.1136/jitc-2023-006728
24. Linke M, Pham HTT, Katholnig K, Schnöller T, Miller A, Demel F, et al. mTORC1 and mTORC2 as regulators of cell metabolism in immunity. *FEBS Lett.* (2017) 591:3027–41. doi: 10.1002/1873-3468.12711
25. Weichhart T, Hengstschläger M, Linke M. Regulation of innate immune cell function by mTOR. *Nat Rev Immunol.* (2015) 15:599–614. doi: 10.1038/nri3901
26. Herzog S, Shaw R. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol.* (2017) 19:121–35. doi: 10.1038/nrm.2017.95
27. Cui Y, Wang Q, Li J, Zhang L, He Y, Li H, et al. The role of AMPK in macrophage metabolism, function and polarisation. *J Trans Med.* (2023) 21:837. doi: 10.1186/s12967-023-04772-6
28. Corcoran S, O'Neill L. HIF1 α and metabolic reprogramming in inflammation. *J Clin Invest.* (2016) 126:3699–707. doi: 10.1172/JCI84431
29. Gao Z, Li C, Li R, Liang Y, Zhang Y, Wang Y, et al. Mechanistic insight into PPAR γ and tregs in atherosclerotic immune inflammation. *Front Pharmacol.* (2021) 12:750078. doi: 10.3389/fphar.2021.750078
30. Tang B. Sirt1 and the mitochondria. *Mol Cells.* (2016) 39:87–95. doi: 10.14348/molcells.2016.2318
31. Dutta S, Sengupta P, Slama P, Roychoudhury S. Somatic-immune cells crosstalk in-the-making of testicular immune privilege. *Reprod Sci.* (2021) 29:2707–18. doi: 10.1007/s43032-021-00721-0
32. Qu N, Ogawa Y, Kuramasu M, Nagahori K, Sakabe K, Itoh M. Immunological microenvironment in the testis. *Reprod Med Biol.* (2019) 19:24–31. doi: 10.1002/rmb2.12293
33. Margret JJ, Jain S. The protective role of L-cysteine in the regulation of blood-testis barrier functions—A brief review. *Genes.* (2024) 15:852. doi: 10.3390/genes15070852
34. Luaces JP, Rossi SP, Matzkin ME, Terradas C, Ponzio R, Munuce MJ, et al. What do we know about blood-testis barrier? current understanding of its structure and physiology. *Front Cell Dev Biol.* (2023) 11. doi: 10.3389/fcell.2023.1114769
35. Liu J, Zhang X, Cheng Y, Cao X. Dendritic cell migration in inflammation and immunity. *Cell Mol Immunol.* (2021) 18:2461–71. doi: 10.1038/s41423-021-00726-4
36. O'Donnell L, Smith LB, Rebouret D, Wu Q, Cruickshanks L, Brown P, et al. Sertoli cell-enriched proteins in mouse and human testicular interstitial fluid. *PLoS One.* (2023) 18:e0290846. doi: 10.1371/journal.pone.0290846
37. Ma Y, Li Z, Zhao J, Zhang X, Wang T, Chen H, et al. Immunoregulation and male reproductive function: Impacts and mechanistic insights into inflammation. *Andrology.* (2024) 12:1021–35. doi: 10.1111/andr.13772
38. Ni F-D, Hao S-L, Yang W-X. Multiple signaling pathways in Sertoli cells: recent findings in spermatogenesis. *Cell Death Dis.* (2019) 10:897. doi: 10.1038/s41419-019-1782-z
39. Li J, Smith A, Johnson B, Davis K, Brown M, Wilson T, et al. L- and D-Lactate: unveiling their hidden functions in disease and health. *Cell Commun Signaling: CCS.* (2025) 23:89. doi: 10.1186/s12964-025-02132-z
40. Wang Y, Chen X, Li Z, Liu Y, Zhang Q, Yang J, et al. Steroidogenesis in Leydig cells: effects of aging and environmental factors. *Reproduction.* (2017) 154:R129–37. doi: 10.1530/REP-17-0064
41. Zhang S, Wang L, Li X, Chen Y, Liu Z, Yang H, et al. Efferocytosis fuels requirements of fatty acid oxidation and the electron transport chain to polarize macrophages for tissue repair. *Cell Metab.* (2019) 29:443–56. doi: 10.1016/j.cmet.2018.12.004
42. Chen Y, Zhang X, Wang J, Li H, Liu Y, Yang Z, et al. Mitochondrial metabolic reprogramming by CD36 signaling drives macrophage inflammatory responses. *Circ Res.* (2019) 125:1137–50. doi: 10.1161/CIRCRESAHA.119.315833
43. Mu Y, Li X, Wang J, Chen Y, Zhang Z, Liu H, et al. Diet-induced obesity impairs spermatogenesis: the critical role of NLRP3 in Sertoli cells. *Inflammation Regen.* (2022) 42:17. doi: 10.1186/s41232-022-00203-z
44. Tavalaei M, Ghaedi K, Nasr-Esfahani MH. The NLRP3 inflammasome: molecular activation and regulation in spermatogenesis and male infertility; a systematic review. *Basic Clin Androl.* (2022) 32:17. doi: 10.1186/s12610-022-00157-9
45. Saleh D, Ahmed K, Mohammed S, Ibrahim R, Hassan M, Abdelaziz A, et al. Eugenol alleviates acrylamide-induced rat testicular toxicity by modulating AMPK/p-AKT/mTOR signaling pathway and blood–testis barrier remodeling. *Sci Rep.* (2024) 14:10198. doi: 10.1038/s41598-024-52259-1
46. George BT, Miller A, Davis S, Clark J, Rodriguez M, Thompson L, et al. The molecular basis of male infertility in obesity: A literature review. *Int J Mol Sci.* (2023) 25:179. doi: 10.3390/ijms25010179
47. Zuo H, Wan Y. Metabolic reprogramming in mitochondria of myeloid cells. *Cells.* (2019) 9:1089. doi: 10.3390/cells9010005
48. Korbecki J, Kojder K, Simińska D, Bohatyrewicz R, Gutowska I, Chlubek D, et al. Chronic and cycling hypoxia: drivers of cancer chronic inflammation through HIF-1 and NF- κ B activation: A review of the molecular mechanisms. *Int J Mol Sci.* (2021) 22:10701. doi: 10.3390/ijms221910701
49. Wei Y, Wang D, Li Z, Chen J, Sun Y, Zhang H, et al. Polystyrene microplastics disrupt the blood-testis barrier integrity through ROS-Mediated imbalance of mTORC1 and mTORC2. *Environ pollut.* (2021) 289:117904. doi: 10.1016/j.envpol.2021.117904
50. Nowicka-Bauer K, Nixon B. Molecular changes induced by oxidative stress that impair human sperm motility. *Antioxidants.* (2020) 9:134. doi: 10.3390/antiox9020134
51. Saki M, Prakash A. DNA damage related crosstalk between the nucleus and mitochondria. *Free Radical Biol Med.* (2017) 107:216–27. doi: 10.1016/j.freeradbiomed.2016.11.050
52. Wang H, Li X, Zhang Y, Chen J, Liu Y, Wang Z, et al. Mechanism of Heshouwuyin inhibiting the Cyt c/Apaf-1/Caspase-9/Caspase-3 pathway in spermatogenic cell apoptosis. *BMC Complement Med Therapies.* (2020) 20:265. doi: 10.1186/s12906-020-02904-9
53. Hu J, Chen L, Wang Y, Li X, Zhang Q, Yang H, et al. Anthocyanins prevent AAPH-induced steroidogenesis disorder in leydig cells by counteracting oxidative stress and sTAR abnormal expression in a structure-dependent manner. *Antioxidants.* (2023) 12:508. doi: 10.3390/antiox12020508
54. Ma Y, Zhang X, Li Z, Wang T, Chen H, Liu Y, et al. Critical illness and sex hormones: response and impact of the hypothalamic–pituitary–gonadal axis. *Ther Adv Endocrinol Metab.* (2025) 16:204201882513281. doi: 10.1177/20420188251328192
55. Dibble C, Cantley L. Regulation of mTORC1 by PI3K signaling. *Trends Cell Biol.* (2015) 25:545–55. doi: 10.1016/j.tcb.2015.06.002
56. Castoldi A, Naffah de Souza C, Câmara NOS, Moraes-Vieira PM. The macrophage switch in obesity development. *Front Immunol.* (2016) 6. doi: 10.3389/fimmu.2015.00637
57. Bini EI, D'Attilio L, Marquina-Castillo B, Mata-Espinosa D, Bay ML, del Sasiain M, et al. The implication of pro-inflammatory cytokines in the impaired production of gonadal androgens by patients with pulmonary tuberculosis. *Tuberculosis.* (2015) 95:701–6. doi: 10.1016/j.tube.2015.06.002
58. Zhang X-N, Li Y, Wang Y, Chen T, Liu K, Zhou J, et al. Ldh-dependent metabolic programs in sertoli cells regulate spermiogenesis in mouse testis. *Biology.* (2022) 11:1791. doi: 10.3390/biology11121791
59. Daryabor G, Atashzar MR, Kabelitz D, Meri S, Kalantar K. The effects of type 2 diabetes mellitus on organ metabolism and the immune system. *Front Immunol.* (2020) 11. doi: 10.3389/fimmu.2020.01582
60. Ling Q, Wang J, Chen Y, Zhang H, Li X, Liu Z, et al. Analysis of the inflammatory gene expression characteristics and immune microenvironment regulatory mechanisms in the testicular tissue of patients with non-obstructive azoospermia. *PLoS One.* (2025) 20:e0324948. doi: 10.1371/journal.pone.0324948
61. Peirouvi T, Hajipour H, Farjah GH, Karimi A, Moghadaszadeh M. COVID-19 disrupts the blood–testis barrier through the induction of inflammatory cytokines and disruption of junctional proteins. *Inflammation Res.* (2021) 70:1165–75. doi: 10.1007/s00111-021-01497-4
62. Newsholme P, Cruzat VF, Keane KN, Carlessi R, de Bittencourt PIH Jr. Metabolic adaptations/reprogramming in islet beta-cells in response to physiological stimulators—What are the consequences. *Antioxidants.* (2022) 11:108. doi: 10.3390/antiox11010108
63. Li X, Zhang Y, Wang J, Chen T, Liu H, Yang Z, et al. Adipose tissue macrophages as potential targets for obesity and metabolic diseases. *Front Immunol.* (2023) 14. doi: 10.3389/fimmu.2023.1153915
64. Xu L, Ma Y, Zhang J, Wang H, Li X, Chen Y, et al. Macrophage polarization mediated by mitochondrial dysfunction induces adipose tissue inflammation in obesity. *Int J Mol Sci.* (2022) 23:9252. doi: 10.3390/ijms23169252

65. Luo D, Liu Y, Wang H, Li Z, Chen X, Zhang Y, et al. High fat diet impairs spermatogenesis by regulating glucose and lipid metabolism in Sertoli cells. *Life Sci.* (2020) 256:118028. doi: 10.1016/j.lfs.2020.118028
66. Liu Y, Ding Z. Obesity, a serious etiologic factor for male subfertility in modern society. *Reproduction.* (2017) 154:4. doi: 10.1530/REP-17-0161
67. Zhang Z, Li X, Wang Y, Chen J, Liu Y, Zhang H, et al. The impact of oxidative stress-induced mitochondrial dysfunction on diabetic microvascular complications. *Front Endocrinol.* (2023) 14. doi: 10.3389/fendo.2023.1112363
68. Wang F, Zhang S, Jeon R, Vuckovic I, Jiang X, Lerman A, et al. Interferon gamma induces reversible metabolic reprogramming of M1 macrophages to sustain cell viability and pro-inflammatory activity. *EBioMedicine.* (2018) 30:303–16. doi: 10.1016/j.ebiom.2018.02.009
69. Neirijnck Y, Papaioannou MD, Nef S. Insulin and IGF1 receptors are essential for the development and steroidogenic function of adult Leydig cells. *FASEB J.* (2018) 32:3321–35. doi: 10.1096/fj.201700769RR
70. Mu Y, Li X, Wang J, Chen Y, Zhang Z, Liu H, et al. Bezafibrate alleviates diabetes-induced spermatogenesis dysfunction by inhibiting inflammation and oxidative stress. *Heliyon.* (2024) 10:e28284. doi: 10.1016/j.heliyon.2024.e28284
71. Wang J, Chen Y, Zhang H, Li X, Liu Z, Yang H, et al. Gut-microbiota-derived metabolites maintain gut and systemic immune homeostasis. *Cells.* (2023) 12:793. doi: 10.3390/cells12050793
72. Sharma A, Sharma G, Im S-H. Gut microbiota in regulatory T cell generation and function: mechanisms and health implications. *Gut Microbes.* (2025) 17:2457143. doi: 10.1080/19490976.2025.2516702
73. Di Vincenzo F, Del Gaudio A, Petito V, Lopetuso LR, Scaldaferrri F. Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Internal Emergency Med.* (2023) 19:275–93. doi: 10.1007/s11739-023-03374-w
74. Kaldirim M, Lang J, Ehnert S, Nussler AK. Modulation of mTOR signaling in cardiovascular disease to target acute and chronic inflammation. *Front Cardiovasc Med.* (2022) 9. doi: 10.3389/fcvm.2022.907348
75. Wang Y, Li M, Zha A. mTOR promotes an inflammatory response through the HIF1 signaling pathway in ulcerative colitis. *Int Immunopharmacol.* (2024) 134:112217. doi: 10.1016/j.intimp.2024.112217
76. Yang J, Li X, Wang Y, Chen T, Zhang H, Liu Z, et al. Rapamycin ameliorates radiation-induced testis damage in mice. *Front Cell Dev Biol.* (2022) 10. doi: 10.3389/fcell.2022.783884
77. Liu R, Wang H, Zhang Y, Chen J, Li X, Yang Z, et al. Spermidine endows macrophages anti-inflammatory properties by inducing mitochondrial superoxide-dependent AMPK activation, Hif-1 α upregulation and autophagy. *Free Radical Biol Med.* (2020) 161:339–50. doi: 10.1016/j.freeradbiomed.2020.10.029
78. Alam F, Syed H, Amjad S, Baig M, Khan TA, Rizvi A. Interplay between oxidative stress, SIRT1, reproductive and metabolic functions. *Curr Res Physiol.* (2021) 4:119–24. doi: 10.1016/j.crphys.2021.03.002
79. Janani C, Kumari R. PPAR gamma gene—a review. *Diabetes Metab Syndrome.* (2015) 9:46–50. doi: 10.1016/j.dsx.2014.09.015
80. Wang Y, An H, Liu T, Qin C, Sesaki H, Guo S, et al. Metformin improves mitochondrial respiratory activity through activation of AMPK. *Cell Rep.* (2019) 29:1511–23. doi: 10.1016/j.celrep.2019.09.070
81. Leisegang K, Almaghrabi W, Henkel R. The effect of Nigella sativa oil and metformin on male seminal parameters and testosterone in Wistar rats exposed to an obesogenic diet. *Biomed Pharmacother = Biome Pharmacother.* (2021) 133:111085. doi: 10.1016/j.biopha.2020.111085
82. Cappoli N, Tabolacci E, De Chiara G, Scala E, Penta R, Nardone AM, et al. The mTOR kinase inhibitor rapamycin enhances the expression and release of pro-inflammatory cytokine interleukin 6 modulating the activation of human microglial cells. *EXCLI J.* (2019) 18:779–98. doi: 10.17179/excli2019-1814
83. Ciccone L, Nencetti S, Socci S, Sheppard D, Nuti E, Orlandini E, et al. Resveratrol-like compounds as SIRT1 activators. *Int J Mol Sci.* (2022) 23:15105. doi: 10.3390/ijms232315105
84. Lahimer M, Montjean D, Cabry R, Bach V, Benkhalifa M, Mauvieux L, et al. Micronutrient–antioxidant therapy and male fertility improvement during ART cycles. *Nutrients.* (2025) 17:324. doi: 10.3390/nu17020324
85. Longo SK, Guo MG, Ji AL, Khavari PA. Integrating single-cell and spatial transcriptomics to elucidate intercellular tissue dynamics. *Nat Rev Genet.* (2021) 22:627–44. doi: 10.1038/s41576-021-00370-8
86. Ferreira RM, Sabo AR, Winfree S, Collins KS, Janosevic D, Gulbranson CJ, et al. Integration of spatial and single-cell transcriptomics localizes epithelial cell–immune cross-talk in kidney injury. *JCI Insight.* (2021) 6:e147703. doi: 10.1172/jci.insight.147703
87. Yu P, Li X, Zhang Y, Chen J, Wang H, Liu Y, et al. The microRNA-mediated apoptotic signaling axis in male reproduction: a possible and targetable culprit in male infertility. *Cell Biol Toxicol.* (2025) 41:15. doi: 10.1007/s10565-025-10006-w
88. Sanches P, Azevedo V, Rocha C, Lima A, Oliveira E, Santos R, et al. Integrating molecular perspectives: strategies for comprehensive multi-omics integrative data analysis and machine learning applications in transcriptomics, proteomics, and metabolomics. *Biology.* (2024) 13:848. doi: 10.3390/biology13110848
89. Day E, Ford R, Steinberg G. AMPK as a therapeutic target for treating metabolic diseases. *Trends Endocrinol Metab.* (2017) 28:545–60. doi: 10.1016/j.tem.2017.05.004
90. Chen W, Zhang Y, Li X, Wang J, Liu Z, Yang H, et al. The potential influence and intervention measures of gut microbiota on sperm: it is time to focus on testis-gut microbiota axis. *Front Microbiol.* (2024) 15:1478082. doi: 10.3389/fmicb.2024.1478082