

OPEN ACCESS

EDITED BY Imene Soufli, University of Science and Technology Houari Boumediene, Algeria

REVIEWED BY Suzana Stanisavljević, University of Belgrade, Serbia Samiya Bessad, Institut Pasteur d'Algérie, Algeria

*CORRESPONDENCE
Luca Masucci
Luca.masucci@unicatt.it

[†]These authors have contributed equally to this work and share first authorship

[‡]These authors have contributed equally to this work and share senior authorship

RECEIVED 03 August 2025 REVISED 10 November 2025 ACCEPTED 11 November 2025 PUBLISHED 26 November 2025

CITATION

Bibbò S, De Maio F, Capone F, Quaranta G, Rondinella D, Rosato R, Minelli M, De Lorenzis D, Sanguinetti M, Cammarota G, Di Lazzaro V and Masucci L (2025) Case Report: Fecal microbiota transplantation via capsules ameliorated clinical outcomes in a patient with multiple sclerosis. *Front. Immunol.* 16:1678759. doi: 10.3389/fimmu.2025.1678759

COPYRIGHT

© 2025 Bibbò, De Maio, Capone, Quaranta, Rondinella, Rosato, Minelli, De Lorenzis, Sanguinetti, Cammarota, Di Lazzaro and Masucci. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Case Report: Fecal microbiota transplantation via capsules ameliorated clinical outcomes in a patient with multiple sclerosis

Stefano Bibbò^{1†}, Flavio De Maio^{2†}, Fioravante Capone^{3†}, Gianluca Quaranta^{2†}, Debora Rondinella¹, Roberto Rosato², Mauro Minelli⁴, Desy De Lorenzis², Maurizio Sanguinetti^{2,3,4,5}, Giovanni Cammarota^{1,2,3,4,5,6‡}, Vincenzo Di Lazzaro^{3,4,5,6,7‡} and Luca Masucci^{2,3,4,5*‡}

¹Department of Medical and Surgical Sciences, Unità Operativa Complessa (UOC) Gastroenterologia, Fondazione Policlinico Universitario Gemelli Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS), Rome, Italy, ²Department of Laboratory and Haematological Sciences, Fondazione Policlinico Universitario A. Gemelli Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS), Rome, Italy, ³Research Unit of Neurology, Department of Medicine and Surgery, Università Campus Bio-Medico di Roma, Via Alvaro del Portillo, Roma, Italy, ⁴Department of Legal and Business Sciences. Università Libera Università Mediterranea (LUM) Giuseppe Degennaro. Casamassima, Bari, Italy, ⁵Department of Basic Biotechnological Sciences, Intensive and Perioperative Clinics, Università Cattolica del Sacro Cuore, Rome, Italy, ⁶Department of Translational Medicine and Surgery, Università Cattolica del Sacro Cuore, Rome, Italy, ⁷Fondazione Policlinico Universitario Campus Bio-Medico, Via Alvaro del Portillo, Rome, Italy

Multiple Sclerosis (MS) has long been recognized as a multifactorial disease, associated with both genetic and environmental factors. Its link to inflammatory processes has led to significant advances in understanding the immunological and neurobiological mechanisms underlying the disease. The presumed autoimmune etiology is currently guiding the development of therapeutic protocols in this direction. The intestinal bacteria population, known as Gut Microbiota (GM), plays a well-documented role in autoimmune and inflammatory diseases. Gut microbiota dysbiosis is associated in patients affected by MS. Fecal Microbiota Transplantation (FMT) is emerging as a potential strategy to restore eubiosis and modulate systemic inflammation. We treated a 42-year-old woman with severe MS disability by FMT via colonoscopic infusion followed by a 3-month treatment of daily oral capsules, containing frozen microbiota, in order to resolve gastrointestinal symptoms. Clinical followup was conducted at 30, 60, and 90 days. Microbiota profiling (16S rRNA sequencing) and intestinal permeability testing were performed at several time points before and post infusion. Post-FMT, gastrointestinal symptoms improved significantly, as well as in limb spasticity, trunk stability, and fine motor skills. Microbiota analysis revealed a marked reduction in the abundance of Akkermansia muciniphila (22.5% vs 6.6%). At phylum level, Actinomycetota remained elevated (22%), while Bacteroidota consistently showed low abundance (14%). The most favorable microbiota profile was observed at 90 days, which coincided with the peak of clinical improvement. Intestinal permeability also improved over time, despite the patient's microbiota profile

remaining distinct from the donor. This is the first report about combined FMT in MS. The procedure was safe, well tolerated, and associated with both gastrointestinal and neurological improvements. These findings support further exploration of FMT as a therapeutic adjunct in MS through controlled clinical trials.

KEYWORDS

case report, multiple sclerosis, fecal microbiota transplantation, autoimmunity, bacterial therapy

Introduction

Multiple sclerosis (MS) is a disease of the central nervous system (CNS), also associated to autoimmune dysregulation. For this reason, recent therapeutic protocols include treatments targeting the underlying mechanisms (1). The intestinal bacterial population, gut microbiota (GM), plays a key role in immunomodulation (2). The GM profile is altered in MS (3). In recent years, the fecal microbiota transplantation (FMT) has proven to be a valid therapeutic option for the restoring of intestinal eubiosis in *Clostridioides difficile* (4) infection and in autoimmune diseases (2). MS patients also report intestinal discomfort which may be due to both alterations in bacterial composition and neurological involvement (5).

Basing on our findings, the efficacy of FMT in patients with MS has documented in only a few cases (6). We report the first case of a patient affected by MS treated using a combined FMT approach via colonoscopy and oral capsules, aimed at alleviating gastrointestinal symptoms. To assess the efficacy of the procedure, next-generation sequencing (NGS) was performed on fecal samples collected before the infusion of microbiota and over a five-month period. Moreover, clinical assessments were carried out pre-FMT and at monthly intervals for three months. This interventional approach was effective in normalizing intestinal function and improving the patient's neurological status.

Case

We describe a 42-year-old female affected by secondary progressive MS (SPMS). Disease onset was in 2009 with lower limbs impairment. Since 2018, she was restricted to wheelchair. The patient underwent multiple treatment protocols, including interferon β -1A, Natalizumab (Tysabri[®]), Teriflunomide (Aubagio[®]), Methotrexate, and Simvastatin (for its potential neuroprotective effects), all of which were discontinued due to lack of clinical efficacy or poor tolerability. She is currently not receiving any immunosuppressive therapy and has not been under such treatment for approximately 10 years; furthermore, she

did not resume any immunosuppressive therapy after the infusion procedure.

The most recent brain MRI showed stable multiple demyelinating lesions without Gadolinium-enhancement.

From an immuno-allergological perspective, the patient had reduced MTHFR enzymatic activity and a positive HLA-DQ7 haplotype, factors that may predispose to intestinal inflammation (7) and food intolerances (8). Documented intolerances to nickel sulfate and cobalt chloride are associated with clinical symptoms following metal exposure.

The patient reported longstanding chronic gastrointestinal symptoms, including postprandial abdominal cramping, foul-smelling flatulence, urgent bowel movements, and altered bowel habits. Additional symptoms included dyspepsia, intermittent constipation, and delayed gastric emptying.

She experienced systemic and vascular symptoms, such as cold extremities, thermal dysregulation, muscle stiffness, fatigue, slowed movements, and neurovegetative disturbances.

The patient underwent fecal microbiota transplantation (FMT) through the following protocol: an initial infusion was administered via colonoscopy using a 200 mL solution derived from a healthy donor. Starting the day after the infusion, the patient began oral administration of frozen capsules containing microbiota from the same donor, at a dosage of three capsules per day (10⁶–10⁹ CFU/ mL) for a duration of three months. (Table 1). No antibiotic or probiotic therapy was administered to the patient in the five months prior to the procedure.

Donor selection

The donor selection phase for fecal microbiota transplantation (FMT) consists of a multi-step screening process (9). Donors are initially evaluated through an anamnesis-based questionnaire aimed at excluding risk factors for transmissible diseases and relevant comorbidities, including neurological diseases. Subsequently, the potential donor undergoes a rigorous series of serological and microbiological screening tests performed on blood and stool samples.

TABLE 1 Clinical follow-up and FMT administration.

Time point	Functional/clinical notes	FMT administration
Pre-FMT Evaluation	Severe impairment, limited passive mobility; high disability	Initial infusion via colonoscopy (200 mL donor microbiota solution); start of oral capsules 3/day (10^6–10^9 CFU/mL) from day after infusion
30-Day Follow- up Post-FMT	Initial clinical improvement; standing more stable; physiotherapy ongoing	Continuation of oral capsules 3/day
60-Day Follow- up Post-FMT	Improved autonomy in fine motor tasks; independence in some daily activities	Continuation of oral capsules 3/day
90-Day Follow- up Post-FMT	Improved GI symptoms, thermoregulation, and food tolerance; physiotherapy mostly active	End of oral capsules course (3 months)

Blood and serum analyses include testing for Cytomegalovirus (CMV), Epstein–Barr virus (EBV), hepatitis viruses A (HAV), B (HBV), C (HCV), and E (HEV), HIV-1/2, and syphilis (*Treponema pallidum*), as well as general blood chemistry tests (complete blood count, C-reactive protein [CRP], erythrocyte sedimentation rate [ESR], albumin, creatinine, electrolytes, aminotransferases [ALT, AST], bilirubin, gammaglutamyl transferase [GGT], and alkaline phosphatase).

In parallel, stool samples are examined using cultural, molecular, and microscopic methods to exclude intestinal pathogens such as bacteria (Salmonella spp., Campylobacter spp., Shigella spp., Yersinia spp., Escherichia coli O157:H7), viruses, protozoa, and helminths, as well as Clostridioides difficile toxin A/B-producing strains, detected by chemiluminescence assays. Additional tests are also performed to assess fecal occult blood, calprotectin, and Helicobacter pylori fecal antigen.

Furthermore, screening includes the evaluation of possible colonization by multidrug-resistant organisms (MDROs), such as vancomycin-resistant *Enterococcus faecium* and *Enterococcus faecalis* (VRE), methicillin-resistant *Staphylococcus aureus* (MRSA), and Gram-negative bacteria producing carbapenemases or beta-lactamases, as well as *Listeria monocytogenes*. Starting from March 2020, due to the COVID-19 pandemic, the screening protocol was updated to include the detection of SARS-CoV-2 in fecal samples.

FMT solution

Two aliquots of 50 g of fecal material were prepared. Each aliquot was suspended in 150 mL of 0.9% NaCl sterile saline solution supplemented with 10% glycerol ((MONICO s.p.a.-Venezia). The subsequent filtration step was carried out using sterile plastic bags with an integrated gauze filter, STOMACHER[®] 400 Circulator Strainer Bags (Seward, UK), and homogenized with a STOMACHER[®] 400 Circulator (Fisher Scientific, Waltham, USA) at a speed of 230 rpm for 30 seconds (10).

The suspension resulting from the filtration of each aliquot was transferred into dedicated 200 mL sterile containers. One aliquot was stored at -80 °C, while the other was used for the preparation of frozen capsules. In addition, two further aliquots (1.5 ml) were stored for possible quality control testing.

Capsule preparation

Gastro-resistant capsules, size <ns/>1 (FARMALABOR SRL – Canosa di Puglia, BT), were filled with 400 μ L of fecal suspension. Than each size <ns/>1 capsule was inserted into a gastro-resistant capsule, size <ns/>00 (FARMALABOR SRL – Canosa di Puglia, BT). The prepared capsules were stored in a freezer at –80 °C (11).

Pre-FMT evaluation

The patient presented with a severe neurological condition, characterized by marked spastic hypertonia of the lower limbs and the left upper limb. Ambulation was absent, while upright posture was possible only for a few seconds with bilateral support. Neurological examination revealed bilateral Babinski sign and deep tendon reflexes hyperactive in the lower limbs and hypoactive in the upper limbs. Motor function in the right upper limb was partially preserved, whereas the left upper limb was severely impaired. Spasticity was evident at lower limbs and left arm, limiting passive mobilization of arms. The Expanded Disability Status Scale (EDSS) score was 8.

30-day follow-up post-FMT

The patient reported initial clinical improvement, including mild reduction in spasticity affecting limbs and increased stability while standing. Neurological examination confirmed the improved ability to maintain an upright posture with bilateral support (although ambulation remained impossible), the presence of brisk reflexes in the lower limbs, and no deterioration in sensory function or emergence of new neurological deficits.

60-day follow-up post FMT

The patient regained fine motor skills in the right hand, now able to hold a pen and sign her name. She is also capable of combing her hair and washing her face independently. All other neurological findings remain unchanged.

90-day follow-up post-FMT

The most recent evaluation documented further clinical improvement. The patient was able to maintain upright posture with greater stability compared to previous assessments and

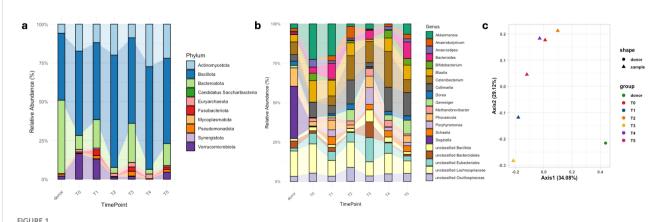
reported a more marked reduction in spasticity, particularly in the lower limbs. Both subjective and objective benefits were observed from physiotherapy, which is now conducted primarily in active mode. The patient reported improved food tolerance, gastrointestinal symptoms and thermoregulation. A summary is reported in Table 1.

To assess the modulation of the gut microbiota, six stool samples from the patient and the donor sample were analyzed using 16s metabarcoding and analyzed as described above (12). The samples were collected prior to the colonoscopic infusion (T0) and then monthly for five consecutive months. At baseline (T0), the microbiota profile showed a marked increase in Verrucomicrobiota (17%) and Actinomycetota (17%), along with a reduced abundance of Bacteroidota (9%) and an elevated proportion of Bacillota (54%). Concerning these, two peakes in the latter were observed at T2 (73%) and T4 (66%), followed by a return to baseline level. This variation coincided with a marked decrease in Bacteroidota, with relative abundances of 4% and 3% respectively (Figure 1a). At the genus level, a notably high abundance of Akkermansia (23%) was observed at T0; however, its levels remained consistently reduced and closer to normal values in all post-FMT time points. Among other genera, Collinsella showed a relative abundance of 9.8% at T0, which dropped to 2.5% at T3, and subsequently rose to around 15% in the following time points. Blautia exhibited fluctuating values between 8% and 14%, except at T3, where it showed a marked decrease to 1.4%. Additionally, Phocaeicola had a low relative abundance at T0 (2.8%), peaked at T3 (7.2%), dropped again at T4 (1.7%), and then stabilized at 4.3% at T5. Unclassified Bacteroidales and Unclassified Eubacteriales were more represented always in T3 (10% and 15.3%, starting from 1.5% and 6% respectively), while Unclassified Lachnospiraceae decreased (9% vs 18.5% at T0) (Figure 1b). Interestingly, Bray Curtis beta diversity highlighted that the patient's samples remained consistently distant from the donor's profile (Figure 1c). However, intestinal permeability, assessed by the lactulose-mannitol test, showed a mannitol value at before FMT (193.0 mg) higher than at four months (<0.2 mg)) by HPLC PAD method.

Discussion

Studies evaluating the efficacy and safety of FMT capsules primarily relate to their use in *Clostridioides difficile* infections. In a meta-analysis including a total of 664 patients, most adverse events were mild gastrointestinal symptoms such as nausea, diarrhea, and abdominal pain. Only 14 serious adverse events were reported, all of which were attributed to pre-existing comorbidities, except for two cases of hospitalization due to recurrent *Clostridioides difficile* infection. Therefore, no serious adverse events were considered potentially attributable to oral FMT capsules (13). With regard to patients affected by multiple sclerosis, we can only hypothesize that their use should first be evaluated by a neurologist to rule out any potential issues related to swallowing.

This report documents, to the best of our knowledge, the first combined FMT, via colonoscopy and oral capsules, in a patient with SPMS, primarily aimed to treat gastrointestinal symptoms. This approach, which combines direct enteral delivery with oral administration, allows for an initial deep colonization of the gut followed by prolonged maintenance of the donor microbiota. MS is recognized as a multifactorial disease in which immune dysregulation plays a central role. The gut microbiota is now well established as a key player in the immune modulation and therefore in the development of autoimmune diseases, including MS (14). Particularly, it has been clearly demonstrated that, in these patients, the abundance of butyrateproducing bacteria is reduced (15). Butyrate belongs to the group of bacterial metabolites known as short-chain fatty acids (SCFAs), which play a role in the differentiation of regulatory T cells (Tregs) in the gut district (16). Recently, a study documented that butyrate induces the expansion of CD69⁺CD56^{dim} NK cells and enhances the cytotoxicity of CD56^{bright} NK cells against hyperactivated CD25⁺CD69⁺ T cells, which



Microbiota profiling of the patient's samples before (T0) and following Faecal Material Transplantation every 30 days (T1, T2, T3, T4, T5) and donor's sample characterization. (a) shows relative abundances at phylum level. (b) shows relative abundances of the top 20 genera, while (c) represents Bray Curtis beta diversity represented as Principal Coordinate analysis.

are potentially harmful in autoimmune settings, while modulating the responce toward overall autologus CD4⁺ T cell compartment (17). In this context, other studies have highlighted how specific bacterial taxa are observed in MS. Akkermansia muciniphila is particularly overrepresented in MS patients compared to healthy controls (14). A proposed hypothesis is that, due to its specific mucin-degrading activity, an increased abundance of this strain may enhance its effect on the mucosal layer, thereby exerting a pro-inflammatory action (18). Similarly, Collinsella genus and other bacteria has been associated with worse disease course, also evident on radiological evaluation (19). Considering this background, FMT emerges as a promising immunomodulatory strategy even in MS patients, documented only in limited number of cases (6). In the present case, patient's neurological condition showed a positive trend particularly in spasticity and balance, following FMT and the maintenance of physiotherapy. Spasticity was reduced, upright posture became more stable and sustained, and some subtle objective neurological parameters improved. The main clinical response was observed 90 days after beginning of procedure. The efficacy of FMT is further supported by 16S rRNA gene sequencing analysis. Examining the evaluated time points, the most favorable microbiota redistribution was observed at T3, precisely 90 days. At the phylum level, this was characterized by a decrease in Actinomycetota and Verrucomicrobiota, accompainied by an increase in Bacteroidota. This pattern reflected a reduction in Akkermansia muciniphila and Collinsella genus and a rise in Phocaeicola genus and unclassified Bacteroidales. The association of Akkermansia muciniphila and Collinsella with MS have been previously described. Regarding Unclassified Bacteriodales, we can only state that they contribute to raising the overall level of Bacteroidota. Phocaeicola reflects the update taxonomy of Parabacteroides genus and certain strains may confer favorable effects by the production of SCFAs (20) Unclassified Eubacteriales showed an increase, potentially contributing to beneficial effects through the production of SCFAs, including butyrate (21). The use of encapsulated microbiota was well tolerated by the patient and may contribute to maintaining a protective effect mainly against the overgrowth of Akkermansia muciniphila or other yet-to-be-identified species. Importantly, the fluctuations observed in microbiota profiles are likely attributable to wellknown confounding factors (genetic, dietary, or environmental) which can hinder the identification of biomarkers and impair successful engraftment of the donor microbiota. Indeed, the comparison with the donor sample revealed a stable distance between microbial profiles, despite prolonged microbiota administration. However, the overall improvement suggests a beneficial effect of FMT, which can be considered an adjunctive approach when combined with intensive physiotherapy and standard care protocols.

Conclusion

The few cases reported to date, including the present one, consistently show a shift toward a more balanced microbial

profile and the potential role of FMT in modulating gut-immune-CNS axis. Moreover, the use of capsules containing intestinal microbiota represents a clear advantage in the FMT procedure, as they can be taken by the patient at home. However, the major limitation of this study is that it evaluates a single patient. This makes it impossible to identify potential microbial species-level clusters associated with MS, which can only be determined through comparison across multiple subjects by metagenomic. Therefore larger, controlled or pilot studies are required to improve the robustness of these preliminary finding.

Data availability statement

The data presented in this article have been deposited with NCBI BioProject, accession number PRJNA1365666.

Ethics statement

The studies involving humans were approved by local Ethics Committee "Territoriale Lazio area 3" (prot N 0000623/24- 22/03/ 2024). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

SB: Conceptualization, Data curation, Resources, Writing – original draft. FD: Resources, Writing – original draft, Formal Analysis, Investigation. FC: Resources, Writing – original draft. GQ: Resources, Writing – original draft. Data curation, Project administration. RR: Writing – original draft, Resources. MM: Resources, Writing – original draft. DD: Resources, Writing – original draft. MS: Formal Analysis, Writing – review & editing. GC: Conceptualization, Writing – review & editing. VD: Conceptualization, Writing – review & editing. LM: Resources, Conceptualization, Data curation, Formal Analysis, Funding acquisition, Project administration, Supervision, Validation, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, and/or publication of this article.

Acknowledgments

Ministry of Health RC2025.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Generative AI statement

The author(s) declare that Generative AI was used in the creation of this manuscript. Artificial intelligence tools were employed solely to verify the English language and style of the manuscript, without generating any original scientific content.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- 1. Baecher-Allan C, Kaskow BJ, Weiner HL. Multiple sclerosis: mechanisms and immunotherapy. Neuron.(2018) 97:742–68. doi: 10.1016/j.neuron.2018.01.021
- iMSMS Consortium. Electronic address: sergio.baranzini@ucsf.edu; iMSMS Consortium. Gut microbiome of multiple sclerosis patients and paired household healthy controls reveal associations with disease risk and course. Cell. (2022) 185:3467–3486.e16. doi: 10.1016/j.cell.2022.08.021
- 3. Yang R, Chen Z, Cai J. Fecal microbiota transplantation: Emerging applications in autoimmune diseases. *J Autoimmun*. (2023) 141:103038. doi: 10.1016/j.jaut.2023.103038
- 4. Cammarota G, Masucci L, Ianiro G, Bibbò S, Dinoi G, Costamagna G, et al. Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent Clostridium difficile infection. . *Aliment Pharmacol Ther.* (2015) 41:835–43. doi: 10.1111/apt.13144
- 5. Khanna L, Zeydan B, Kantarci OH, Camilleri M. Gastrointestinal motility disorders in patients with multiple sclerosis: A single-center study. Neurogastroenterol Motil. (2022) 34:e14326. doi: 10.1111/nmo.14326
- Laeeq T, Vongsavath T, Tun KM, Hong AS. The potential role of fecal microbiota transplant in the reversal or stabilization of multiple sclerosis symptoms: A literature review on efficacy and safety. Microorganisms (2023) 11:2840. doi: 10.3390/microorganisms11122840
- 7. Mahmud N, Molloy A, McPartlin J, Corbally R, Whitehead AS, Scott JM, et al. Increased prevalence of methylenetetrahydrofolate reductase C677T variant in patients with inflammatory bowel disease, and its clinical implications. *Gut.*(1999) 45:389–94. doi: 10.1136/gut.45.3.389
- 8. Tinto N, Cola A, Piscopo C, Capuano M, Galatola M, Greco L, et al. High frequency of haplotype HLA-DQ7 in celiac disease patients from south Italy: retrospective evaluation of 5, 535 subjects at risk of celiac disease. *PloS One.* (2015) 10:e0138324. doi: 10.1371/journal.pone.0138324
- 9. Cammarota G, Ianiro G, Tilg H, Rajilić S, Mirjana Kump P, et al. European consensus conference on faecal microbiota transplantation in clinical practice, ". *Gut.* (2017) 66:569–80. doi: 10.1136/gutjnl-2016-313017
- 10. Quaranta G, Fancello G, Ianiro G, Graffeo R, Gasbarrini A, Cammarota G, et al. Laboratory handling practice for faecal microbiota transplantation, ". *J Appl Microbiol.* (2020) 128:893–8. doi: 10.1111/JAM.14522
- 11. Youngster I, Mahabamunuge J, Systrom HK, Sauk J, Khalili H, Levin J, et al. Oral, frozen fecal microbiota transplant (FMT) capsules for recurrent Clostridium difficile infection. *BMC Med.* (2016) 14:134. doi: 10.1186/s12916-016-0680-9

- 12. Niccolai E, Martinelli I, Quaranta G, Nannini G, Zucchi E, De Maio F, et al. Fecal microbiota transplantation in amyotrophic lateral sclerosis: clinical protocol and evaluation of microbiota immunity axis. *Methods Mol Biol.* (2024) 2761:373–96. doi: 10.1007/978-1-0716-3662-6_27
- 13. Walsh SMLS, Grinspan AMD. Oral fecal microbiota transplant capsules are safe and effective for recurrent *clostridioides difficile* infection. A systematic review and meta-analysis. *J Clin Gastroenterol*. (2021) 55:300–8. doi: 10.1097/MCG.000000000001495
- 14. Cekanaviciute E, Yoo BB, Runia TF, Debelius JW, Singh S, CA N, et al. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. *Proc Natl Acad Sci U S A.*(2017) 114:10713–8. doi: 10.1073/pnas.1711235114
- Levi I, Gurevich M, Perlman G, Magalashvili D, Menascu S, Bar N, et al. Potential role of indolelactate and butyrate in multiple sclerosis revealed by integrated microbiome-metabolome analysis. *Cell Rep Med.* (2021) 2:100246. doi: 10.1016/j.xcrm.2021.100246
- 16. Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature*.(2013) 504:451–5. doi: 10.1038/nature12726
- 17. Carlini F, Squillario M, Casella V, Capaia M, Lusi V, Bagnara D, et al. Butyrate enhances CD56^{bright} NK cell-driven killing of activated T cells and modulates NK cell chromatin accessibility. *Genes Immun.* (2025) 26:342–51. doi: 10.1038/s41435-025-00338-2
- 18. Si J, Kang H, HJ Y, Ko G. Revisiting the role of Akkermansia muciniphila as a therapeutic bacterium. *Gut Microbes.* (2022) 14:1. doi: 10.1080/19490976.20
- 19. Jette S, de Schaetzen C, CC T, Tremlett H. The multiple sclerosis gut microbiome and disease activity: A systematic review. *Mult Scler Relat Disord.* (2024) 92:106151. doi: 10.1016/j.msard.2024.106151
- Clausen U, ST V, Lambertus P, Gehler M, Scheve S, Wöhlbrand L, et al. Catabolic network of the fermentative gut bacterium phocaeicola vulgatus (Phylum bacteroidota) from a physiologic-proteomic perspective. *Microb Physiol.* (2024) 34:88– 107. doi: 10.1159/000536327
- 21. Sorbara MT, Littmann ER, Fontana E, Moody TU, Kohout CE, Gjonbalaj M, et al. Functional and genomic variation between human-derived isolates of lachnospiraceae reveals inter- and intra-species diversity. *Cell Host Microbe.* (2020) 28:134–146.e4. doi: 10.1016/j.chom.2020.05.005