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Alpha-lipoic acid ameliorates nab-paclitaxel-induced peripheral neuropathy by inhibiting IL-17 signaling pathway

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Background: The precise mechanisms by which alpha-lipoic acid (LA) alleviates nab-paclitaxel (nab-PTX)-induced peripheral neuropathy have yet to be fully elucidated. The objective of this study was to investigate the mechanisms underlying the neuroprotective effects of LA in mitigating nab-PTX-induced peripheral neuropathy.

Methods: We established a rat model of nab-PTX-induced peripheral neuropathy to evaluate the efficacy of LA. To systematically elucidate the mechanisms by which LA alleviates nab-PTX-induced peripheral neuropathy, we utilized an integrated approach that combined network toxicology and network pharmacology. Subsequently, molecular docking analysis was performed to assess the binding affinity of the LA to the target proteins involved in the key signaling pathway. Furthermore, experimental validation was conducted to confirm the role of the key signaling pathway in the neuroprotective mechanism of LA.

Results: LA was demonstrated to effectively alleviate nab-PTX-induced peripheral neuropathy. The network analysis indicated that LA ameliorated nab-PTX-induced peripheral neuropathy primarily through the AGE-RAGE signaling pathway in diabetic complications, IL-17 signaling pathway, fluid shear stress and atherosclerosis, NOD-like receptor signaling pathway, and pathways of neurodegeneration - multiple diseases. The molecular docking indicated a potential impact of LA on the IL-17 signaling pathway. Further experiment validation revealed that nab-PTX activated the IL-17 signaling pathway, whereas LA could mitigate nab-PTX-induced peripheral neuropathy by inhibiting this pathway.

Conclusion: By integrating network toxicology analysis, network pharmacology analysis, and experimental validation, this study provides a clearer understanding of the mechanisms by which LA ameliorates nab-PTX-induced peripheral neuropathy.

KEYWORDS

alpha-lipoic acid, nab-paclitaxel, peripheral neuropathy, network toxicology, network pharmacology

1 Introduction

Nanoparticle albumin-bound paclitaxel (nab-PTX) is a widely used chemotherapeutic agent that has demonstrated efficacy in various cancers (1–6). However, it induces a high incidence and severity of peripheral neuropathy, which represents a major dose-limiting toxicity that severely compromises patients' quality of life and treatment outcomes (7–9). The increased neurotoxicity of nab-PTX, compared to solvent-based paclitaxel, is likely attributable to its improved delivery and higher accumulation in peripheral nerves (10, 11). Unfortunately, efficient therapeutic strategies to manage this complication are largely limited. Therefore, it is essential to monitor peripheral neuropathy during nab-PTX treatment, and there is an urgent need to identify effective strategies to mitigate its development.

Alpha-lipoic acid (LA), a natural antioxidant with established efficacy in mitigating diabetic peripheral neuropathy (DPN), has shown promise in alleviating chemotherapy-induced peripheral neuropathy (CIPN) (12–15). Its potential protective effects against nab-PTX-induced neuropathy, however, remain incompletely understood. While previous research on LA has illuminated LA's impact on isolated pathways like Nrf2, the comprehensive network of molecular targets and the primary signaling pathways through which LA counteracts nab-PTX neurotoxicity are still elusive (16).

Given the multifactorial pathogenesis of peripheral neuropathy, a systematic approach is crucial for elucidating complex drug actions. In this study, we employed an integrative strategy combining network toxicology and network pharmacology to holistically predict the potential targets and pathways involved in LA's protection against nab-PTX-induced peripheral neuropathy (17, 18). The integrated approach might help us identify multiple signaling pathways through which LA mitigates nab-PTX-induced peripheral neuropathy, such as Nrf2/ARE, NF- κ B, PI3K/Akt and others. Experimental validation was then performed to confirm the role of the key signaling pathway in the neuroprotective mechanism. Our study addresses the limitations of previous research, which usually focused on isolated aspects, thereby providing a more comprehensive and systematic investigation into the mechanisms by which LA alleviates nab-PTX-induced peripheral neuropathy.

Abbreviations: LA, alpha-lipoic acid; nab-PTX, nab-paclitaxel; DPN, diabetic peripheral neuropathy; CIPN, chemotherapy-induced peripheral neuropathy; ELISA, enzyme-linked immunosorbent assay; SD, Sprague-Dawley; LLA, low dose LA; MLA, medium dose LA; HLA, high dose LA; PWT, paw withdrawal threshold; CTD, Comparative Toxicogenomics Database; OMIM, Online Mendelian Inheritance in Man; PPI, Protein-Protein Interaction; GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; PDB, Protein Data Bank; DRG, dorsal root ganglia; BP, biological processes; CC, cellular components; MF, molecular functions; SABV, sex as a biological variable; NIH, National Institutes of Health.

2 Materials and methods

2.1 Drugs and reagents

The lipoic acid injection (Yabao Pharmaceutical Group Co. Ltd.), nab-PTX injection (Jiangsu Hengrui Pharmaceuticals Co., Ltd.), and normal saline injection (Sichuan Kelun Pharmaceutical Co., Ltd.) were obtained from Eye Ear Nose Throat Hospital of Fudan University. The enzyme-linked immunosorbent assay (ELISA) kits for IL-17, TNF- α , and MMP-13 were obtained from Shanghai Jianglai Biotechnology Co., Ltd., while the kit for HSP-90 α was sourced from UpingBio.

2.2 Animals and treatment

Twenty male Sprague-Dawley (SD) rats were procured from Shanghai Bikai Keyi Biotechnology Co., Ltd., and were housed under standardized laboratory conditions, which included a 12-hour light/dark cycle, ambient temperature of 25°C, and relative humidity maintained between 55% and 60%. The animals were free access to food and water throughout the study. The experimental procedures adhered strictly to the guidelines and recommendations set forth by the International Association for the Study of Pain. Every effort was made to minimize both the number of animals utilized and their potential suffering throughout the study. After a one-week adaptation period, the rats were randomly assigned into five groups (n=4 per group) using a computer-based random order generator: control group (administered normal saline), nab-PTX group (10 mg/kg), low dose LA (LLA)+nab-PTX group (nab-PTX 10 mg/kg combined with 15 mg/kg LA), medium dose LA (MLA)+nab-PTX group (nab-PTX 10 mg/kg combined with 30 mg/kg LA), and high dose (HLA)+nab-PTX group (nab-PTX 10 mg/kg combined with 60 mg/kg LA). The nab-PTX was administered once a week for three consecutive weeks via tail vein injection; LA was given once daily via intraperitoneal injection. The assessment of peripheral neuropathy was conducted in a blinded manner with respect to drug administration on days 0, 1, 8, 15, and 22. This experiment was approved by the Experimental Animal Ethics Committee of School of Pharmacy Fudan University (2025-03-LY-GY-44). Rats were excluded from the study if they died prematurely, as this would prevent the collection of complete behavioral and histological data.

2.3 Peripheral neuropathy assessment

We employed the fixed-threshold von Frey method to assess the mechanical pain of rats (19). Each rat was individually placed in a transparent chamber (20×10×20 cm) containing a specially designed iron wire mesh platform, which formed a uniform 10 mm grid. Seven calibrated von Frey filaments (1, 2, 4, 6, 8, 10, and 15 g) were applied in ascending order to the central area of the plantar surface of one hind paw. Each filament was pressed against the midplantar skin five times until it slightly bent, and then held in place for 5 seconds. Trials consisted of five applications of each

filament to the hind paw, with a 15-second interval between each application. If the hind paw withdrew in response to a particular filament in at least 4 out of 5 applications, the force value of that filament (in grams) was defined as the paw withdrawal threshold (PWT). The withdrawal responses from both hind paws were documented, and the measurements from each paw were analyzed separately. The percentage response for the 4 g, 8 g, and 15 g von Frey filaments was calculated. Acetone test was performed to assess the cold allodynia of rats (20). A 0.05 ml drop of acetone was delicately applied to the central region of the ventral surface of the rat's hindpaw using a 1-ml syringe. Utmost care was exercised to prevent any contact between the syringe tip and the hindpaw, thereby avoiding any mechanical stimulation that could confound the results. The rats' responses to acetone application were assessed using a scoring system. A score of 0 indicated no cold allodynia, while 1, 2, and 3 corresponded to mild (scratching the hind paw), moderate (intense licking, biting, or withdrawal of the hind paw), and severe (vigorous licking, multiple withdrawals, or vocalizations) cold allodynia, respectively. Five trials were carried out at intervals of approximately 5 min for each hind paw. The observation window for each trial was 30 seconds, during which the rats were observed continuously, and the highest score observed within this period was recorded for each trial. The ambient temperature in the testing room was maintained at $25 \pm 1^\circ\text{C}$ to ensure a stable and controlled environment for the experiments. The statistical analysis was performed using one-way ANOVA followed by Dunnett's *post-hoc* test.

2.4 Network toxicology analysis

The potential mechanisms underlying nab-PTX-induced peripheral neuropathy were explored using network toxicology. The SwissTargetPrediction (<http://www.swisstargetprediction.ch/>) (21), DrugBank (<https://go.drugbank.com/>) (22), and Comparative Toxicogenomics Database (CTD) (<https://ctdbase.org/>) databases (23) were utilized to identify the potential targets of nab-PTX. Due to the relatively limited number of targets associated with nab-PTX, we retained all the potential targets. The pathological targets associated with peripheral neuropathy were obtained from the GeneCards (<https://www.genecards.org/>; relevance score > median value) and Online Mendelian Inheritance in Man (OMIM) (<https://omim.org/>) databases (24). Venn diagram was employed to identify the targets potentially involved in nab-PTX-induced peripheral neuropathy. The Protein-Protein Interaction (PPI) network was constructed using the STRING database (<https://cn.string-db.org/>), with interactions filtered based on a medium confidence score (25). Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses were performed using the DAVID (<https://davidbioinformatics.nih.gov/tools.jsp>) (26). The visualization of enrichment analysis was plotted using the Chiplot platform (<https://www.chiplot.online/>) (27).

2.5 Network pharmacology analysis

Network pharmacology was employed to investigate the potential protective effects of LA against nab-PTX-induced peripheral neuropathy. The SwissTargetPrediction, DrugBank, and CTD were used to identify the potential targets of LA. We had also retained all the potential targets of LA. The overlapping targets between LA and peripheral neuropathy were determined using Venn diagrams. Additionally, PPI network analysis with interactions filtered based on a medium confidence score, GO enrichment analysis, and KEGG pathway analysis were conducted to further elucidate the mechanisms.

2.6 Integration analyses of network toxicology and network pharmacology

The potential mechanisms by which LA ameliorates nab-PTX-induced peripheral neuropathy were initially identified through network toxicology and network pharmacology approaches. To achieve a more precise understanding of these mechanisms, further integrated analyses combining network toxicology and network pharmacology were performed. The overlapping targets among nab-PTX, LA, and peripheral neuropathy were identified using Venn diagrams. Additionally, PPI network analysis and KEGG pathway analysis were conducted to provide deeper insights into the underlying mechanisms.

2.7 Molecular docking analysis

The three-dimensional structure of LA was obtained from the PubChem Compound database (<https://www.ncbi.nlm.nih.gov/pccompound/>). Meanwhile, the crystal structure of the protein was sourced from the Protein Data Bank (PDB) database (<https://www.rcsb.org/>). Following this, the files of receptor and ligand were processed via the CB-Dock2 portal to carry out molecular docking analysis (28).

2.8 Validation of the IL-17 signaling pathway

All rats were put under deep anesthesia with a ketamine after the trial was finished. The serum, spinal cords and dorsal root ganglia (DRG) were collected and kept in liquid nitrogen in a freezer at a temperature of -80°C . The levels of IL-17, HSP-90 α , TNF- α , and MMP-13 in serum, spinal cords, and DRG were measured using enzyme-linked immunosorbent assay (ELISA) kits, following the manufacturer's instructions. The statistical analysis was performed using one-way ANOVA followed by Dunnett's *post-hoc* test.

3 Results

3.1 LA ameliorated nab-PTX-induced peripheral neuropathy

To investigate mechanical and cold allodynia, we performed von Frey and acetone tests on days 0, 1, 8,15, 22 after administration (Figure 1A). The results demonstrated that rats in the treatment of nab-PTX exhibited a significant decrease in bilateral paw withdrawal thresholds compared to those in the control group (Figure 1B). In terms

of 4, 8, and 15 g von Frey tests, rats treated with nab-PTX exhibited significantly more responses than those in the control group (Figure 1C). Similarly, the acetone test for cold allodynia revealed that nab-PTX-treated rats had significantly higher scores compared to controls (Figure 1D). LA demonstrated significant effects on nab-PTX-induced neuropathic pain. Rats treated with LA (in three dosage regimens) in combination with nab-PTX showed a significant increase in mechanical withdrawal thresholds compared to those treated with nab-PTX alone (Figure 1B). Regarding 4, 8, and 15 g von Frey tests, rats treated with LA plus nab-PTX exhibited significantly fewer responses than those treated

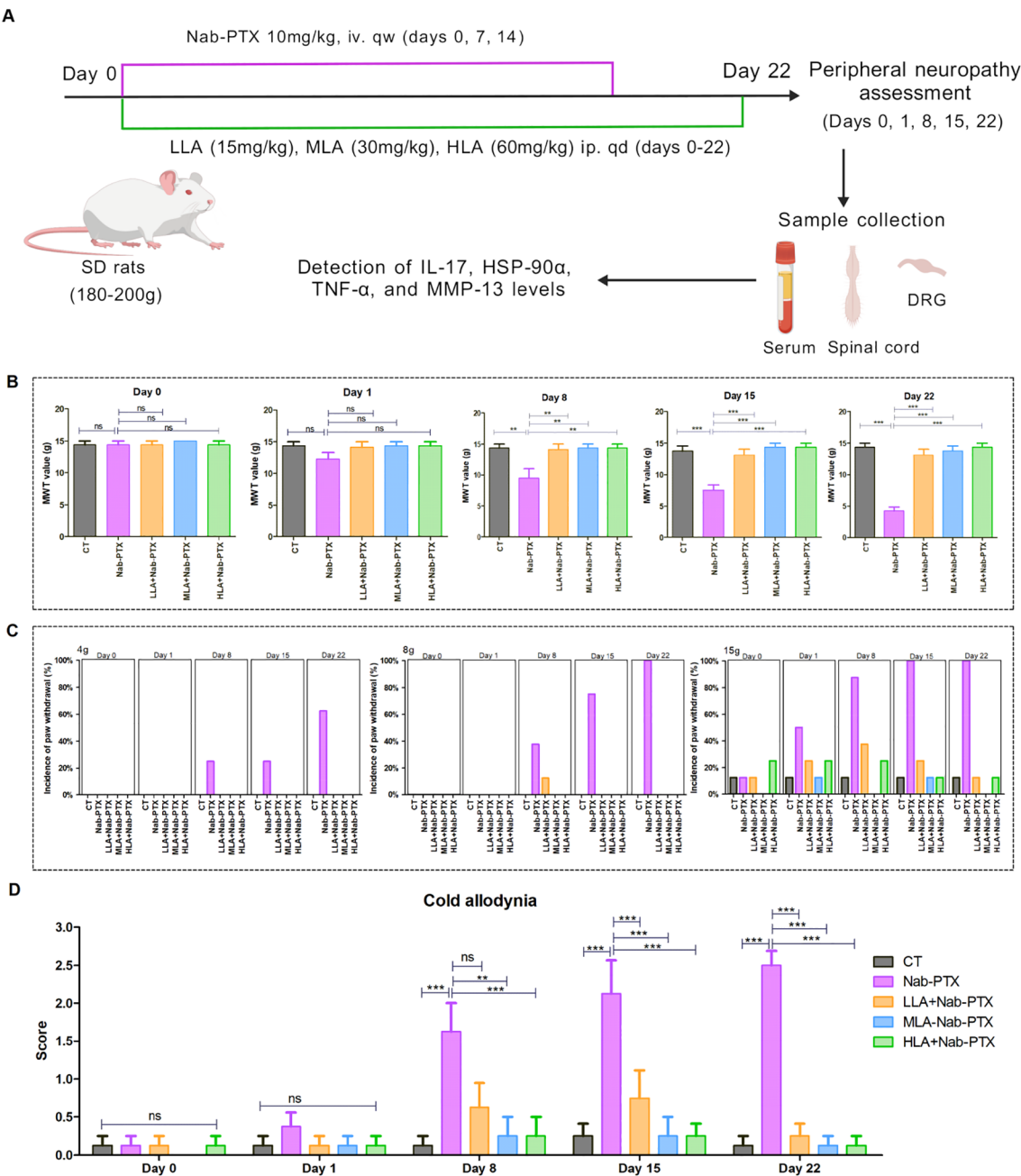


FIGURE 1
Neuroprotective effects of LA in rats of nab-PTX-induced peripheral neuropathy. (A) Schematic representation of animal experiment [created with biogdp.com (29)]; (B, C) Results of mechanical allodynia tests (n=8 paws); (D) Results of cold allodynia tests (n=8 paws). **P<0.01, ***P<0.001.

with nab-PTX alone (Figure 1C). Additionally, the cold allodynia scores of rats treated with LA plus nab-PTX were significantly lower compared to those treated with nab-PTX alone (Figure 1D). These results collectively suggested that LA could effectively alleviate mechanical and cold allodynia caused by nab-PTX. Rats treated with nab-PTX exhibited a slight decrease in body weight (Supplementary Figure S1).

3.2 Network toxicology analysis of the potential mechanisms of nab-PTX-induced peripheral neuropathy

A total of 154 target genes were identified in association with nab-PTX, while 4807 target genes were linked to peripheral neuropathy. Upon further analysis, an intersection of these two sets of genes identified 73 common target genes (Figure 2A). The PPI network of these targets was illustrated in Figure 2D. The network was constructed

based on the intricate interactions among the core targets. This optimized network diagram provided a clear visual representation of the relationships and functional associations among these core targets. GO enrichment analysis showed that, for biological processes (BP), the genes were predominantly involved in protein phosphorylation, epidermal growth factor receptor signaling pathway, ephrin receptor signaling pathway, and collagen-activated tyrosine kinase receptor signaling pathway. It suggested that kinase-mediated signal transduction disorder could be the core mechanism behind nerve damage. Regarding cellular components (CC), the genes were mainly localized to the ficolin-1-rich granule lumen, cytosol, secretory granule lumen, and plasma membrane, hinting abnormal secretion of neurotransmitters/inflammatory factors and dysfunction of membrane receptors may be involved in the toxic process. In terms of molecular functions (MF), the genes were primarily enriched in protein kinase activity, ATP binding, histone H2AXY142 kinase activity, and histone H3Y41 kinase activity (Figure 2C), which further confirmed the driving

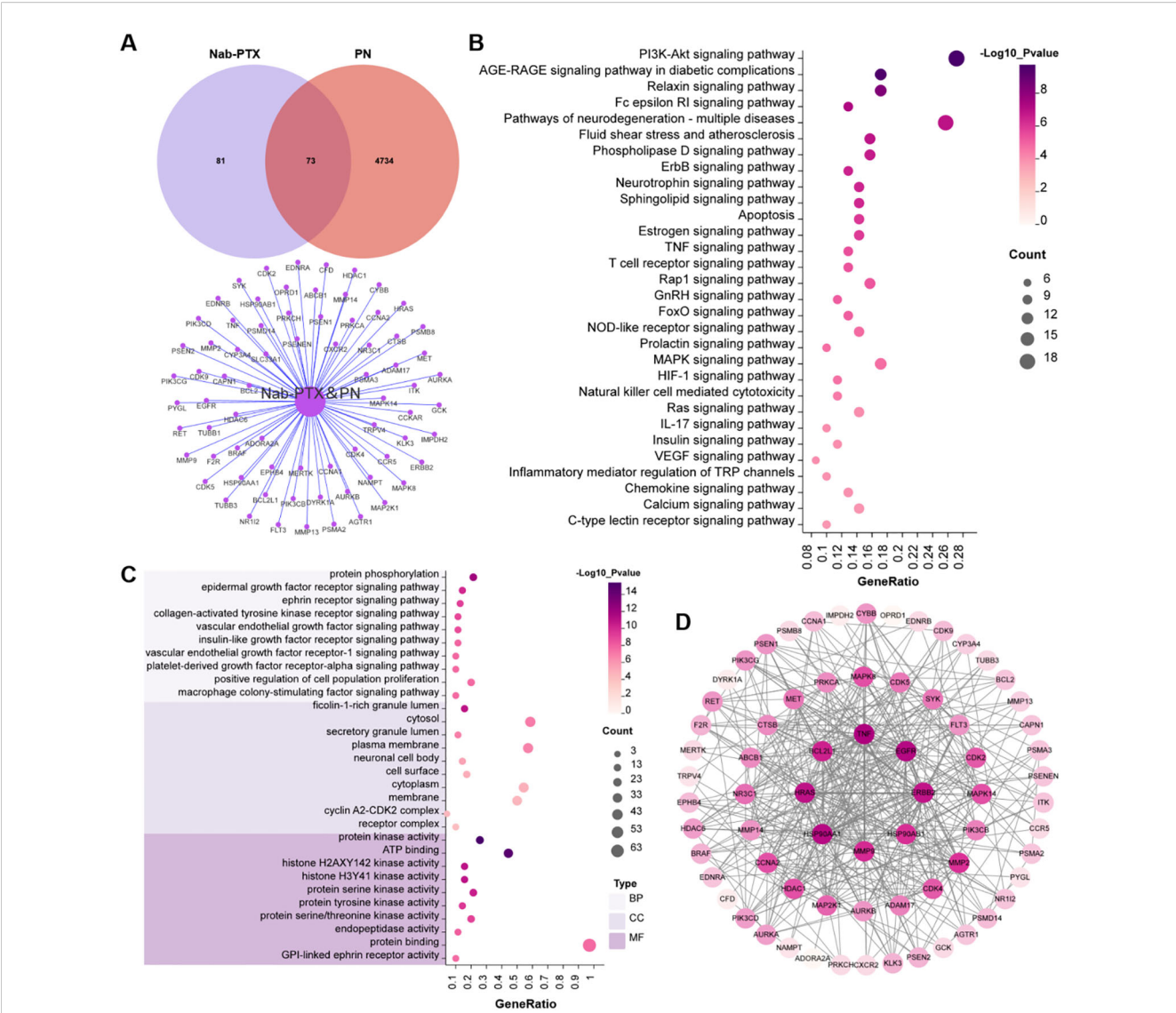


FIGURE 2 Results of network toxicology analysis. (A) Venn diagram; (B) KEGG analysis; (C) GO enrichment analysis; (D) PPI network.

effect of kinase activity imbalance in neuropathy. KEGG enrichment analysis highlighted the involvement of these genes in key pathways such as PI3K-Akt signaling pathway, AGE-RAGE signaling pathway in diabetic complications, fluid shear stress and atherosclerosis pathway, TNF signaling pathway, and IL-17 signaling pathway (Figure 2B). The result of KEGG enrichment analysis denoted that inflammatory response, oxidative stress, and imbalanced neurotrophic support are the core pathological basis of nab-PTX induced peripheral neuropathy.

3.3 Network pharmacology analysis of the potential neuroprotective mechanism of LA

As shown in Figure 3A, a total of 335 genes were identified in association with LA. Among these, 234 genes were found to overlap with those implicated in peripheral neuropathy. The PPI network exhibited a highly complex structure (Figure 3D). The GO enrichment

analysis revealed that LA primarily influenced biological processes associated with response to xenobiotic stimulus, response to hypoxia, positive regulation of gene expression, and response to oxidative stress (Figure 3C). It implied that LA had multidimensional protective effects such as antioxidant stress and anti-inflammatory effects. Additionally, KEGG enrichment analysis highlighted that the neuroprotective effects of LA were predominantly mediated through key pathways, including the AGE-RAGE signaling pathway in diabetic complications, apoptosis, fluid shear stress and atherosclerosis pathway, TNF signaling pathway, and IL-17 signaling pathway (Figure 3B).

3.4 Potential mechanism of LA ameliorating nab-PTX-induced peripheral neuropathy

Based on the results of network toxicology and network pharmacology, we found that the mechanism by which LA

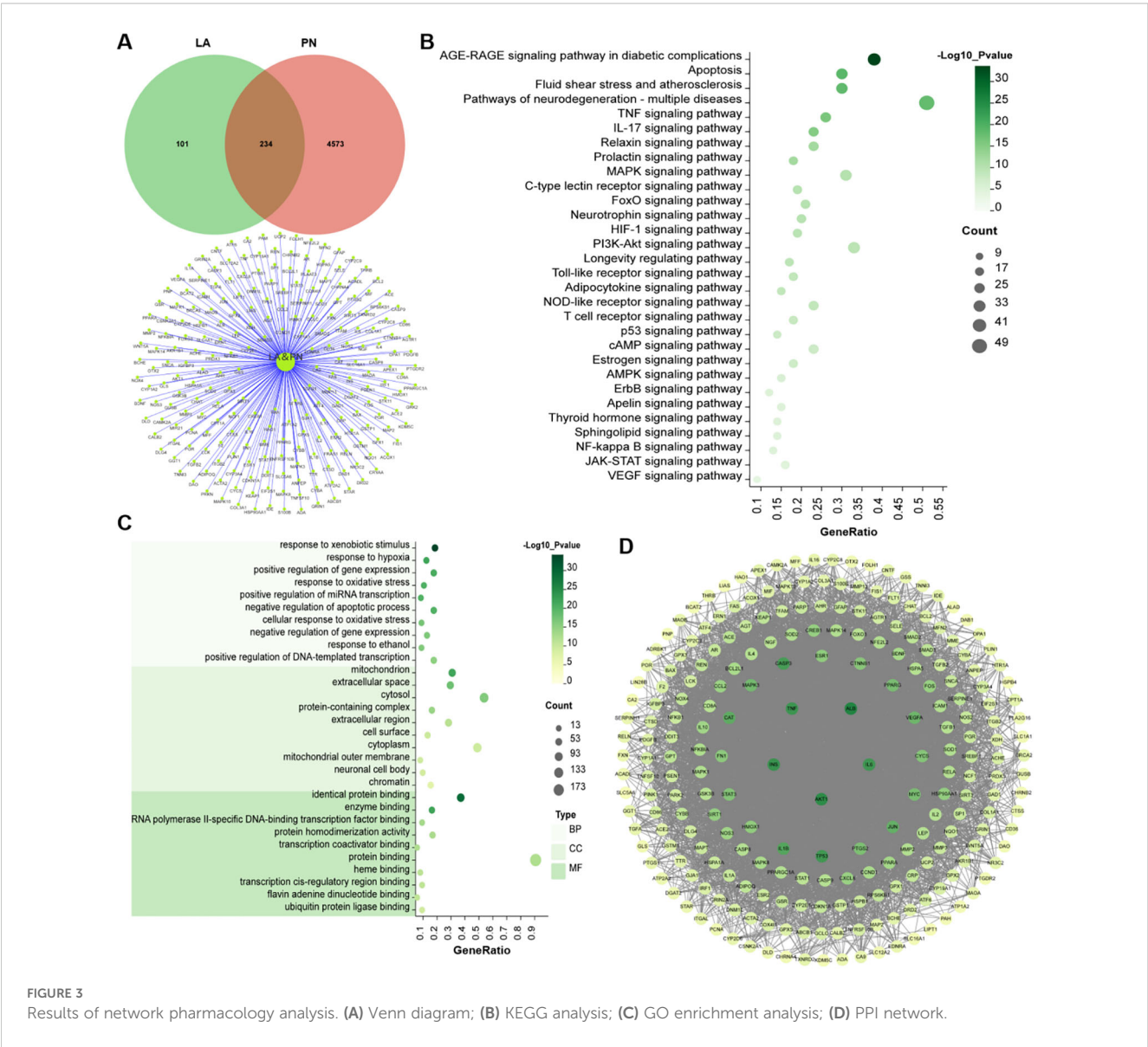


FIGURE 3 Results of network pharmacology analysis. (A) Venn diagram; (B) KEGG analysis; (C) GO enrichment analysis; (D) PPI network.

ameliorates peripheral neuropathy induced by nab-PTX might be associated with multiple signaling pathways such as PI3K-Akt signaling pathway, AGE-RAGE signaling pathway in diabetic complications, TNF signaling pathway and the IL-17 signaling pathway. Further integrated analyses, combining approaches from network toxicology and network pharmacology, identified 14 common genes shared among nab-PTX, LA, and peripheral neuropathy (Figures 4A, B). KEGG analysis revealed that the key pathways involved in these common genes included the AGE-RAGE signaling pathway in diabetic complications, IL-17 signaling pathway (Figure 4D), fluid shear stress and atherosclerosis, NOD-like receptor signaling pathway, and pathways of neurodegeneration - multiple diseases (Figure 4C).

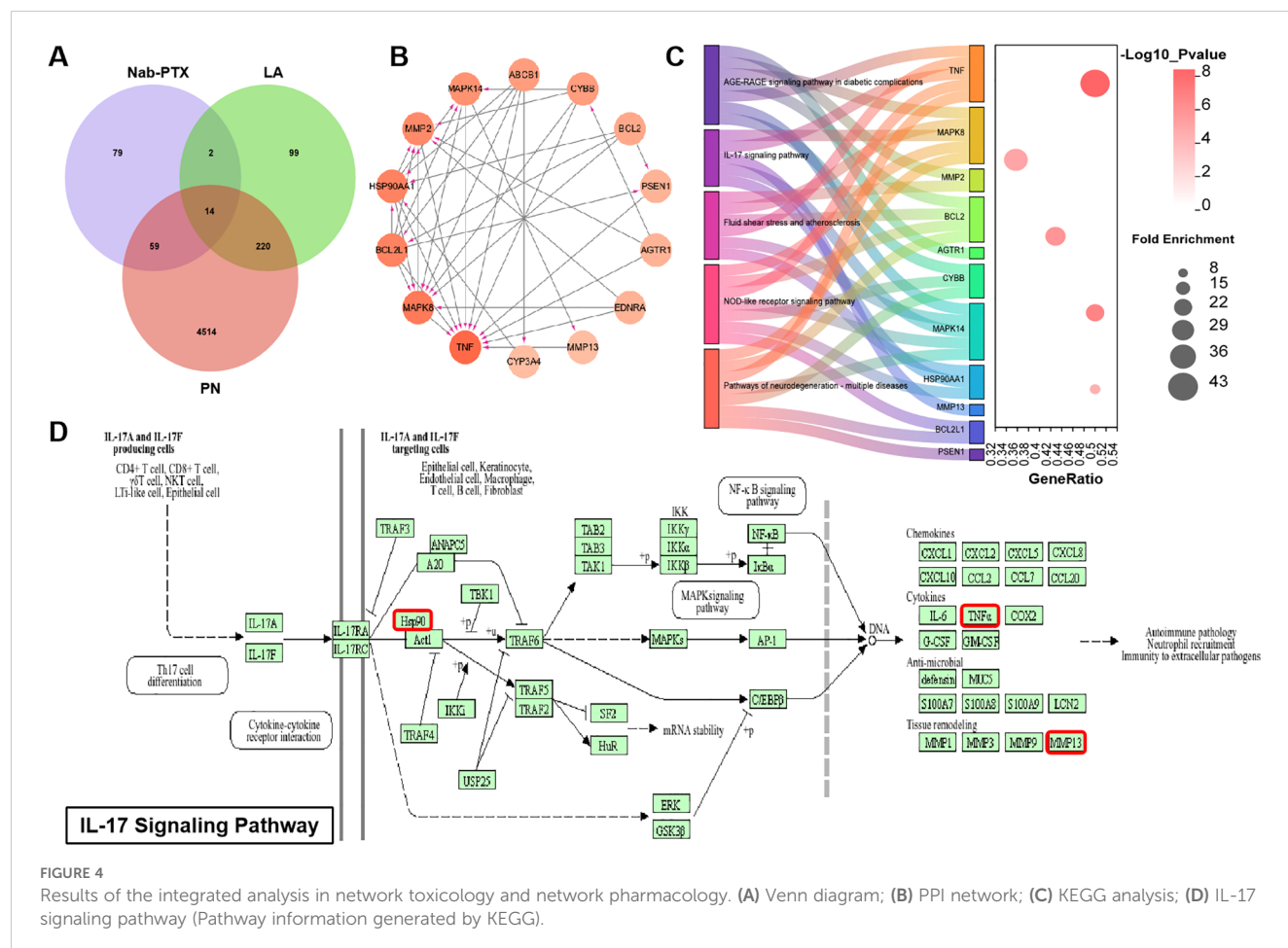
3.5 LA may ameliorate nab-PTX-induced peripheral neuropathy by IL-17 signaling pathway

Based on the predictions of network toxicology and network pharmacology, we identified several potential pathways but prioritized the IL-17 signaling pathway for further validation. This is due to its well-documented role in neuroinflammation

and allodynia, and its high concordance with the anti-inflammatory mechanisms of LA. The molecular docking analysis was performed to assess the binding affinity of the LA to the target proteins involved in IL-17 signaling pathway (Figures 5A-D). The PubChem CID of LA and the PDB IDs of all proteins employed for docking were provided in Supplementary Table S1. Given that all the results scored lower than -5.0 according to the Vina scoring system, this indicated that the ligand exhibited favorable binding interactions with the receptor. The findings suggest that LA may have a potential impact on the IL-17 signaling pathway.

3.6 LA ameliorated nab-PTX-induced peripheral neuropathy by inhibiting IL-17 signaling pathway

IL-17 signaling pathway was selected for experimental validation. The levels of IL-17, HSP-90 α , TNF- α , and MMP-13 in serum, spinal cord, and DRG of rats were presented in Figure 6. IL-17 expression analysis revealed distinct compartment-specific regulation. While serum IL-17 levels remained unchanged across all groups (Figure 6A), neural tissues showed significant pathway activation. In the spinal cord, nab-PTX treatment induced a



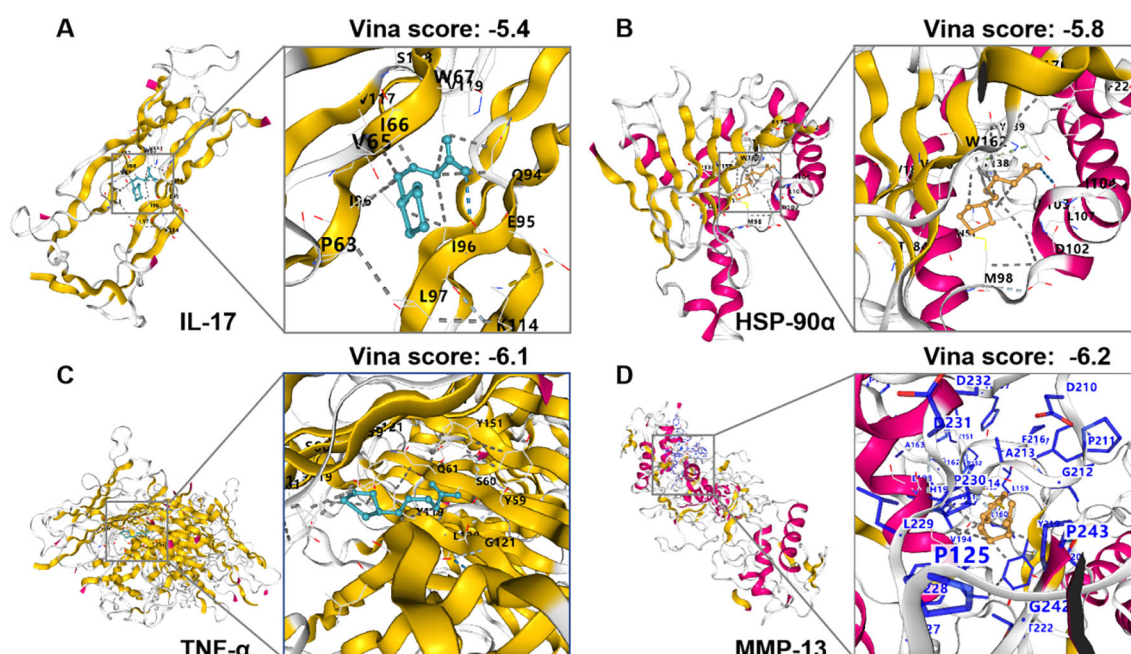


FIGURE 5
Docking results of LA with the (A) IL-17; (B) HSP-90α; (C) TNF-α; (D) MMP-13.

substantial increase in IL-17 compared to controls ($P < 0.05$, Figure 6B), which was effectively normalized by LA co-treatment. A parallel pattern was observed in DRG tissues (Figure 6C), where nab-PTX elevated IL-17 levels and LA administration significantly suppressed this increase.

HSP-90α measurements demonstrated a similar tissue-specific profile. No significant inter-group differences were detected in serum HSP-90α (Figure 6D). However, both spinal cord (Figure 6E) and DRG (Figure 6F) tissues from nab-PTX-treated rats exhibited marked HSP-90α upregulation ($P < 0.05$), which was significantly attenuated by LA treatment.

TNF-α expression patterns followed the established trend. Serum TNF-α levels showed no treatment-induced alterations (Figure 6G). In contrast, spinal cord (Figure 6H) and DRG (Figure 6I) tissues from the nab-PTX group displayed significantly elevated TNF-α ($P < 0.05$), with LA treatment effectively reducing these levels toward baseline.

MMP-13 quantification revealed distinct regulatory patterns across compartments. Serum analysis showed no significant difference in MMP-13 levels between the control and nab-PTX groups (Figure 6J). In contrast, MLA and HLA treatment significantly reduced serum MMP-13 compared to nab-PTX group ($P < 0.05$). Meanwhile, in neural tissues, nab-PTX-induced MMP-13 upregulation was observed in both spinal cord (Figure 6K) and DRG (Figure 6L), with LA treatment significantly suppressing these increases ($P < 0.05$).

In summary, by targeting the IL-17 inflammatory axis in spinal cord and DRG, LA coordinately suppresses the expression of TNF-α, HSP-90α, and MMP-13, thereby blocking the neuroinflammation-injury cascade. Moreover, these findings indicate that even low-dose LA is sufficient to effectively curb the

overactivation of the IL-17 pathway. These results not only validated prior network pharmacology predictions but provided mechanistically defined experimental evidence for LA's therapeutic potential against CIPN.

4 Discussion

Peripheral neuropathy is a major dose-limiting side effect of nab-PTX, significantly impacting patient quality of life (30). The management of peripheral neuropathy induced by nab-PTX continues to be an important challenge for both clinicians and cancer patients. In clinical practice, if a patient develops peripheral neuropathy while being treated with nab-PTX, the clinicians usually reduce the dose or discontinue the use of nab-PTX for the patient. However, dose reduction or discontinuation of nab-PTX may compromise the efficacy of cancer treatment, highlighting the urgent need for effective strategies to manage and mitigate peripheral neuropathy without sacrificing therapeutic outcomes.

The pathogenesis of CIPN and DPN exhibits considerable overlap. Both conditions are underpinned by comparable pathophysiological processes, such as oxidative stress, neuroinflammation, mitochondrial dysfunction, and changes in neuronal ion channel activities (31). Therefore, theoretically, drugs that could alleviate DPN should also be effective for CIPN. LA is a versatile compound boasting a wide range of potential health benefits, including potent antioxidant, anti-inflammatory, and neuroprotective properties (32, 33). In our study, to elucidate the mechanisms underlying the neuroprotective effects of LA in mitigating nab-PTX-induced peripheral neuropathy, we initially conducted network toxicology and network pharmacology analyses.

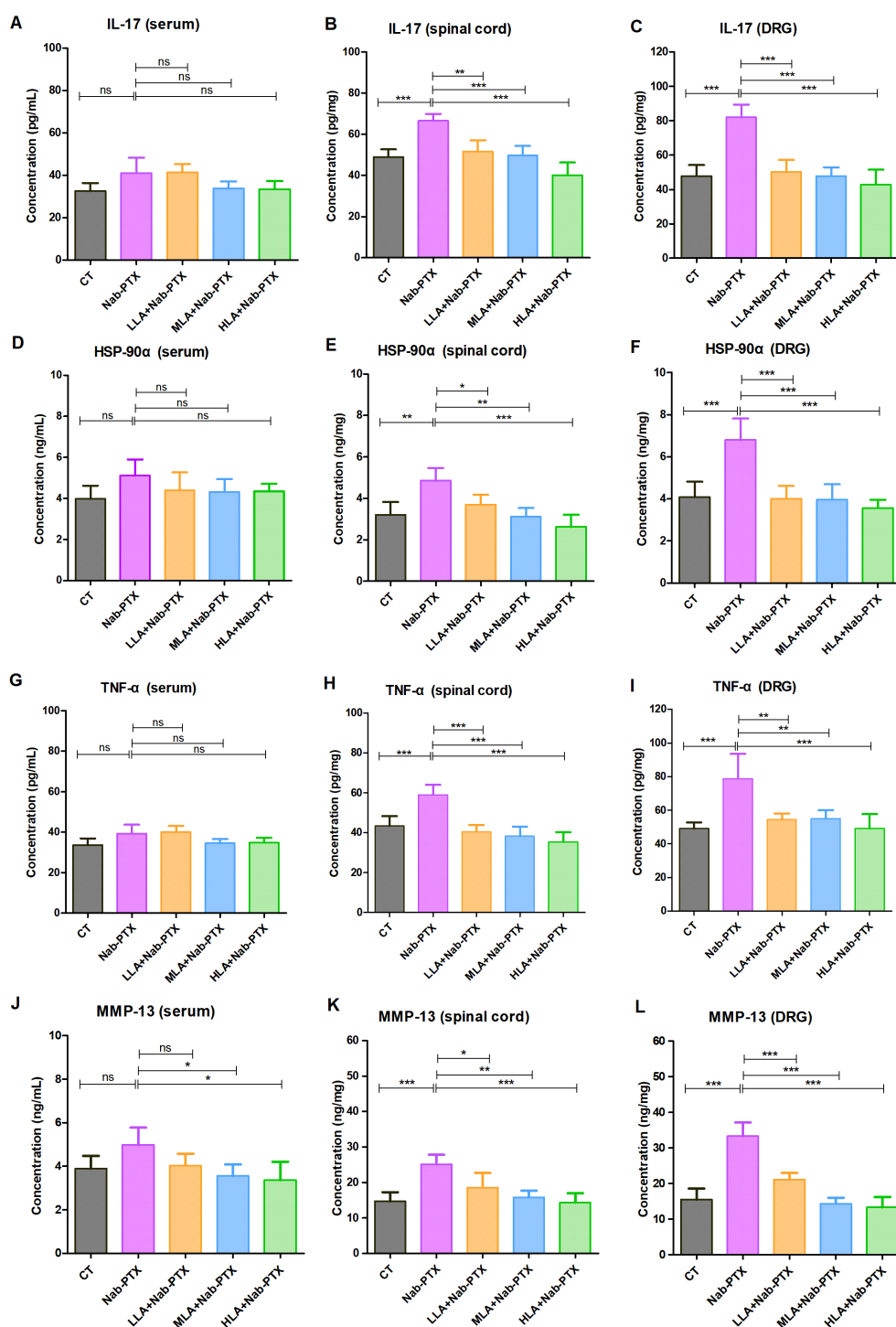


FIGURE 6

The detection results of IL-17, HSP-90α, TNF-α, and MMP-13. (A) IL-17 levels in serum; (B) IL-17 levels in spinal cord; (C) IL-17 levels in DRG; (D) HSP-90α levels in serum; (E) HSP-90α levels in spinal cord; (F) HSP-90α levels in DRG; (G) TNF-α levels in serum; (H) TNF-α levels in spinal cord; (I) TNF-α levels in DRG; (J) MMP-13 levels in serum; (K) MMP-13 levels in spinal cord; (L) MMP-13 levels in DRG. $n=4$, * $P<0.05$, ** $P<0.01$, *** $P<0.001$.

The former aimed to uncover the potential mechanisms of nab-PTX-induced peripheral neuropathy, while the latter focused on exploring how LA exerts its protective effects against peripheral neuropathy. Subsequently, we performed integrated analyses that combined insights from both network toxicology and network

pharmacology to pinpoint the precise neuroprotective mechanisms of LA against nab-PTX-induced peripheral neuropathy. Finally, we selected the IL-17 signaling pathway for validation through molecular docking analysis and experimental approaches. Through a comprehensive approach combining

network toxicology analysis, network pharmacology analysis, molecular docking analysis, and experimental validation, we have systematically elucidated the molecular mechanisms underlying LA's therapeutic effects against nab-PTX-induced peripheral neuropathy. Our network analysis has unveiled that the mechanism underlying the amelioration of nab-PTX-induced peripheral neuropathy by LA might be intricately linked to multiple signaling pathways. These pathways mainly include the AGE-RAGE signaling pathway in diabetic complications, apoptosis, fluid shear stress and atherosclerosis pathway, TNF signaling pathway, and IL-17 signaling pathway. Molecular docking analysis indicates a potential impact of LA on the IL-17 signaling pathway. Further experimental validation has elucidated that nab-PTX could activate the IL-17 signaling pathway. In contrast, LA could attenuate nab-PTX-induced peripheral neuropathy by inhibiting this pathway. The novelty of our study lies in the systematic identification and subsequent validation of the IL-17 signaling pathway as an important mechanism for LA's effect specifically in the context of nab-PTX-induced peripheral neuropathy, which to our knowledge, has not been reported before. Previous studies mainly focused on LA's antioxidant properties, our integrated approach of network toxicology/pharmacology and experimental validation provides a new mechanistic insight into its immunomodulatory role.

IL-17 is a pro-inflammatory cytokine that plays a crucial role in modulating immune responses, neuroinflammation and is implicated in pain regulation (34, 35). IL-17 was reported to play roles in neuropathic pain and neurological disorders (36–38). Research has shown that IL-17 may exacerbate neuropathic pain by stimulating the proliferation of astrocytes and increasing the secretion of pro-inflammatory cytokines in models of spinal nerve ligation-induced neuropathic pain (39). TNF- α is a cytokine primarily produced by immune cells, and it plays a significant role in immune responses, inflammatory processes, and apoptosis. TNF- α also plays a pivotal role in the pathogenesis of peripheral neuropathic pain (40). HSP-90 α is a multifunctional protein that plays critical roles in maintaining cellular homeostasis, responding to stress, and regulating key biological processes (41). MMP-13, a member of the peptidase M10 family of matrix metalloproteinases (MMPs), has been reported to promote paclitaxel-induced peripheral neuropathy (42, 43). These targets were involved in IL-17 signaling pathway.

Our findings demonstrated that in nab-PTX-induced peripheral neuropathy, IL-17 pathway activation occurs predominantly within neural microenvironments (spinal cord/DRG). By targeting the IL-17 inflammatory axis in spinal cord and DRG, LA coordinately suppresses the expression of TNF- α , HSP-90 α , and MMP-13, thereby blocking the neuroinflammation-injury cascade. The finding that LA modulates this pathway adds a new dimension to the understanding of nab-PTX-induced peripheral neuropathy as not just a neuronal but also an immune-mediated pathology. In addition to the IL-17 signaling pathway, other pathways may also be implicated in both the pathogenesis of nab-PTX-induced peripheral neuropathy and the therapeutic effects of LA. For instance, the AGE-RAGE pathway is implicated in oxidative stress and

inflammation (44, 45). Similarly, the TNF signaling pathway plays a crucial role in regulating immune responses and apoptosis (46, 47). These signaling pathways are not isolated; they form a complex network that collectively contributes to the alleviation of nab-PTX-induced peripheral neuropathy by LA. For instance, TNF- α is involved in these predicted pathways, highlighting the interconnected nature of these processes. Additionally, HSP90AA1 is involved in the IL-17 signaling pathway, fluid shear stress and atherosclerosis, and NOD-like receptor signaling pathway, further emphasizing the interplay between these pathways. The interplay among these pathways suggests that the development of nab-PTX-induced peripheral neuropathy is a multifactorial process involving oxidative stress, inflammation, and immune responses. The pleiotropic modulation of these key pathways by LA underscores its potential as a multi-target therapeutic strategy.

The findings of this study have significant clinical implications for the management of nab-PTX-induced peripheral neuropathy. Given that the doses of nab-PTX and LA utilized in this study were primarily extrapolated from clinically relevant dosing regimens, it is reasonable to directly apply the standard clinical dosages of LA for the prophylaxis of nab-PTX-induced peripheral neuropathy in clinical practice. In addition, LA is widely used in clinical practice and has a good safety profile, so there is no need to be overly concerned about safety issues. However, there are currently no clinical trials on the prevention of nab-PTX-induced peripheral neuropathy by LA. Future research should focus on validating these findings in clinical settings. Well-designed clinical trials are needed to assess the efficacy and safety of LA in treating nab-PTX-induced peripheral neuropathy.

While our study offers compelling evidence for the therapeutic potential of LA in mitigating nab-PTX-induced peripheral neuropathy, several limitations should be acknowledged. On the one hand, network analysis relies on the existing databases and computational models, which may not fully capture the complexity of biological systems. On the other hand, experimental validation was performed in animal models, which may not fully translate to humans. In addition, we only included male rats in this study. We recognize the importance of integrating sex as a biological variable (SABV) in preclinical research, as mandated by the National Institutes of Health (NIH). The generalizability of our findings regarding the protective effects of LA against nab-PTX-induced neuropathy to females may be limited. This aspect warrants careful consideration when interpreting our results. Future studies systematically investigating the efficacy and mechanisms of LA in both male and female animal models are essential to comprehensively understand its therapeutic potential. Moreover, while our *in vivo* findings robustly demonstrate the association between LA treatment and the modulation of the IL-17 pathway in neural tissues, the present study does not include *in vitro* experiments to elucidate the direct cellular mechanisms. Future studies utilizing cell culture models are essential to confirm the direct cellular targets of LA, delineate the precise molecular interactions, and establish causal relationships within the pathway, for instance, through techniques such as Western

blotting, qRT-PCR, and immunofluorescence. Furthermore, we have only experimentally validated the IL-17 signaling pathway; other predicted signaling pathways still need to be verified. Future studies should address these limitations by integrating more comprehensive datasets, conducting clinical studies, investigating the effect of gender on the outcome, conducting *in vitro* experiments, and verifying other predicted signaling pathways.

Despite the above limitations, our integrated approach—combining network toxicology, network pharmacology, molecular docking with *in vivo* validation provides strong, physiologically relevant evidence for the protective role of LA. The consistent downregulation of key pathway components (IL-17, TNF- α , HSP-90 α , MMP-13) at the protein level in the target tissues offers compelling support for our findings.

5 Conclusion

In conclusion, this study provides a comprehensive understanding of the molecular mechanisms by which LA ameliorates nab-PTX-induced peripheral neuropathy. The findings suggest that LA ameliorates nab-PTX-induced peripheral neuropathy in part by inhibiting the IL-17 signaling pathway in the rat model. Future research should focus on elucidating the direct cellular mechanisms of LA and subsequently translating these mechanistic insights into clinical validation through well-designed trials.

Data availability statement

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding authors.

Ethics statement

The animal study was approved by Experimental Animal Ethics Committee of School of Pharmacy Fudan University. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

HS: Conceptualization, Data curation, Formal Analysis, Investigation, Methodology, Resources, Software, Visualization,

Writing – original draft, Writing – review & editing. YC: Data curation, Formal Analysis, Investigation, Methodology, Resources, Writing – original draft, Writing – review & editing. LL: Resources, Writing – review & editing. XX: Resources, Writing – review & editing. JY: Conceptualization, Supervision, Writing – review & editing. TH: Conceptualization, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2025.1674709/full#supplementary-material>

References

1. Stinchcombe TE. Nanoparticle albumin-bound paclitaxel: a novel Cremphor-EL-free formulation of paclitaxel. *Nanomed (Lond)*. (2007) 2:415–23. doi: 10.2217/17435889.2.4.415
2. Futamura M, Ishihara K, Nagao Y, Ogiso A, Niwa Y, Nakada T, et al. Neoadjuvant chemotherapy using nanoparticle albumin-bound paclitaxel plus trastuzumab and pertuzumab followed by epirubicin and cyclophosphamide for operable HER2-positive primary breast cancer: a multicenter phase II clinical trial (PerSeUS-BC04). *Breast Cancer*. (2023) 30:293–301. doi: 10.1007/s12282-022-01425-2
3. Nakashima K, Umeda Y, Demura Y, Sonoda T, Tada T, Yamaguchi M, et al. Efficacy of nanoparticle albumin-bound paclitaxel (nab-PTX) monotherapy can be improved after treatment with immune checkpoint inhibitor in patients with non-small cell lung cancer: long-term follow-up and updated analysis of two previous prospective clinical studies. *Oncology*. (2024) 102:593–603. doi: 10.1159/000535994
4. Yamaguchi J, Yokoyama Y, Fujii T, Yamada S, Takami H, Kawashima H, et al. Results of a phase II study on the use of neoadjuvant chemotherapy (FOLFIRINOX or GEM/nab-PTX) for borderline-resectable pancreatic cancer (NUPAT-01). *Ann Surg*. (2022) 275:1043–49. doi: 10.1097/SLA.0000000000005430
5. Liu G, Ye Y, Jiang Y, Chen GJ, Xia W, Huang Y, et al. Nab-paclitaxel, cisplatin, and capecitabine versus cisplatin and gemcitabine as first line chemotherapy in patients with recurrent or metastatic nasopharyngeal carcinoma: randomised phase 3 clinical trial. *BMJ*. (2024) 385:e77890. doi: 10.1136/bmj-2023-077890
6. Wu D, Li Y, Xu P, Fang Q, Cao F, Lin H, et al. Neoadjuvant chemo-immunotherapy with camrelizumab plus nab-paclitaxel and cisplatin in resectable locally advanced squamous cell carcinoma of the head and neck: a pilot phase II trial. *Nat Commun*. (2024) 15:2177. doi: 10.1038/s41467-024-46444-z
7. Guo X, Sun H, Dong J, Feng Y, Li H, Zhuang R, et al. Does nab-paclitaxel have a higher incidence of peripheral neuropathy than solvent-based paclitaxel? Evidence from a systematic review and meta-analysis. *Crit Rev Oncol Hematol*. (2019) 139:16–23. doi: 10.1016/j.critrevonc.2019.04.021
8. Untch M, Jackisch C, Schneeweiss A, Conrad B, Aktas B, Denkert C, et al. Nab-paclitaxel versus solvent-based paclitaxel in neoadjuvant chemotherapy for early breast cancer (GeparSepto-GBG 69): a randomised, phase 3 trial. *Lancet Oncol*. (2016) 17:345–56. doi: 10.1016/S1470-2045(15)00542-2
9. Kida K, Yamada A, Shimada K, Narui K, Sugae S, Shimizu D, et al. A prospective comparison study utilizing patient-reported outcomes of taxane-related peripheral neuropathy between nab-paclitaxel and standard paclitaxel in patients with breast cancer. *Breast Cancer*. (2024) 31:409–16. doi: 10.1007/s12282-024-01551-z
10. Chen N, Brachmann C, Liu X, Pierce DW, Dey J, Kerwin WS, et al. Albumin-bound nanoparticle (nab) paclitaxel exhibits enhanced paclitaxel tissue distribution and tumor penetration. *Cancer Chemother Pharmacol*. (2015) 76:699–712. doi: 10.1007/s00280-015-2833-5
11. Girdenyte M, Hu Y, Ginosyan A, Hammarlund-Udenaes M, Loryan I. Formulation-dependent differences in paclitaxel distribution to anatomical sites relevant to chemotherapy-induced peripheral neuropathy. *Front Pharmacol*. (2024) 15:1486686. doi: 10.3389/fphar.2024.1486686
12. Baicus C, Purcarea A, Von E, Delcea C, Furtunescu F. Alpha-lipoic acid for diabetic peripheral neuropathy. *Cochrane Database Syst Rev*. (2024) 1:CD12967. doi: 10.1002/14651858.CD012967.pub2
13. Hsieh R, Huang I, Chen C, Sung J. Effects of oral alpha-lipoic acid treatment on diabetic polyneuropathy: A meta-analysis and systematic review. *Nutrients*. (2023) 15:3634. doi: 10.3390/nu15163634
14. Zhou L, Yang H, Wang J, Liu Y, Xu Y, Xu H, et al. The therapeutic potential of antioxidants in chemotherapy-induced peripheral neuropathy: evidence from preclinical and clinical studies. *Neurotherapeutics*. (2023) 20:339–58. doi: 10.1007/s13311-023-01346-8
15. Melli G, Taiana M, Camozzi F, Triolo D, Podini P, Quattrini A, et al. Alpha-lipoic acid prevents mitochondrial damage and neurotoxicity in experimental chemotherapy neuropathy. *Exp Neurol*. (2008) 214:276–84. doi: 10.1016/j.expneurol.2008.08.013
16. Sun H, Guo X, Wang Z, Wang P, Zhang Z, Dong J, et al. Alphas-lipoic acid prevents oxidative stress and peripheral neuropathy in nab-paclitaxel-treated rats through the nrf2 signalling pathway. *Oxid Med Cell Longev*. (2019) 2019:3142732. doi: 10.1155/2019/3142732
17. Huang S. Analysis of environmental pollutant Bisphenol F elicited prostate injury targets and underlying mechanisms through network toxicology, molecular docking, and multi-level bioinformatics data integration. *Toxicology*. (2024) 506:153847. doi: 10.1016/j.tox.2024.153847
18. Zhao L, Zhang H, Li N, Chen J, Xu H, Wang Y, et al. Network pharmacology, a promising approach to reveal the pharmacology mechanism of Chinese medicine formula. *J Ethnopharmacol*. (2023) 309:116306. doi: 10.1016/j.jep.2023.116306
19. Chaplan S, Bach F, Pogrel J, Chung J, Yaksh T. Quantitative assessment of tactile allodynia in the rat paw. *J Neurosci Methods*. (1994) 53:55–63. doi: 10.1016/0165-0270(94)90144-9
20. Yoon C, Wook YY, Sik NH, Ho KS, Mo CJ. Behavioral signs of ongoing pain and cold allodynia in a rat model of neuropathic pain. *Pain (Amsterdam)*. (1994) 59:369. doi: 10.1016/0304-3959(94)90023-X
21. Daina A, Michielin O, Zoete V. SwissTargetPrediction: updated data and new features for efficient prediction of protein targets of small molecules. *Nucleic Acids Res*. (2019) 47:W357–64. doi: 10.1093/nar/gkz382
22. Knox C, Wilson M, Klinger CM, Franklin M, Oler E, Wilson A, et al. DrugBank 6.0: the drugBank knowledgebase for 2024. *Nucleic Acids Res*. (2024) 52:D1265–75. doi: 10.1093/nar/gkad976
23. Davis AP, Wiegiers TC, Sciaky D, Barkalow F, Strong M, Wyatt B, et al. Comparative Toxicogenomics Database's 20th anniversary: update 2025. *Nucleic Acids Res*. (2025) 53:D1328–34. doi: 10.1093/nar/gkae883
24. Hamosh A, Amberger JS, Bocchini C, Scott AF, Rasmussen SA. Online Mendelian Inheritance in Man (OMIM®): VictorMcKusick's magnum opus. *Am J Med Genet A*. (2021) 185:3259–65. doi: 10.1002/ajmg.a.62407
25. Szklarczyk D, Nastou K, Koutrouli M, Kirsch R, Mehryary F, Hachilif R, et al. The STRING database in 2025: protein networks with directionality of regulation. *Nucleic Acids Res*. (2025) 53:D730–37. doi: 10.1093/nar/gkae1113
26. Sherman BT, Hao M, Qiu J, Jiao X, Baseler MW, Lane HC, et al. DAVID: a web server for functional enrichment analysis and functional annotation of gene lists (2021 update). *Nucleic Acids Res*. (2022) 50:W216–21. doi: 10.1093/nar/gkac194
27. Ji X, Tang J, Zhang J. Effects of salt stress on the morphology, growth and physiological parameters of *juglansmicrocarpa* L. *Seedlings Plants (Basel)*. (2022) 11:2381. doi: 10.3390/plants11182381
28. Liu Y, Yang X, Gan J, Chen S, Xiao Z, Cao Y. CB-Dock2: improved protein-ligand blind docking by integrating cavity detection, docking and homologous template fitting. *Nucleic Acids Res*. (2022) 50:W159–64. doi: 10.1093/nar/gkac394
29. Jiang S, Li H, Zhang L, Mu W, Zhang Y, Chen T, et al. Generic Diagramming Platform (GDP): a comprehensive database of high-quality biomedical graphics. *Nucleic Acids Res*. (2025) 53:D1670–76. doi: 10.1093/nar/gkae973
30. Morimoto M, Toba H, Aoyama M, Nakagawa M, Takechi H, Yoshida T, et al. Phase 1 dose-escalation study of triweekly nab-paclitaxel combined with S-1 for HER2-negative metastatic breast cancer. *Clin Breast Cancer*. (2020) 20:448–53. doi: 10.1016/j.clbc.2020.07.012
31. Jali AM, Banji D, Banji OJF, Hurubi KY, Tawhari FY, Alameer AA, et al. Navigating preclinical models and medications for peripheral neuropathy: A review. *Pharm (Basel)*. (2024) 17:1010. doi: 10.3390/ph17081010
32. Tripathi AK, Ray AK, Mishra SK, Bishen SM, Mishra H, Khurana A. Molecular and therapeutic insights of alpha-lipoic acid as a potential molecule for disease prevention. *Rev Bras Farmacogn*. (2023) 33:272–87. doi: 10.1007/s43450-023-00370-1
33. Shay KP, Moreau RF, Smith EJ, Smith AR, Hagen TM. Alpha-lipoic acid as a dietary supplement: Molecular mechanisms and therapeutic potential. *Biochim Et Biophys Acta (Bba) - Gen Subj*. (2009) 1790:1149–60. doi: 10.1016/j.bbagen.2009.07.026
34. Kim CF, Moalem-Taylor G. Interleukin-17 contributes to neuroinflammation and neuropathic pain following peripheral nerve injury in mice. *J Pain*. (2011) 12:370–83. doi: 10.1016/j.jpain.2010.08.003
35. Jiang X, Zhou R, Zhang Y, Zhu T, Li Q, Zhang W. Interleukin-17 as a potential therapeutic target for chronic pain. *Front Immunol*. (2022) 13:999407. doi: 10.3389/fimmu.2022.999407
36. Liang F, Xu Z, Ding L, Zhu Z, Chen M, Shu H, et al. Biomass fuel induces neuroinflammation and neurodegeneration via the astrocyte-microglia IL-17A/IL-17RA pathway. *J Hazard Mater*. (2025) 494:138569. doi: 10.1016/j.jhazmat.2025.138569
37. Park SE, Ferreira AFF, Kwon H, Yu J, Diniz L, Ulrich H, et al. The role of interleukin-17 in neurological disorders. *Anim Cells Syst (Seoul)*. (2025) 29:372–86. doi: 10.1080/19768354.2025.2510994
38. Wang J, Zhang N, Liu H, Wang J, Zhang Y, Su D, et al. NaHS alleviates neuropathic pain in mice by inhibiting IL-17-mediated dopamine (DA) neuron necroptosis in the VTA. *Brain Res Bull*. (2025) 220:111168. doi: 10.1016/j.brainresbull.2024.111168
39. Sun C, Zhang J, Chen L, Liu T, Xu G, Li C, et al. IL-17 contributed to the neuropathic pain following peripheral nerve injury by promoting astrocyte proliferation and secretion of proinflammatory cytokines. *Mol Med Rep*. (2017) 15:89–96. doi: 10.3892/mmr.2016.6018
40. Leung L, Cahill CM. TNF- α and neuropathic pain - a review. *J Neuroinflamm*. (2010) 7:27. doi: 10.1186/1742-2094-7-27
41. Zuehlke AD, Beebe K, Neckers L, Prince T. Regulation and function of the human HSP90AA1 gene. *Gene*. (2015) 570:8–16. doi: 10.1016/j.gene.2015.06.018
42. Lisse TS, Elias LJ, Pellegrini AD, Martin PB, Spaulding EL, Lopes O, et al. Paclitaxel-induced epithelial damage and ectopic MMP-13 expression promotes neurotoxicity in zebrafish. *Proc Natl Acad Sci*. (2016) 113:E2189–98. doi: 10.1073/pnas.1525096113
43. Staff NP, Fehrenbacher JC, Caillaud M, Damaj MI, Segal RA, Rieger S. Pathogenesis of paclitaxel-induced peripheral neuropathy: A current review of *in vitro* and *in vivo* findings using rodent and human model systems. *Exp Neurol*. (2020) 324:113121. doi: 10.1016/j.expneurol.2019.113121

44. Sharma N, Kumar P, Shukla KS, Maheshwari S. AGE RAGE pathways: cardiovascular disease and oxidative stress. *Drug Res (Stuttgart)*. (2023) 73:408–11. doi: 10.1055/a-2047-3896
45. Chen J, Peng H, Chen C, Wang Y, Sang T, Cai Z, et al. NAG-1/GDF15 inhibits diabetic nephropathy via inhibiting AGE/RAGE-mediated inflammation signaling pathways in C57BL/6 mice and HK-2 cells. *Life Sci*. (2022) 311:121142. doi: 10.1016/j.lfs.2022.121142
46. Laha D, Grant R, Mishra P, Nilubol N. The role of tumor necrosis factor in manipulating the immunological response of tumor microenvironment. *Front Immunol*. (2021) 12:656908. doi: 10.3389/fimmu.2021.656908
47. Milani D, Zauli G, Rimondi E, Celeghini C, Marmioli S, Narducci P, et al. Tumour necrosis factor-related apoptosis-inducing ligand sequentially activates pro-survival and pro-apoptotic pathways in SK-N-MC neuronal cells. *J Neurochem*. (2003) 86:126–35. doi: 10.1046/j.1471-4159.2003.01805.x