

#### **OPEN ACCESS**

EDITED BY Yadong Guo, Tongji University, China

REVIEWED BY
Qi-Jun Wu,
China Medical University, China
Liwen Feng,
Capital Medical University, China
Lingjiao Meng,
Fourth Hospital of Hebei Medical University,
China

\*CORRESPONDENCE

Xiaoyan Lin

Iinxiaoyanyan@163.com

Wei Chong

Hao Chen

<sup>†</sup>These authors have contributed equally to this work

RECEIVED 10 July 2025
ACCEPTED 20 October 2025
PUBLISHED 03 November 2025

#### CITATION

Chen X, Liu X, Xu J, Zhu X, Liu B, Yang X, Cong G, Lin X, Chong W and Chen H (2025) The inspiration of methyltransferase in RNA methylation modification for targeted therapy of malignant tumors. Front. Immunol. 16:1663423. doi: 10.3389/fimmu.2025.1663423

© 2025 Chen, Liu, Xu, Zhu, Liu, Yang, Cong,

#### COPYRIGHT

Lin, Chong and Chen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# The inspiration of methyltransferase in RNA methylation modification for targeted therapy of malignant tumors

Xinru Chen<sup>1,2†</sup>, Xinyu Liu<sup>1,2†</sup>, Jiaqi Xu<sup>3†</sup>, Xingyu Zhu<sup>1,2</sup>, Bingyao Liu<sup>1,2</sup>, Xinhao Yang<sup>1,2</sup>, Ge Cong<sup>1,2</sup>, Xiaoyan Lin<sup>4\*</sup>, Wei Chong<sup>1,2\*</sup> and Hao Chen<sup>5\*</sup>

<sup>1</sup>Department of Gastrointestinal Surgery, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, Shandong, China, <sup>2</sup>Key Laboratory of Engineering of Shandong Province, Shandong Provincial Hospital, Medical Science and Technology Innovation Center Shandong First Medical University & Shandong Academy of Medical Sciences, Jinan, Shandong, China, <sup>3</sup>Department of Pathology, Shandong Provincial Hospital, Shandong University, Jinan, Shandong, China, <sup>4</sup>Department of Pathology, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, Shandong, China, <sup>5</sup>Clinical Research Center of Shandong University, Clinical Epidemiology Unit Qilu Hospital of Shandong University, Jinan, Shandong, China

RNA methylation modification has always been a research hotspot. RNA methylation modification can regulate processes such as transcription, translation, splicing, stability, and degradation of RNA, in which effector proteins play an important role, including 'writers', 'erasers', and 'readers'. There are various types of proteins involved in cancer progression, and in recent years, research on their mechanisms of action has been increasing, providing new ideas for targeted cancer therapy. By regulating the expression of related genes and affecting signaling pathways, protein writing plays a role in promoting or inhibiting cancer in the proliferation, invasion, migration, and metastasis of different tumors, providing direction for the treatment of malignant tumors. This article reviews the mechanisms of common RNA methylation modified writers and their prospects in targeted cancer therapy.

#### KEYWORDS

RNA methylation modification, writer, malignant tumor, mechanism of action, targeted therapy

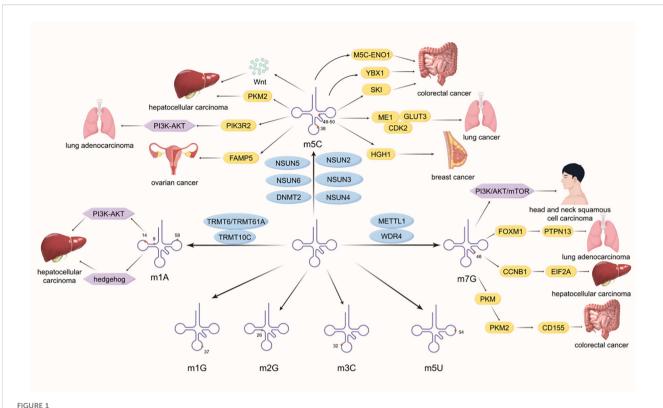
# 1 Introduction

RNA modification was first discovered and recorded in the 1950s, and now over 170 types of RNA modifications have been discovered, which are commonly present in both coding and non-coding RNA. RNA modification is mostly reversible and has functions such as splicing, localization, transport, translation, and degradation, which are crucial for regulating various

types of RNA (1, 2). At the cellular level, RNA modification is involved in various cellular processes including cell death, proliferation, aging, differentiation, migration, metabolism, autophagy, DNA damage response, and liquid-liquid phase separation (3). The abnormal expression of RNA modification can serve as a signal for the occurrence of malignant tumors, and is related to the functions of malignant tumor cell growth, proliferation, self-renewal, stress adaptation, invasion, and resistance to therapy. RNA methylation is the most common characteristic modification of RNA modification. According to the methylation sites, RNA methylation can be classified into various forms, including N6-methyladenosine (m6A), 5methylcytosine (m5C), N1-methyladenosine (m1A), N7methylguanosine(m7G), N6,2'-O-dimethyladenosine (m6Am), N4acetylcytosine (ac4C), Pseudouridine (Ψ). N6 methyladenosine (m6A) modification is a common RNA methylation modification that was first discovered in research in the 1970s and has been a hot topic in disease mechanism studies both domestically and internationally in recent years (4, 5). Many studies have shown that m6A plays an important role in human cancer development and tumorigenesis (6, 7). It is mainly related to three regulatory factors, which are the methyltransferase (writers), demethylase (erasers), and binding proteins (readers) (8, 9). Writers, also known as m6A methyltransferase complex, are composed of a variety of components, mainly including methyltransferase-like 3 (METTL3) and methyltransferase-like 14 (METTL14) with some regulatory subunits such as the Wilms' tumor 1 (WT1) associated protein (WTAP) (8). METTL3 is a 70 kda protein that is one of the important components of writers and belongs to a conserved family of methyltransferases (10). Structurally, METTL3 contains 580 amino acids in its full length and is composed of a methyltransferase domain (MTD)and a zinc finger domain (ZFD) (11). Besides, it contains Sadenosylmethionine (SAM) to catalyze methyl transfer (12). METTL3 is a monomer with catalytic activity, while METTL14 plays a structural role in RNA substrate recognition (13), and the two can form stable heterodimers (14). The heterodimers are called M6A-METTL Complex (MAC) (15) and formed by the MTDs of both. The solution structure of the ZFD of METTL3 provides methyltransferase activity of METTL3-METTL14 (16). METTL3 (11) and METTL14 (17) play a crucial role in the progression of human malignant tumors and provide new ideas for the treatment of malignant tumors, such as immunotherapy (18). WTAP can interact with METTL3 and METTL14 to regulate the nuclear speckle localization of methyltransferases and their catalytic activity in vivo (19). It can also regulate recruitment of the m6A methyltransferase complex to mRNA targets (19). Based on its multiple functions and its close relationship with the cyclical activity, metabolism and autophagy of tumor cells, WTAP has great potential in clinical cancer treatment and prognosis (20). The methyltransferase complex is also composed of KIAA1429 (17, 21), METTL5, tRNA methyltransferase activator subunit 11-2(TRMT112), Cbl proto-oncogene like 1 (CBLL1), zinc finger protein 217 (ZFP217), RNA binding motif protein 15 (RBM15), RNA binding motif protein 15B (RBM15B), vir-like m6A methyltransferase-associated protein (VIRMA), zinc finger CCCHtype containing 13 (ZC3H13), and methyltransferase-like protein 6 (METTL16) (9). METTL5 can modify 18S rRNA (22) and interact with SAM (23), which is associated with the occurrence of pancreatic cancer (24). ZC3H13 has the effect of inducing the translocation of methylase complexes into the nucleus (25). METTL16 can regulate bone marrow cell differentiation, and the absence of red blood cells can cause apoptosis, which may be associated with malignant hematologic tumors (12). In recent years, researchers have also discovered that ZCCHC4 acts as an m6A methyltransferase to modify 28S rRNA (22) and plays an important role in the translation process (26). 5methylcytosine (m5C) is also a common RNA methylation modification, widely present in tRNA, mRNAs, rRNAs, and enhancer RNA (eRNA) (27, 28). It involves adding a methyl group to the fifth carbon atom of cytosine, including the seven members of the NOL1/NOP2/Sun domain family (NSUN) NSUN1 to NSUN7 and DNA methyltransferase like 2 (DNMT2) (29). N1 methyladenosine (m1A) is the addition of a methyl group to the nitrogen atom at the first position of adenosine (30, 31), and has been found in transfer RNAs (tRNAs), rRNAs, mRNAs, lncRNAs, and mitochondrial RNAs (32, 33). M1A mainly exists in tRNA and rRNA, first identified in tRNA, usually located at positions 9, 14 and 58 of tRNA (34)(Figure 1). The methyltransferase TRMT6/61A present in tRNA recognizes the T-Loop like structure (35). Another methyltransferase responsible for m1A modification in tRNA is tRNA methyltransferase 10 homolog A (TRMT10) (36). M7G is an RNA modification that adds a methyl group to the N7 position of the nucleoside (37, 38), and is widely present in the 5'CAP structure of eukaryotic mRNA and pre tRNA (39). It plays an important role in affecting processing maturation (40), nuclear cytoplasmic transport (41), stability (42), transcription, and translation (43, 44). The methyltransferases responsible for m7G modification are mainly methyltransferase like 1 (METTL1) and its cofactor WD repeat domain 4 (WDR4) (45, 46). M5C, m1A, and m7G modifications can also play a role in the progression of malignant tumors. NSUN2 promotes the progression of lung cancer and hepatocellular carcinoma by affecting cellular metabolism (47, 48). The TRMT6/TRMT61A driving signaling pathway affects the progression of malignant tumors (49, 50). METTL1 and WDR4 promote cancer progression and poor prognosis by synergistically regulating the translation process (42, 51, 52). This review focuses on the mechanism of action of RNA writer in malignant tumors and provides inspiration and assistance for targeted therapy (Figure 1).

# 2 RNA methylation modification

More and more studies have shown that RNA methylation modified writers are closely related to the occurrence and development of various malignant tumors (53, 54), but their mechanisms of action varied in different malignant tumors. The following will discuss the mechanisms of action in different malignant tumors and thoughts on targeted therapy.



This demonstrates the distribution of different RNA methylation modifications in tRNA and the mechanism and principles of how some methyltransferases affect the development of malignant tumors. Common methylation sites include N1 methyladenosine (m1A), N3 methylcytosine (m3C), N5 methylcytosine (m5C), N1 methylguanosine (m1G), N2 methylguanosine (m2G), and N7 methylguanosine (m7G).

#### 2.1 m6A

### 2.1.1 METTL3

In recent years, a number of studies have found that METTL3 is involved in cancer development as a cancer promoting factor. METTL3 can promote the proliferation and metastasis of esophageal cancer cells (55) and renal cell carcinoma (RCC) cells (56). In addition, researchers have provided a new direction for cancer treatment through continuous exploration of its mechanism(Table 1). EPPK1 is a gene which inhabits the proliferation of esophageal cells obviously. METTL3, as a m6A writer, its deficiency can down-regulate the expression of EPPK1 and thereby inhibit the PI3K/AKT pathway, then curbing the development of esophageal cancer (57). METTL3 promotes cancer stem-like cell (CSC) phenotype in oral squamous cell carcinoma(OSCC) by targeting SALL4 through the Wnt/β-catenin signaling pathway (58). METTL3 mediates miR-99a-5p to drive OSCC cell metastasis through targeting ZBTB7A (59). METTL3 can also promote the occurrence of OSCC via m6A modification-dependent stabilization of LAMA3 transcripts (60). There are many kinds of lncRNAs, including LINC00894 (61), HOXA10-AS (62) and more. In papillary thyroid carcinoma (PTC), METTL3 can improve and enhance the stability of LINC00894 and thus promote the development of PTC through the m6A-YTHDC2-dependent pathway (61). METTL3 can modify lncRNA DSCAM-AS1 to enhance the stability of endogenous antioxidant factor SLC7A11, thereby inhibiting iron death and promoting the poor prognosis of breast cancer (63). In addition, METTL3 can promote the m6A modification of HOXA10-AS and enhance its RNA stability, thereby driving the malignant progression of laryngeal squamous cell carcinoma (LSCC) (62). LncRNA and related genes and their signaling pathways may be effective mechanistic targets for cancer treatment targeting METTL3. MicroRNAs also play a role in tumorigenesis. MiR-1908-5p is a type of microRNA that plays a role in the progression of nasopharyngeal carcinoma (NPC). METTL3 mediates the m6A modification of miR-1908-5p by targeting HOPX, thereby promoting the occurrence of NPC (64). METTL3 is involved in the m6A modification of the nuclear protein DEK, which improves the stability and expression of DEK, thereby promoting the proliferation and migration of gastric cancer (GC) cells (65). METTL3 mediates STAT5A regulation of KLF4 to promote proliferation and migration of GC cells (66). METTL3 promotes the development of ovarian cancer (OC) by increasing the mRNA level of oncogene MALAT1 through RNA binding protein ELAVL1 (67). METTL3 mediates HSPA9 m6A modification to promote the malignant development of cervical cancer (CC) (68). In another mechanism, METTL3 promotes the malignant progression of CC by up-regulating the expression of NEK2 (69). METTL3 can also enhance the stability of the glutamine transporter SLC38A1 mRNA, drive the growth of CC cells, and inhibit their apoptosis (70). METTL3 mediates Ran GTPase activating protein 1 (RanGAP1) to contribute to the progress of colorectal cancer (CRC) (71). METTL3 can promote the senescence of CRC cells through the mediator CDKN2B, providing a new target for the treatment of CRC

(72). METTL3 mediated m6A modification of matrix metalloproteinase-9 (MMP9) mRNA promotes tumorigenicity of CRC (73). METTL3 can mediate the expression of circular RNA circ\_0008345, which can bind to miR-182-5p and prevent miR-182-5p from inhibiting the oncogene CYP1A2, thereby contributing to the malignant progression of CRC (74). Meanwhile, METTL3 promotes epithelial-mesenchymal transition in CRC through m6A modification of SNAIL mRNA, where SNAIL enhances CXCL2 secretion through the NF-κB pathway to recruit M2 macrophages and promote CRC lung metastasis (75). METTL3 mediates m6A modification of exonuclease DCLRE1B to promote the development of pancreatic cancer (PC) (76). METTL3 induces m6A modification of FBXO31 and enhances its mRNA translation, thereby promoting the occurrence of PC (77). METTL3 mediates MiR-589-5p maturation and expression, which is positively correlated and over-expression can promote the deterioration of liver cancer (78). METTL3 promotes the proliferation and stemness of liver cancer stem cells (LCSCs) by targeting SOCS3 mRNA through the JAK2/STAT3 pathway (79). METTL3 mediates m6A modification of lnc-TSPAN12 to drive migration and invasion of hepatocellular carcinoma(HCC) cells, which in turn promotes hepatic metastasis (80). The transcription factor RELA can bind to the METTL3 promoter region to promote the transcription of METTL3, thereby promoting the malignant growth of HCC cells (81). Another circular RNA also plays a role in METTL13's involvement in tumor progression. In prostate cancer (PCa), METTL3 induces homeobox C6 (HOXC6) m6A modification to stabilize its expression and promote cancer cell proliferation, invasion, migration, stemness, and glycolysis (82). METTL3 mediates CircGlis3 (HSA\_CIRC\_0002874) to upregulate MDM2 and regulate the p53 signaling pathway, promoting PCa cell proliferation, migration, and invasion (83). Aromatase (CYP19A1) is an enzyme involved in limiting the rate of estrogen biosynthesis. METTL3 promotes estrogen production by enhancing CYP19A1 translation and mediates miR-196a, thereby driving nonsmall cell lung cancer (NSCLC) metastasis (84). In another mechanism, METTL3 promotes the occurrence and metastasis of NSCLC by inhibiting FDX1 by promoting the maturation of miR-21-5p (85). Due to different mechanisms, METTL3 can also play different roles in the same cancer. The over-expression of SH3 domain binding protein 5 (SH3BP5) can inhibit the invasion ability of lung cancer cells, METTL3 can mediate m6A modification to promote its expression, and METTL3 itself also has the function of inhibiting the migration and invasion of lung cancer cells, thereby METTL3 can inhibit the deterioration of lung cancer (86). METTL3 inhibits malignant progression of lung adenocarcinoma by blocking the PI3K/AKT/NFκB signaling pathway (87). METTL3 exhibits a dual role in cancer, highlighting its highly environmentally dependent function. This contradictory phenomenon may stem from METTL3's ability to modify various RNA substrates, including transcripts of oncogenes and tumor suppressor genes. The ultimate effect is likely to be the result of the combined effects of specific cellular backgrounds, tumor microenvironments, and transcriptome landscapes. This reminds us that strict biomarker stratification is necessary when applying METTL3 inhibitors in clinical practice to achieve precise treatment (Table 1).

#### 2.1.2 METTL14

METTL14, as one of the main members of m6A writers, plays an important role in the mechanisms related to malignant tumors (Table 2). METTL14 enhances the m6A modification of CDKN2A, inhibits the activation of p53 pathway, and promotes retinoblastoma (RB) progression (88). In nasopharyngeal carcinoma, METTL14 promotes cancer progression by regulating the stability of AOC1 mRNA (89). METTL14 catalyzes m6A modification of ANKRD22 mRNA to improve mRNA stability and translation efficiency. At the same time, ANKRD22 interacts with SLC25A1 to increase intracellular acetyl-CoA content, promoting lipid metabolism reprogramming and cellular lipid synthesis, thereby advancing the progression of nasopharyngeal carcinoma (90). METTL14 can up-regulate the expression of lncRNA MSTRG.292666.16 and enhance the level of m6A modification, which can promote the development of NSCLC (91). In another mechanism, METTL14 increases miR-93-5p expression and matures pri-miR-93-5p through m6A alteration to target and inhibit TXNIP, thereby inhibiting NSCLC cell apoptosis and promoting cancer development and metastasis (92). Moreover, METTL14 can target CSF1R to accelerate the proliferation, migration, and invasion of NSCLC cells (93). METTL14 activates miR-29c-3p through m6A and regulates the ubiquitination of pyruvate kinase isoform M2 (PKM2) mediated by the tripartite motif containing 9 (TRIM9), driving aerobic glycolysis of glucose and promoting the progression of triple-negative breast cancer (TNBC) (94). METTL14 mediates lncRNA RP1-228H13.5 to promote the development of liver cancer through targeting hsamiR-205 and regulating the expression of zinc finger protein interacting with K protein 1 (ZIK1) (95). In glioma cells, downregulation of METTL14 induces down-regulation of m6A modification and expression of circRNA\_103239, driving upregulation of miR-182-5p and down-regulation of MTSS1, thereby promoting EMT in glioma and promoting tumor progression (96). METTL14 targets downstream gene MN1 to enhance its stability and translation efficiency, thereby promoting malignant progression of OC (97). METTL14, by forming the METTL3-METTL14 complex, enhances the m6A modification of SETBP1 mRNA, thereby increasing the stability of SETBP1 mRNA and subsequently activating the PI3K-AKT signaling pathway to promote cell proliferation in myelodysplastic tumors (98). In addition, METTL14 also plays an equally important role in inhibiting the progression of malignant tumors. METTL14 promotes autophagy in OSCC cells and inhibits its progression by targeting the autophagy related gene RB1CC1 through m6A modification (99). METTL14 mediates miR-30c-1-3p maturation, and miR-30c-1-3p targets MARCKSL1 to inhibit its expression, thereby suppressing the development of lung cancer through this mechanism (100). METTL14 up-regulates the m6A level of lnc-PLCB1 to enhance its stability and inhibit the migration and invasion of GC cells (101). METTL14 promotes the degradation of ATF5 mRNA, reduces WDR74 transcription and  $\beta$  - catenin nuclear translocation, thereby inhibiting GC cell stemness and

TABLE 1 The mechanism of METTL3 in malignant tumors.

Malignant tumors types	Roles	Mechanism (target/signaling pathway)	References
esophageal cancer	tumor promoter	EPPK1/PI3K/AKT	(57)
OSCC	tumor promoter	SALL4/Wnt/β-catenin	(58)
OSCC	tumor promoter	miR-99a-5p/ZBTB7A	(59)
OSCC	tumor promoter	LAMA3	(60)
PTC	tumor promoter	LINC00894/m6A- YTHDC2-dependent	(61)
LSCC	tumor promoter	HOXA10-AS	(62)
breast cancer	tumor promoter	DSCAM-AS1/SLC7A11	(63)
NPC	tumor promoter	miR-1908-5p/HOPX	(64)
GC	tumor promoter	DEK	(65)
GC	tumor promoter	STAT5A/KLF4	(66)
OC	tumor promoter	MALAT1/ELAVL1	(67)
СС	tumor promoter	HSPA9	(68)
CC	tumor promoter	NEK2	(69)
СС	tumor promoter	SLC38A1	(70)
CRC	tumor promoter	RanGAP1	(71)
CRC	tumor promoter	CDKN2B	(72)
CRC	tumor promoter	MMP9	(73)
CRC	tumor promoter	miR-182-5p/CYP1A2	(74)
CRC	tumor promoter	SNAIL/CXCL2/NF-ĸB/M2	(75)
PC	tumor promoter	DCLRE1B	(76)
PC	tumor promoter	FBXO31	(77)
liver cancer	tumor promoter	miR-589-5p	(78)
liver cancer	tumor promoter	SOCS3/JAK2/STAT3	(79)

(Continued)

TABLE 1 Continued

Malignant tumors types	Roles	Mechanism (target/signaling pathway)	References
НСС	tumor promoter	lnc-TSPAN12	(80)
НСС	tumor promoter	RELA	(81)
PCa	tumor promoter	HOXC6	(82)
PCa	tumor promoter	CircGlis3/MDM2/p53	(83)
NSCLC	tumor promoter	CYP19A1/miR-196a	(84)
NSCLC	tumor promoter	FDX1/miR-21-5p	(85)
lung cancer	tumor suppressor	SH3BP5	(86)
lung cancer	tumor suppressor	PI3K/AKT/NF-κB	(87)

slowing down cancer progression (102). In colon cancer, METTL14 targets SCD1, and its over-expression significantly enhances the m6A modification of SCD1 mRNA and reduces SCD1 mRNA levels, possibly through SCD1 mediated Wnt/β- Catenin signaling inhibits the stemness and metastasis of colon cancer cells, thereby hindering the tumorigenic process of colon cancer (103). METTL14 inhibits the expression of transcription factors NANOG and βcatenin in CRC cells, thereby inhibiting the phenotype of CRC stem cells and hindering cancer progression (104). METTL14 targets FTH1 to reduce its mRNA stability, thereby enhancing sorafenib induced ferroptosis and helping to inhibit CC progression through the PI3K/Akt signaling pathway (105). Similar to METTL3, METTL14 also exhibits dual functionality. The role of METTL14 may be related to its cooperative relationship with METTL3 and other co factors. When dissociated from METTL3, METTL14 may be more inclined to stabilize tumor suppressor RNA. This suggests that targeting the METTL3-METTL14 interaction interface may be a more precise strategy than comprehensive inhibition (Table 2).

### 2.1.3 WTAP

The high expression of WTAP is closely related to the poor prognosis caused by many malignant tumors such as NSCLC (106), CRC (107). Therefore, targeted therapy research can be conducted based on the development mechanism of WTAP in different malignant tumors (Table 3). WTAP increases the stability of plasminogen activator urokinase (PLAU) and promotes the migration, invasion, and proliferation of LSCC cells (108). WTAP positively regulates the expression of m6A target PTP4A1, activates the AKT mTOR pathway, and promotes esophageal squamous cell carcinoma (ESCC) cell proliferation (109). WTAP can also promote the growth and metastasis of ESCC tumors by reducing the

TABLE 2 The mechanism of METTL14 in malignant tumors.

Malignant tumors types	Roles	Mechanism (target/signaling pathway)	References
RB	tumor promoter	CDKN2A/p53	(88)
nasopharyngeal carcinoma	tumor promoter	AOC1	(89)
nasopharyngeal carcinoma	tumor promoter	ANKRD22/SLC25A1	(90)
NSCLC	tumor promoter	MSTRG.292666.16	(91)
NSCLC	tumor promoter	miR-93-5p/TXNIP	(92)
NSCLC	tumor promoter	CSF1R	(93)
TNBC	tumor promoter	miR-29c-3p/TRIM9/PKM2	(94)
liver cancer	tumor promoter	RP1-228H13.5/hsa-miR- 205/ZIK1	(95)
glioma	tumor promoter	circRNA_103239/miR- 182-5p/MTSS1	(96)
ОС	tumor promoter	MN1	(97)
myelodysplastic tumor	tumor promoter	SETBP1/PI3K/AKT	(97)
OSCC	tumor suppressor	RB1CC1	(99)
lung cancer	tumor suppressor	miR-30c-1-3p/MARCKSL1	(100)
GC	tumor suppressor	lnc-PLCB1	(101)
GC	tumor suppressor	ATF5/WDR74/β - catenin	(102)
colon cancer	tumor suppressor	SCD1/Wnt/β- Catenin	(103)
CRC	tumor suppressor	NANOG/β - catenin	(104)
CC	tumor suppressor	FTH1/PI3K/Akt	(105)

expression of CPSF4 (110). WTAP can affect the progression of malignant tumors by mediating circular RNAs (circRNAs). WTAP mediates circEEF2 to regulate the expression of CANT1 and promote the occurrence of lung adenocarcinoma (LUAD) (111). WTAP can promote the expression of NUPR1, eliminate the role of WTAP knockdown in promoting iron death and inhibiting the malignant behavior of TNBC cells, while up-regulating LCN2 can inhibit iron death and promote the progress of TNBC (112). WTAP mediated FAM83H-AS1 promotes GC cell migration, proliferation, and invasion through m6A modification, thereby promoting cancer development (113). The Warburg effect is an abnormal energy metabolism in which cancer cells obtain energy through oxidative

glycolysis, thereby promoting proliferation and cancer progression (114, 115). WTAP can enhance the stability of HK2, accelerate the Warburg effect of GC, and promote GC progression (116). WTAP can also accelerate the migration and epithelial mesenchymal transition of GC cells by promoting the expression of transforming growth factor-beta (TGF-B) (117). WTAP inhibits downstream gene FLNA, enhances the proliferation of colon cancer cells, and inhibits cancer cell autophagy (118). Meanwhile, WTAP mediates downstream molecule VEGFA activation of MAPK signaling, accelerating CRC cell proliferation, migration, invasion, and angiogenesis (119). WTAP also mediates PDK4 and inhibits its expression to drive the proliferation, migration, and invasion of CRC cells (120). In addition, WTAP has also become an important target and research direction in immunotherapy for tumors. PD-L1 (121), as an immunosuppressive checkpoint, can promote immune escape, while WTAP can enhance its expression, strengthen the immune escape mechanism, prevent the anti-tumor effect caused by T cell proliferation (122), and promote the progression of CRC (123) (Figure 2). The decreased expression of the tumor suppressor gene PTEN leads to cellular dysregulation and loss, thereby promoting the characteristics of endometrial cancer (EC) stem cells. The reason for this is that down-regulation of WTAP reduces m6A modification of EGR1 mRNA and lowers EGR1 levels (124). Moreover, WTAP can also down-regulate the expression of CAV-1, activate the nuclear factor kappa B (NF-κB) signaling pathway in EC cells, promote EC cell proliferation, migration, and invasion, and accelerate cancer progression (125). WTAP can increase the stability of GBE1 mRNA, and overexpression of GBE1 promotes PC cell proliferation and stemnesslike properties (126). Overexpression of WTAP can counteract the tumor suppressive effect of miR-455-3p in PCa cells, thereby promoting PCa progression (127). WTAP can target NRF2 to inhibit iron death and accelerate the malignant progression of bladder cancer(BLCA) (128). In addition, the mRNA of WTAP can interact with long non-coding RNAs (lncRNA) to promote the migratory progression of malignant tumors, such as breast cancer (129). WTAP can also target lncRNA DIAPH1-AS1 to enhance its stability and promote the progression of NPC (130). In renal cell carcinoma (RCC), WTAP regulates the m6A modification of long non coding RNA TEX41 to promote cancer cell proliferation and metastasis (131). WTAP often promotes cancer progression through mechanisms involving immune escape, metabolic reprogramming, and RNA stability. The stabilizing effect of WTAP on PD-L1 mRNA under hypoxic conditions reveals a noteworthy association between m6A modification and immune therapy resistance. Targeting WTAP may enhance the efficacy of immune checkpoint blockade therapy, especially in hypoxic tumors (Figure 2, Table 3).

#### 2.1.4 METTL16

Current research indicates that METTL16 plays a dual role in cancer biology, either promoting or inhibiting tumor progression depending on the context. In lung cancer, METTL16 interacts with eIF4E2 to enhance the translation of key oncogenes, thereby driving tumor development (132). In breast cancer, METTL16 regulates

TABLE 3 The mechanism of WTAP in malignant tumors.

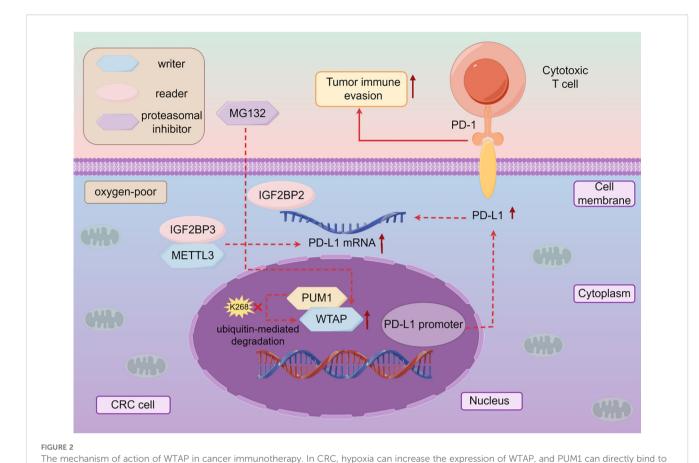
Malignant tumors types	Roles	Mechanism(target/ signaling pathway)	References
LSCC	tumor promoter	PLAU	(108)
ESCC	tumor promoter	PTP4A1/AKT mTOR	(109)
ESCC	tumor promoter	CPSF4	(110)
LUAD	tumor promoter	circEEF2/CANT1	(111)
TNBC	tumor promoter	NUPR1/LCN2	(112)
GC	tumor promoter	FAM83H-AS1	(113)
GC	tumor promoter	HK2	(116)
GC	tumor promoter	TGF-β	(117)
colon cancer	tumor promoter	FLNA	(118)
CRC	tumor promoter	VEGFA/MAPK	(119)
CRC	tumor promoter	PDK4	(120)
CRC	tumor promoter	PD-L1	(123)
EC	tumor promoter	EGR1/PTEN	(124)
EC	tumor promoter	CAV-1/NF-κB	(125)
PC	tumor promoter	GBE1	(126)
PCa	tumor promoter	miR-455-3p	(127)
BLCA	tumor promoter	NRF2	(128)
NPC	tumor promoter	DIAPH1-AS1	(130)
RCC	tumor promoter	TEX41	(131)

GPX4 expression, inhibiting ferroptosis and promoting cancer cell proliferation (133). In HCC, METTL16 targets the eukaryotic translation initiation factor 3 subunit a (eIF3a) to promote rRNA maturation and enhance mRNA translation efficiency, thereby facilitating the self-renewal and carcinogenesis of liver cancer stem cells (134). Additionally, METTL16 regulates the stability of SENP3 mRNA, with which it is positively correlated, and promotes the malignant progression of HCC through the METTL16-SENP3-LTF signaling axis by modulating ferroptosis (135). In lung cancer metastasis, METTL16 regulates the stability of SYNPO2L, mediates

the secretion of COL10A1 by cancer-associated fibroblasts (CAFs), and promotes epithelial-mesenchymal transition (EMT), making tumor cells more prone to metastasis (136). Furthermore, METTL16 is implicated in metal carcinogenesis; it targets glutamate-ammonia ligase (GLUL) to regulate glutamine metabolism, thereby promoting chromium (VI) exposure-induced tumorigenesis (137). In osteosarcoma, METTL16 targets vacuolar protein sorting protein 33b (VPS33B) for degradation, driving tumor progression (138). In CRC, METTL16 up-regulates pyruvate dehydrogenase kinase 4 (PDK4) by targeting suppressor of glucose by autophagy (SOGA1), thereby promoting CRC progression (139). In acute myeloid leukemia (AML), METTL16 drives tumor development by promoting the expression of branched-chain amino acid (BCAA) transaminases BCAT1 and BCAT2, reprogramming BCAA metabolism (140). In cholangiocarcinoma (CCA), METTL16 targets PRDM15 to regulate FGFR4 expression, promoting cell proliferation and malignant progression (141). These mechanisms collectively highlight the tumor-promoting role of METTL16 in various cancers. Conversely, METTL16 also exhibits tumor-suppressive functions in certain contexts. In PTC, METTL16 increases m6A abundance in SCD1, activates lipid metabolism, and inhibits tumor progression (142). In BLCA, METTL16 reduces the stability of prostate transmembrane protein androgen-induced 1 (PMEPA1) mRNA, inhibiting cell proliferation and cisplatin chemoresistance, thus offering a potential therapeutic strategy for BLCA (143). In epithelial ovarian cancer (EOC), METTL16 negatively correlates with lncRNA MALAT1, and their interaction up-regulates βcatenin, thereby inhibiting the growth, migration, and invasion of EOC cells (144). These findings underscore the dual role of METTL16 in cancer biology and provide valuable insights for clinical interventions and targeted cancer therapies. METTL16 is emerging as an environmentally dependent regulatory factor, playing a role in translation, metabolism, and ferroptosis. It participates in rRNA methylation, which extends the research scope of m6A beyond mRNA. The effect of METTL16 on ferroptosis suggests that it may regulate therapeutic responses, particularly to drugs that induce oxidative stress.

#### 2.1.5 RBM15

RBM15 plays a pivotal role in the progression of various cancers through diverse molecular mechanisms. In triple-negative breast cancer, RBM15 facilitates tumor growth by enhancing serine and glycine metabolism (145). In LSCC, RBM15 mediates m6A modification of TMBIM6 mRNA, increasing its stability and contributing to poor prognosis (146). In CRC, RBM15 binds to E2F2 and stabilizes its mRNA, thereby promoting malignant cellular processes (147). In CC, RBM15 drives m6A modification and protein translation of EZH2, leading to enhanced cancer cell proliferation, invasiveness, and epithelial-mesenchymal transition (EMT) (148). Additionally, RBM15 stabilizes the expression of the oncogenic lncRNA HEIH, promoting the proliferation, metastasis, and stemness of CC cells (149). In NSCLC, RBM15 up-regulates Kruppel-like factor 1 (KLF1), which suppresses the expression of TRIM13, a member of the tripartite motif (TRIM) family, and



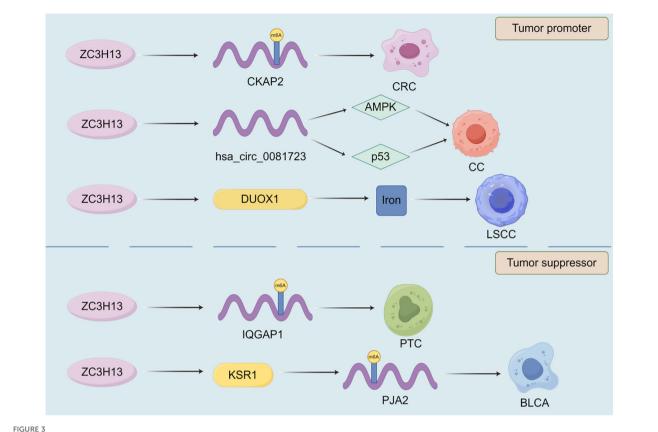
WTAP to inhibit ubiquitin mediated degradation of WTAP under hypoxic conditions, thereby enhancing the expression of PD-L1 and the stability of

IGF2BP2 and PD-L1 mRNA binding, inhibiting the anti-tumor activity of T cells, in other words, promoting cancer progression.

promotes ANXA8, a member of the annexin A (ANXA) family, thereby accelerating cancer progression (150). In LUAD, RBM15 enhances the stability of LDHA mRNA, further supporting tumor progression (151). Moreover, RBM15 up-regulates TGF-β/Smad2 expression, promoting the growth, invasion, and migration of lung cancer cells (152). In CC, RBM15 activates the AKT/mTOR signaling pathway to promote the expression of the oncogene OTUB2, driving malignant progression (153). Conversely, knockdown of RBM15 suppresses CC tumorigenesis by inhibiting the JAK-STAT pathway (154). In paclitaxel resistant ovarian cancer, silencing RBM15 down-regulates multidrug resistance 1 (MDR1) expression and inhibits malignant progression (155). Furthermore, RBM15-mediated m6A modification of MyD88 mRNA promotes CRC occurrence and metastasis, while its knockdown inhibits these processes (156). In clear cell renal cell carcinoma (ccRCC), RBM15 stabilizes CXCL11 mRNA, enhancing cell proliferation, colony formation, macrophage infiltration, and EMT (157). Interestingly, RBM15 exhibits a contrasting role in certain cancers. For instance, its elimination inhibits PC progression and enhances macrophage infiltration and phagocytosis, offering a novel direction for PC immunotherapy (158). The above mechanism suggests that RBM15 may serve as a "core stabilizer" for malignant tumors, generating diverse phenotypes in different cancer backgrounds by regulating the basic mechanism of transcriptional stability. Targeting RBM15 may simultaneously disrupt the energy supply and biosynthetic ability of tumors. In addition, the regulation of tumor immune microenvironment by RBM15 is highly environment dependent, and future immune combination strategies require precise tumor typing guidance. RBM15 inhibitors may have a synergistic effect with traditional chemotherapy drugs and serve as an important strategy for future clinical treatment.

#### 2.1.6 KIAA1429

KIAA1429 plays a significant role in cancer progression through its regulation of m6A modification and mRNA stability. In nasopharyngeal carcinoma, KIAA1429 mediates m6A modification of PTGS2, enhancing its mRNA stability and promoting cancer cell growth, proliferation, migration, and invasion (159). In LUAD, KIAA1429 increases m6A levels in ARHGAP30 mRNA, thereby driving tumor proliferation and metastasis (160). Additionally, KIAA1429 mediates LINC01106 to enhance its expression, increasing phosphorylated JAK2 and STAT3 levels, which further promotes LUAD development (161). KIAA1429 also up-regulates BTG2 expression and enhances its stability, contributing to LUAD progression (162). Furthermore, KIAA1429 promotes LUAD cell proliferation, migration, and invasion by up-regulating MUC3A expression (163). In GC,



The role of ZC3H13 in the progression of malignant tumors. Among many mechanisms, ZC3H13 can promote or inhibit the progression of malignant tumors by affecting the targeted gene m6A modification.

KIAA1429 down-regulates RASD1 expression by destabilizing its mRNA, thereby facilitating tumor growth and metastasis (164). KIAA1429 also enhances GC resistance to oxaliplatin (OXA) chemotherapy by targeting FOXM1 and stabilizing its mRNA (165). Moreover, KIAA1429 induces LINC00958 to promote aerobic glycolysis in GC cells, leading to poor prognosis (166). In HCC, KIAA1429 inhibits the expression of the tumor suppressor RND3, exerting oncogenic effects and promoting metastasis (167). Additionally, KIAA1429 targets HK1 to regulate the Warburg effect, reducing the sensitivity of liver cancer cells to the chemotherapy drug sorafenib (168). In CRC, KIAA1429 regulates lncRNA POU6F2-AS1 to promote malignant behavior (169) and reduces WEE1 stability, inhibiting its expression and driving CRC proliferation and progression (170). In colon adenocarcinoma (COAD), KIAA1429 activates the HIF-1 signaling pathway to promote tumor development (171). In ovarian cancer, KIAA1429 enhances ENO1 mRNA stability, promoting tumor progression and aerobic glycolysis (172). Finally, in multiple myeloma, KIAA1429 increases FOXM1 expression and stabilizes its mRNA, driving aerobic glycolysis and malignant behavior (173). From the above research, it can be seen that unlike some m6A writers with dual roles, KIAA1429 plays a strong pro cancer role in various types of cancer. It participates in almost all known malignant features of tumors, including proliferation, metastasis, metabolic

reprogramming, and chemotherapy resistance, by regulating different downstream targets.

# 2.1.7 Others

The expression of zinc finger CCCH-type containing 13 (ZC3H13) plays a crucial role in the development of malignant tumor (Figure 3). ZC3H13 promotes the malignant progression of CC by mediating CKAP2 m6A modification (174). ZC3H13 mediated circRNA hsa\_circ\_0081723 m6A modification promotes CC progression by regulating the AMPK/p53 pathway (175). ZC3H13 reverses the increase in iron content in LSCC cells by inhibiting the double oxidase 1 (DUOX1) gene, thereby reducing iron death (176). And, ZC3H13 can mediate m6A modification of centromere protein K (CENPK) to promote the progression of cervical cancer (177). In addition to promoting the development of malignant tumors, ZC3H13 also has inhibitory effects on them. ZC3H13 can inhibit the migration and invasion of hepatocellular carcinoma cells (178). ZC3H13 inhibits the proliferation, invasion, and migration of PTC cells by promoting the degradation of IQGAP1 mRNA (179). ZC3H13 can degrade KSR1, increase the stability of PJA2 mRNA, promote autophagy in BLCA cells, and inhibit BLCA progression (180). As one of the m6A writers, VIRMA also affects the progression of malignant tumors (Figure 4). VIRMA promotes the progression of TNBC by up regulating the expression of KIF15

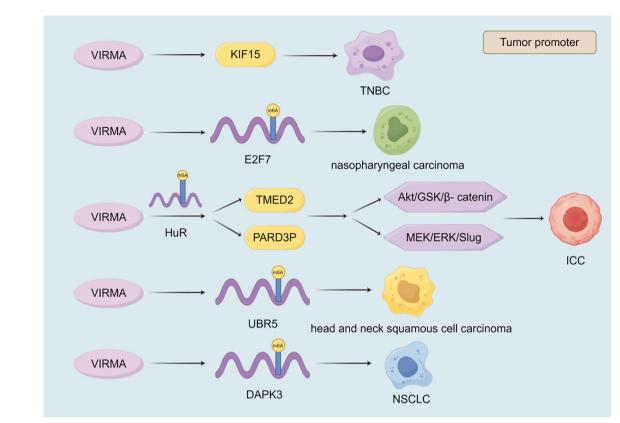


FIGURE 4

The role of VIRMA in the progression of malignant tumors. The currently discovered roles of VIRMA in malignant tumors are all promoting. VIRMA promotes the progression of malignant tumors by regulating the expression of target genes, the stability of their transcripts, and signaling pathways.

TABLE 4 The mechanism of NSUN2 in malignant tumors.

Malignant tumors types	Roles	Mechanism (target/ signaling pathway)	References
breast cancer	tumor promoter	HGH1	(186)
lung cancer	tumor promoter	ME1/GLUT3/CDK2	(47)
lung adenocarcinoma	tumor promoter	PIK3R2/PI3K-AKT	(187)
НСС	tumor promoter	PKM2	(48)
НСС	tumor promoter	Wnt	(188)
CRC	tumor promoter	YBX1/M5C-ENO1	(190)
CRC	tumor promoter	SKIL	(191)
OS	tumor promoter	FABP5	(192)

(181). VIRMA can up-regulate the expression of E2F7 and maintain the stability of E2F7 mRNA, promoting the occurrence and metastasis of nasopharyngeal carcinoma (182). VIRMA can maintain the stability of TMED2 and PARD3B through m6A HuR

mediation, activate the Akt/GSK/β- catenin and MEK/ERK/Slug signaling pathways, thereby promoting further deterioration of intrahepatic cholangiocarcinoma (ICC) (183). VIRMA promotes the progression of head and neck squamous cell carcinoma by upregulating the m6A level of UBR5 (184). VIRMA promotes NSCLC cell proliferation and malignant progression through m6A dependent degradation of DAPK3 mRNA (185). The mechanisms of these relatively less studied writers often involve the stability of lncRNA/circRNA, metabolic reprogramming, and immune regulation, often exhibiting pro cancer effects, but there are exceptions. They participate in stress adaptation, whether it is metabolic, oxidative, or immune related stress, making them potential targets for combination therapies aimed at disrupting tumor tolerance (Figures 3, 4).

#### 2.2 m5C

#### 2.2.1 NSUN2

NSUN2 plays a pivotal role in the development and progression of various cancers by regulating gene expression and enhancing mRNA stability, serving as a key hub connecting m5C modification with tumor malignancy phenotype (Table 4, Figure 5). In breast cancer, NSUN2 promotes tumorigenesis by modulating the expression of HGH1 and stabilizing its mRNA, suggesting that

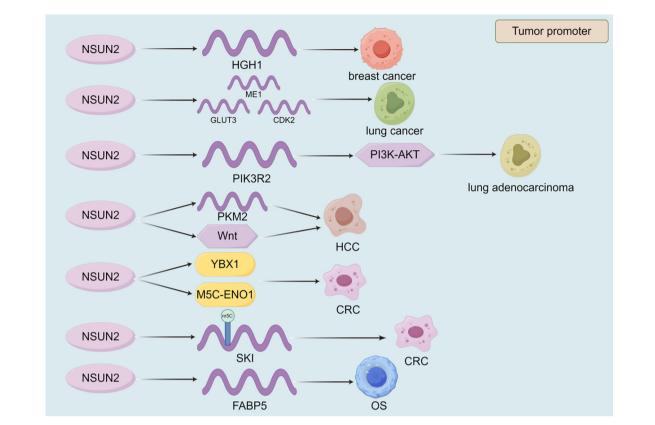


FIGURE 5

The role of NSUN2 in the progression of malignant tumors. The currently discovered roles of NSUN2 in malignant tumors are all promoting. VIRMA promotes the progression of malignant tumors by regulating the expression of target genes, the stability of their transcripts, and signaling pathways.

HGH1 could be a potential therapeutic target (186). Similarly, in lung cancer, NSUN2 facilitates metabolic reprogramming and drives Cr(VI)-induced malignant transformation by stabilizing the mRNA of key genes such as ME1, GLUT3, and CDK2 (47). Furthermore, NSUN2 up-regulates PIK3R2 and enhances its mRNA stability, leading to the activation of the PI3K-AKT signaling pathway and promoting the malignant progression of lung adenocarcinoma, highlighting its potential as a therapeutic target (187). In HCC, NSUN2 contributes to tumor progression by up-regulating PKM2 and stabilizing its mRNA, thereby promoting glycolysis and offering potential therapeutic avenues for HCC patients (48). Additionally, NSUN2 regulates the Wnt signaling pathway to enhance HCC cell proliferation and migration (188). Notably, Niu et al. discovered that NSUN2 lactylation drives drug resistance in cancer cells, suggesting that targeting NSUN2 lactylation could be an effective strategy to improve patient outcomes (189). In CRC, NSUN2 exerts its oncogenic effects through the NSUN2/YBX1/m5C-ENO1 signaling axis, providing a rationale for combining NSUN2 inhibitors in CRC treatment (190). Moreover, NSUN2 induces m5C modification of SKIL, enhancing its mRNA stability and promoting CRC progression (191). In OS, NSUN2 up-regulates FABP5 expression and stabilizes its mRNA, thereby promoting fatty acid metabolism in OS cells and driving tumor progression (192). These mechanisms indicate that

NSUN2 acts as a metabolic core regulator, reshaping glycolysis and fatty acid metabolism by stabilizing key metabolic enzymes such as PKM2 and ENO1 mRNA. NSUN2 also mediates the malignant transformation induced by environmental carcinogen Cr (VI), revealing its special significance in the occurrence of environment related tumors. These findings suggest its enormous potential as a combination therapy target, but highly selective inhibitors need to be developed to overcome the potential toxicity caused by its underlying physiological functions (Figure 5, Table 4).

#### 2.2.2 Others

Numerous m5C modification writers are implicated in the progression of malignant tumors and offer potential therapeutic insights (Table 5). For instance, knocking down NSUN3 increases the infiltration of M1 macrophages while reducing M2 macrophages, thereby promoting the progression of head and neck squamous cell carcinoma (193). In CRC, NSUN3 knockdown activates AMPK-related signaling and inhibits STAT3 signaling, leading to anti-phosphorylation and pro-apoptotic effects, which suppress CRC cell proliferation and migration (194). These findings suggest that NSUN3 inhibitors could serve as a novel therapeutic strategy for malignant tumors. NSUN4 mediates m5C modification of Circacteri3, which targets DNA binding protein 1 (DDB1), affecting mitochondrial function and

TABLE 5 The mechanism of other m5C writers in malignant tumors.

Malignant tumors types	RNA-modifying proteins	Roles	Mechanism(target/signaling pathway)	References
head and neck squamous cell carcinoma	NSUN3	tumor promoter	M1/M2	(193)
CRC	NSUN3	tumor suppressor	AMPK/STAT3	(194)
lung cancer	NSUN4	tumor promoter	DDB1	(195)
gliomas	NSUN4	tumor promoter	CDC42	(196)
HCC	NSUN5	tumor promoter	ZBED3/WNT/β-catenin	(198)
HCC	NSUN5	tumor promoter	WDR5/SMAD/H3K4me3/EMT	(199)
GC	NSUN5	tumor promoter	Wnt/β-catenin	(200)
esophageal cancer	NSUN5	tumor promoter	METTL1	(201)
lung cancer	NSUN6	tumor promoter	NM23-H1	(203)
COAD	NSUN6	tumor promoter	METTL3	(204)
cervical cancer	NSUN6	tumor promoter	NDRG1	(205)
hepatocellular carcinoma	DNMT2	tumor promoter	TNFSF10	(206)

energy metabolism, and thereby promoting lung cancer development (195). In gliomas, NSUN4 drives malignant progression by increasing the stability of CDC42 mRNA (196). Additionally, NSUN4 exacerbates liver cancer and may serve as a new prognostic marker for liver cancer (197). NSUN5 promotes HCC cell proliferation through the ZBED3/WNT/β-catenin signaling pathway (198). It also recruits WDR5 to enrich trimethylated histone H3 at lysine 4 (H3K4me3) in the SMAD3 promoter region, where SMAD3 mediates the EMT pathway, further promoting HCC progression (199). In GC, NSUN5 activates the Wnt/β-catenin signaling pathway, promoting immune escape and malignant progression (200). Moreover, NSUN5 directly binds to METTL1 transcripts, enhancing their m5C modification and driving the malignant progression of esophageal cancer (201). In glioblastoma (GBM), NSUN5 enhances protein synthesis necessary for tumor progression, thereby promoting malignancy (202). In lung cancer, NSUN6 regulates the expression of NM23-H1, promoting cancer cell proliferation, migration, and EMT (203). In COAD cells, NSUN6 up-regulates METTL3 expression and mediates its m5C modification, facilitating cancer progression (204). Additionally, NSUN6 promotes m5C modification of NDRG1 mRNA, driving resistance to radiotherapy in cervical cancer (205). Targeting NSUN6 through these mechanisms may represent a promising approach to improve the poor prognosis of malignant tumors. Lastly, DNMT2 targets the pro-apoptotic gene TNFSF10 (TRAIL), promoting the proliferation of hepatocellular carcinoma cells (206). The above mechanism indicates that different m5C writers play highly specialized roles in tumorigenesis by regulating the fate of specific RNAs. NSUN4 can affect energy metabolism through mitochondrial associated RNA (such as circERI3), revealing the

unique function of m5C at the subcellular level. NSUN3 and NSUN5 have begun to demonstrate their ability to regulate the tumor immune microenvironment, affecting macrophage polarization and T cell function, providing new targets for combined immunotherapy. In the future, it is necessary to continue exploring effective strategies targeting m5C modification to reverse tumor metabolic abnormalities and overcome immune resistance (Table 5).

#### 2.3 m1A

M1A modification is an emerging but highly promising field in the tumor epigenetic transcriptome. There is not much research on the mechanism of m1A writer in tumors, and currently only the following studies provide new strategies for the treatment of malignant tumors. TRMT6 promotes cell proliferation in HCC through the PI3K/AKT signaling pathway, providing direction for the treatment of HCC (49). In addition, TRMT6/TRMT61A can drive cholesterol synthesis to activate hedgehog signaling, promoting the occurrence of HCC (50). Another type of m1A writer TRMT10C can promote the malignant development of certain gynecological cancers and may become an effective target for treating gynecological cancers (207). Overall, m1A modification finely regulates protein translation by modifying tRNA, thereby affecting key oncogenic pathways. M1A modification may not directly cause drastic transcriptome changes, but rather exert an impact by optimizing the synthesis efficiency of specific oncogenic proteins, acting as a "translation level" amplifier. In the future, we can further explore the modification of m1A on ribosomal RNA (rRNA) and its regulation of the translation machine's own function. Based on the discovered

TABLE 6 The mechanism of m7G writers in malignant tumors.

Malignant tumors types	RNA-modifying proteins	Roles	Mechanism(target/signaling pathway)	References
head and neck squamous cell carcinoma	METTL1	tumor promoter	PI3K/AKT/ mTOR	(208)
breast cancer	METTL1	tumor suppressor	GADD45A/RB1	(209)
lung adenocarcinoma	METTL1	tumor promoter	FOXM1	(210)
CRC	METTL1	tumor promoter	PKM/PKM2/ CD155	(211)
HCC	WDR4	tumor promoter	CCNB1/EIF2A	(213)

TRMT6/TRMT61A small molecule inhibitors, we can promote their translation into preclinical therapeutic research.

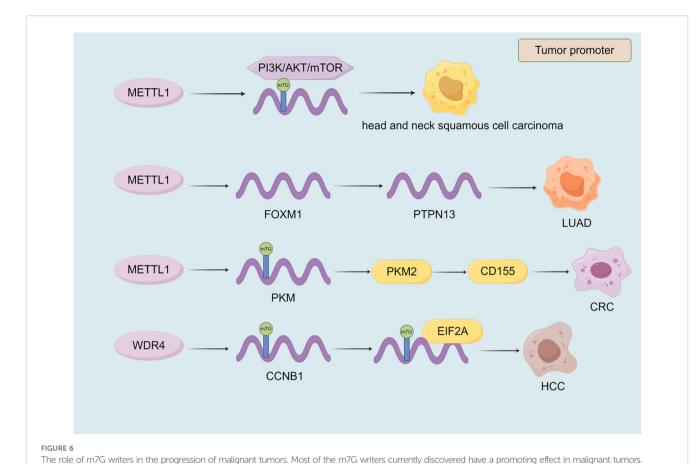
simultaneously coordinating the cell cycle, driving chemotherapy resistance, and promoting immune escape (Figure 6, Table 6).

#### 2.4 m7G

METTL1 plays a significant role in the progression of various cancers by regulating key signaling pathways, mRNA translation, and immune modulation (Table 6, Figure 6). In head and neck squamous cell carcinoma, METTL1 promotes tumor progression by regulating the PI3K/AKT/mTOR signaling pathway, enhancing mRNA translation, and influencing the immune microenvironment (208). Conversely, in breast cancer, METTL1 mediates m7G modification to regulate cell cycle arrest and the translation of genes such as GADD45A and RB1, thereby inhibiting cancer cell proliferation (209). In LUAD, METTL1 enhances the RNA stability of FOXM1 and upregulates its expression. FOXM1, in turn, inhibits PTPN13 expression, reducing the sensitivity of LUAD cells to gefitinib and driving cancer progression (210). In CRC, METTL1 mediates m7G methylation of PKM mRNA, enhancing PKM2 expression. PKM2 activates CD155 expression, inducing immune evasion and promoting CRC progression, which provides a potential direction for CRC immunotherapy (211). Additionally, METTL1 and WDR4 collaborate to promote the malignant progression of lung cancer by mediating m7G tRNA modification and regulating mRNA codon composition to enhance mRNA translation (51). In HCC, WDR4 interacts with METTL1 to synergistically regulate m7G mRNA modification, thereby promoting cancer progression (212). WDR4 also enhances the stability of CCNB1 mRNA and facilitates the binding of EIF2A to CCNB1 mRNA, enhancing CCNB1 translation and driving HCC progression (213). Similarly, in intrahepatic cholangiocarcinoma, METTL1 and WDR4 mediate m7G tRNA modification to enhance mRNA translation, promoting cancer progression and contributing to poor prognosis (42). In osteosarcoma, METTL1 and WDR4 mediate m7G modification of tRNA, enhancing mRNA translation and promoting cancer cell proliferation, migration, and chemoresistance (52). In summary, m7G modification globally reshapes the proteome at the translation level by altering tRNA abundance and codon preference, thereby

# 3 Potential clinical treatment of malignant tumors by RNA methylation modified writers

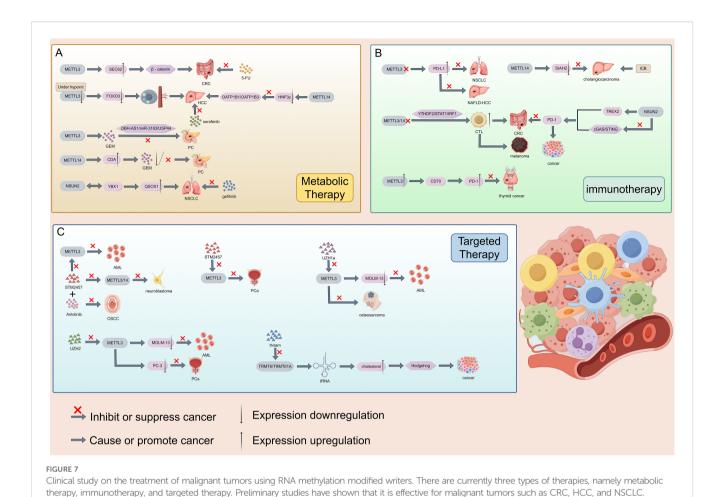
Based on the impact and molecular mechanisms of RNA methylation modification on malignant tumors mentioned above, clinical research on the treatment of malignant tumors targeting RNA methylation modification authors is becoming increasingly popular. At present, researchers are exploring the treatment of various malignant tumors related to RNA methylation modification in clinical practice through the development of metabolic therapy, immunotherapy, and targeted therapy. RNA methylation modification writer is currently involved in regulating the resistance of various malignant tumor drugs. METTL3 can promote CRC resistance to 5-FU by activating the  $\beta$  - catenin signaling pathway through the promotion of SEC62 protein expression (214). Under hypoxic conditions, METTL3 deficiency leads to a decrease in FOXO3 expression, promoting autophagy and angiogenesis, thereby promoting the progression of HCC and sorafenib resistance (215). On the contrary, METTL3, as a sensitizer in PC, can enhance the sensitivity of gemcitabin (GEM) and achieve anti-cancer effects through the DBH-AS1/miR-3163/USP44 pathway (216). In HCC, METTL14 can reduce hepatocyte nuclear factor 3- γ (HNF3γ) and inhibit the activation of drug transporters OATP1B1/OATP1B3, leading to resistance to sorafenib (217). In addition, METTL14, as a resistance factor, can upregulate the expression of cytidine deaminase (CDA) to metabolize and inactivate GEM, promoting the progression of PC (218). In NSCLC, the interaction between m5C writer NSUN2 and YBX1 can enhance the translation of QSOX1 mRNA and promote intrinsic resistance to gefitinib (219). The m6A writer is a key regulator of tumor immune therapy response, particularly in immune checkpoint blockade therapy (ICB) and adoptive cell therapy (ACT). PD-L1 is a key protein used by tumor cells to suppress T cells and achieve immune escape. In the vast majority of cases, inhibiting METTL3 can enhance the efficacy of anti-PD-1 therapy. For example, knocking out METTL3 in NSCLC and non-alcoholic fatty liver cancer-related liver cancer



METTL1 and WDR4 affect the progression of malignant tumors by regulating the expression of target genes, transcriptional stability, and signaling pathways.

(NAFLD-HCC) increases PD-L1 expression and immune cell infiltration, ultimately enhancing the anti-PD-1 effect and achieving anti-tumor efficacy (220, 221). Knocking down METTL3/14 enhances the infiltration and function of cytotoxic T cells (CTLs) through the YTHDF2/STAT1/IRF1 axis, thereby enhancing the anti-PD-1 efficacy of CRC, which also has the same effect in melanoma (222). On the contrary, in certain situations, high expression of METTL3 is beneficial for immunotherapy. For example, in thyroid cancer, high expression of METTL3 inhibits CD70 and maintains its degradation, instead relieving the inhibition of T cells and enhancing PD-1 efficacy (223). The same effect is also observed in macrophages of melanoma and lung cancer (224). In addition, METTL14 may downregulate the expression of SIAH2, making cholangiocarcinoma sensitive to ICB (225). There is relatively little research on the clinical treatment of malignant tumors with ACT, and further exploration is needed. At present, progress has shown that METTL3 can enhance the proliferation and cytotoxicity of NK cells, providing direction for future research (226). In addition, due to the correlation between m6A modified regulatory factors and T cell function, CAR-T cell therapy is also worth looking forward to (227, 228). Researchers have studied the inhibitory effect of m6A writers on certain malignant tumors in the development of targeted agents. Li et al. utilized the protein-protein interaction at the METTL3-METTL14 binding interface to select a candidate peptide RM3 with high anti-cancer activity, and then designed a fixed peptide inhibitor (RSM3) to enhance the stability of RM3, thereby achieving the anticancer effect of targeting carcinogenic METTL3 (229). Small molecule

inhibition is an important anti-cancer strategy. STM2457, as a highly effective catalytic inhibitor, targets METTL3 to promote AML apoptosis (230). In addition, STM2457 can also inhibit the progression of PCa (231) and neuroblastoma (232). STM2457 can provide new insights into the treatment of OSCC in conjunction with Anlotinib (233). There are two other METTL3 inhibitors that can also be effective in clinical treatment. UZH1a has been shown to reduce m6A/A levels in the mRNA component of leukemia cell line MOLM-13, thereby inhibiting tumor cell proliferation and exhibiting the same effect in osteosarcoma U2OS cells (234). UZH2 can also reduce the m6A/A levels of polyadenylation RNA in MOLM-13 (acute myeloid leukemia) and PC-3 (prostate cancer) cell lines (235). In addition to m6A writers' research on the clinical treatment of malignant tumors, other RNA methylation modifications also provide direction for the clinical treatment of malignant tumors. M5C writer NSUN2 can regulate the expression of TREX2 and inhibit the cGAS/STING pathway, thereby developing resistance to PD-1 blockade and providing new ideas for immunotherapy (236). In addition, researchers have identified a potent inhibitor thiram for m1A writer TRMT6/TRMT61A, which exhibits significant therapeutic effects in liver cancer treatment. Thiram inhibits TRMT6/TRMT61A in liver cancer, increases the methylation level of some tRNAs, triggers cholesterol synthesis, activates the Hedgehog signaling pathway, and promotes the process of liver cancer formation (50). At present, there is not much research on other RNA writer inhibitors, and targeting RNA methylation modified writers with inhibitors for malignant tumor



treatment may become the most direct and convenient direction. However, there are still several issues to be addressed on the future research path. Firstly, the actual effect of using inhibitors to treat patients is unknown, and currently it can only be the direction of treatment rather than the exact treatment method. Secondly, patients with malignant tumors may gradually develop resistance during the treatment with RNA methylation modified writer inhibitors, which cannot guarantee the expected therapeutic effect. Thirdly, due to individual differences in constitution, inhibitor drugs are not suitable for every patient with malignant tumors, which may narrow the scope of treatment. Finally, the selection of inhibitors also needs further evaluation to discover their potential effects. In addition, according to the above report, the promotion or blockade of targeted genes and signaling pathways within the mechanism can also become the direction of future treatment, but more detailed and lengthy research is needed, and the task is arduous and the road is long (Figure 7).

### 4 Conclusions and outlook

In this review, we mainly introduce and summarize common RNA writers and their mechanisms of action in different malignant tumors, providing direction for the treatment of malignant tumors. Firstly, in many studies, we have found that m6A writers have a dual role in the progression of malignant tumors, that is, m6A writers can

promote or inhibit the progression of malignant tumors, and the mechanisms exhibited in different malignant tumors are different. For example, METTL3 can promote the progress of esophageal cancer, colorectal cancer, pancreatic cancer and other cancers, and the mechanisms of promoting the occurrence of different targets among them are also different. More interestingly, the same m6A writer within the same malignant tumor can regulate tumor progression through different mechanisms, providing diverse targeted therapies for cancer patients as much as possible. For example, in cervical cancer, we identified three different mechanisms of METTL3 that can promote the progression of cervical cancer. HSPA9 (68), NEK2 (69), and SLC38A1 (70) are the targets corresponding to these three mechanisms and may become effective targets for cancer treatment in the future. We also found that in some studies, the same m6A writer can have opposite effects on the same cancer, such as METTL3 promoting non-small cell lung cancer (84), while also inhibiting lung cancer cell migration and suppressing lung adenocarcinoma progression (86, 87). In addition, there have been many reports on the relationship between m6A writers and long non coding RNAs in the study of their impact on the progression of malignant tumors. For example, METTL3 can promote the development of PTC by increasing the stability of LINC00894 (61), and METTL14 mediated lncRNA RP1-228H13.5 can promote the development of liver cancer (95). However, there are few reports on research related to circular RNA, only in colorectal cancer, prostate

cancer, glioma, lung adenocarcinoma, and cervical cancer. Finally, m6A writer plays a role in immunotherapy and drug-resistant therapy for malignant tumors. WTAP can enhance the immune escape mechanism and promote the occurrence of colorectal cancer (123). KIAA1429 can reduce the sensitivity of liver cancer cells to the chemotherapy drug sorafenib and enhance the resistance of gastric cancer cells to oxaliplatin chemotherapy, thereby affecting the development of cancer (237). The research on m6A writers can help us explore the occurrence of malignant tumors and provide new directions for the treatment of malignant tumors. Currently, there have been breakthroughs in the treatment direction of METTL3 inhibitors, but other aspects of research have not yet been discovered. The road to a comprehensive understanding of the relationship between m6A writers and malignant tumors is still long. In addition, the mechanisms of action of m5C, m1A, and m7G modified writers in malignant tumors have also been gradually studied, providing more possibilities for the treatment of malignant tumors. We found that immunotherapy may play a better role in exploring the treatment of m5C modified writers. For example, in head and neck squamous cell carcinoma, it acts on M1 and M2 macrophages by affecting NSUN3 (193). NSUN5 can promote immune escape in GC (200). At present, there is a lack of research on writer inhibitors for these three modifications, but there are already mechanism directions that provide inspiration. Meanwhile, emerging treatment approaches such as mRNA therapy also require RNA methylation modifications to enhance stability and avoid immunogenicity (237). Despite significant progress in research, there are still many challenges in this field: the functional background of writers is dependent, and the same writer (such as METTL3, METTL14) can play completely opposite roles in different tumors or even within the same tumor. The determining factors of upstream regulation and downstream substrate selectivity are still unclear. Writers have targeted specificity and toxicity, and they typically modify a large number of RNA transcripts. How to develop drugs that can specifically block carcinogenic modification events without interfering with global RNA homeostasis and normal physiological functions is a major challenge. Similar to other targeted therapies, tumors may eventually develop resistance to writer inhibitors. Is combination therapy strategy (such as combining with immune checkpoint inhibitors) an inevitable choice? This issue also needs to be considered. Although small molecule inhibitors such as STM2457 have been studied, their in vivo delivery efficiency, tumor targeting, and long-term safety still need to be further evaluated. Our research on circRNA is insufficient. Compared with lncRNA, the study of how writers affect tumor progression by regulating circular RNA is still in its infancy, which is a huge potential unknown field. Based on the above summary and challenges, we believe that future research should prioritize deeper mechanism exploration: using single-cell multi omics techniques to draw RNA modification profiles in the tumor microenvironment, in order to analyze the cellular background specificity of writer function. In addition, it is necessary to develop new treatment strategies and explore combination therapies, such as combining writer inhibitors with chemotherapy, radiotherapy, immunotherapy, or metabolic drugs, to overcome drug resistance. Develop new modal drugs such as

protein degradation targeted chimeras to achieve more thorough and precise clearance of writer proteins. Strengthen clinical translation and promote the clinical validation of writers or their modified signatures as predictive biomarkers to guide patient stratification and treatment selection. Actively exploring the synergistic effect of mRNA based therapy and RNA methylation modification, utilizing modification techniques to enhance the stability and translation efficiency of therapeutic mRNA. In summary, RNA methylation writers are a dynamic but extremely complex link in the cancer regulatory network. Looking ahead to the future, we hope to discover more types of m6A writers, gain a deeper understanding of the structures and mechanisms of existing molecules, closely integrate epigenetic transcriptomics, functional genomics, and clinical data, develop diversified targeted therapies for different cancers, and provide more support and choices for patients with malignant tumors.

#### **Author contributions**

XC: Writing – original draft. XLiu: Writing – review & editing. JX: Writing – review & editing. XZ: Writing – review & editing. BL: Writing – review & editing. XY: Writing – review & editing. GC: Writing – review & editing. XLin: Conceptualization, Writing – review & editing. WC: Conceptualization, Writing – review & editing. HC: Conceptualization, Writing – review & editing.

# **Funding**

The author(s) declare financial support was received for the research and/or publication of this article. This work was supported by National Natural Science Foundation of China (82473112), Natural Science Foundation of Shandong Province of China (ZR2024MH011), Youth Innovation Science and Technology Program of Shandong Provincial Universities (2022KJ187, 2023KJ018), and Special Foundation for Taishan Scholars Program of Shandong Province (No. tsqn202306373).

#### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

#### Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

#### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- 1. Roundtree IA, Evans ME, Pan T, He C. Dynamic RNA modifications in gene expression regulation. Cell. (2017) 169:1187–200. doi: 10.1016/j.cell.2017.05.045
- 2. Lin S, Kuang M. RNA modification-mediated mRNA translation regulation in liver cancer: mechanisms and clinical perspectives. *Nat Rev Gastroenterol Hepatol.* (2024) 21:267–81. doi: 10.1038/s41575-023-00884-y
- 3. Wilkinson E, Cui Y-H, He Y-Y. Roles of RNA modifications in diverse cellular functions. Front Cell Dev Biol. (2022) 10. doi: 10.3389/fcell.2022.828683
- 4. Desrosiers R FK, Rottman F. Identification of methylated nucleosides in messenger RNA from novikoffHepatoma cells. *Proc Natl Acad Sci U S A.* (1974) 71:3971–5. doi: 10.1073/pnas.71.10.3971
- 5. Adams JM CS. Modified nucleosides and bizarre 5'-termini in mouse myeloma mRNA. Nature. (1975) 255:28–33. doi: 10.1038/255028a0
- 6. An Y, Duan H. The role of m6A RNA methylation in cancer metabolism. *Mol Cancer*. (2022) 21(1):14. doi: 10.1186/s12943-022-01500-4
- 7. Feng H, Yuan X, Wu S, Yuan Y, Cui L, Lin D, et al. Effects of writers, erasers and readers within miRNA-related m6A modification in cancers. *Cell Proliferation*. (2022) 56(1):e13340. doi: 10.1111/cpr.13340
- 8. Su X, Lu R, Qu Y, Mu D. Methyltransferase-like 3 mediated RNA m6A modifications in the reproductive system: Potentials for diagnosis and therapy. *J Cell Mol Med.* (2024) 28(4):e18128. doi: 10.1111/jcmm.18128
- 9. Xie L, Zhang X, Xie J, Xu Y, Li X-J, Lin L. Emerging roles for DNA 6mA and RNA m6A methylation in mammalian genome. *Int J Mol Sci.* (2023) 24(18):13897. doi: 10.3390/ijms241813897
- $10.\,$  Su X, Qu Y, Mu D. The regulatory network of METTL3 in the nervous system: diagnostic biomarkers and therapeutic targets. Biomolecules. (2023) 13(4):664. doi: 10.3390/biom13040664
- 11. Zeng C, Huang W, Li Y, Weng H. Roles of METTL3 in cancer: mechanisms and therapeutic targeting. *J Hematol Oncol.* (2020) 13(1):117. doi: 10.1186/s13045-020-00951-w
- 12. Qi Y-N, Liu Z, Hong L-L, Li P, Ling Z-Q. Methyltransferase-like proteins in cancer biology and potential therapeutic targeting. *J Hematol Oncol.* (2023) 16(1):89. doi: 10.1186/s13045-023-01477-7
- 13. Dou X, Huang L, Xiao Y, Liu C, Li Y, Zhang X, et al. METTL14 is a chromatin regulator independent of its RNA N 6-methyladenosine methyltransferase activity. *Protein Cell.* (2023) 14:683–97. doi: 10.1093/procel/pwad009
- $14.\ Wang\ P,$  Doxtader Katelyn A, Nam Y. Structural basis for cooperative function of mettl3 and mettl14 methyltransferases. *Mol Cell.* (2016) 63:306–17. doi: 10.1016/j.molcel.2016.05.041
- 15. Chen Y, Jiang Z, Yang Y, Zhang C, Liu H, Wan J. The functions and mechanisms of post-translational modification in protein regulators of RNA methylation: Current status and future perspectives. *Int J Biol Macromol.* (2023) 253(Pt 2):126773. doi: 10.1016/j.ijbiomac.2023.126773
- 16. Huang J, Dong X, Gong Z, Qin L-Y, Yang S, Zhu Y-L, et al. Solution structure of the RNA recognition domain of METTL3-METTL14 N6-methyladenosine methyltransferase. *Protein Cell.* (2018) 10:272–84. doi: 10.1007/s13238-018-0518-7
- 17. Zhou H, Yin K, Zhang Y, Tian J, Wang S. The RNA m6A writer METTL14 in cancers: Roles, structures, and applications. *Biochim Biophys Acta (BBA) Rev Cancer*. (2021) 1876(2):188609. doi: 10.1016/j.bbcan.2021.188609
- 18. Su W, Che L, Liao W, Huang H. The RNA m6A writer METTL3 in tumor microenvironment: emerging roles and therapeutic implications. *Front Immunol.* (2024) 15. doi: 10.3389/fimmu.2024.1335774
- 19. Ping X-L, Sun B-F, Wang L, Xiao W, Yang X, Wang W-J, et al. Mammalian WTAP is a regulatory subunit of the RNA N6-methyladenosine methyltransferase. *Cell Res.* (2014) 24:177–89. doi: 10.1038/cr.2014.3
- 20. Ju G, Lei J, Cai S, Liu S, Yin X, Peng C. The emerging, multifaceted role of WTAP in cancer and cancer therapeutics. *Cancers*. (2023) 15(11):3053. doi: 10.3390/cancers15113053
- 21. Huang Q, Mo J, Liao Z, Chen X, Zhang B. The RNA m6A writer WTAP in diseases: structure, roles, and mechanisms.  $Cell\ Death\ Dis.\ (2022)\ 13(10):852.$  doi: 10.1038/s41419-022-05268-9
- 22. van Tran N, Ernst FGM, Hawley BR, Zorbas C, Ulryck N, Hackert P, et al. The human 18S rRNA m6A methyltransferase METTL5 is stabilized by TRMT112. *Nucleic Acids Res.* (2019) 47:7719–33. doi: 10.1093/nar/gkz619

- 23. Turkalj EM, Vissers C. The emerging importance of METTL5-mediated ribosomal RNA methylation. *Exp Mol Med.* (2022) 54:1617–25. doi: 10.1038/s12276-022-00869-v
- 24. Huang H, Li H, Pan R, Wang S, Khan A, Zhao Y, et al. Ribosome 18S m6A methyltransferase METTL5 promotes pancreatic cancer progression by modulating c –Myc translation. *Int J Oncol.* (2021) 60(1):9. doi: 10.3892/ijo.2021.5299
- 25. Zhu L, Zhang H, Zhang X, Xia L. RNA m<br/>6A methylation regulators in sepsis.  $Mol\ Cell\ Biochem.$  (2023). doi: 10.1007/s11010-023-04841-w
- 26. Pinto R, Vågbø CB, Jakobsson ME, Kim Y, Baltissen MP, O'Donohue M-F, et al. The human methyltransferase ZCCHC4 catalyses N6-methyladenosine modification of 28S ribosomal RNA. *Nucleic Acids Res.* (2020) 48:830–46. doi: 10.1093/nar/gkz1147
- 27. Zhang T, Zhao F, Li J, Sun X, Zhang X, Wang H, et al. Programmable RNA 5-methylcytosine (m5C) modification of cellular RNAs by dCasRx conjugated methyltransferase and demethylase. *Nucleic Acids Res.* (2024) 52:2776–91. doi: 10.1093/nar/gkae110
- 28. Ma J, Song B, Wei Z, Huang D, Zhang Y, Su J, et al. m5C-Atlas: a comprehensive database for decoding and annotating the 5-methylcytosine (m5C) epitranscriptome. *Nucleic Acids Res.* (2022) 50:D196–203. doi: 10.1093/nar/gkab1075
- 29. Bohnsack KE, Höbartner C, Bohnsack MT. Eukaryotic 5-methylcytosine (m5C) RNA methyltransferases: mechanisms, cellular functions, and links to disease. *Genes.* (2019) 10(2):102. doi: 10.3390/genes10020102
- 30. Oerum S, Dégut C, Barraud P, Tisné C. m1A post-transcriptional modification in tRNAs. *Biomolecules*. (2017) 7(1):20. doi: 10.3390/biom7010020
- 31. Li J, Zhang H, Wang H. N1-methyladenosine modification in cancer biology: Current status and future perspectives. *Comput Struct Biotechnol J.* (2022) 20:6578–85. doi: 10.1016/j.csbj.2022.11.045
- 32. Xiong X, Li X, Yi C. N1-methyladenosine methylome in messenger RNA and non-coding RNA. *Curr Opin Chem Biol.* (2018) 45:179–86. doi: 10.1016/j.cbpa.2018.06.017
- 33. Li X, Xiong X, Zhang M, Wang K, Chen Y, Zhou J, et al. Base-resolution mapping reveals distinct m1A methylome in nuclear- and mitochondrial-encoded transcripts. *Mol Cell.* (2017) 68:993–1005.e9. doi: 10.1016/j.molcel.2017.10.019
- 34. Shima HIK. N 1-methyladenosine (m1A) RNA modification: the key to ribosome control. *J Biochem.* (2020) 167:535–9. doi: 10.1093/jb/mvaa026
- 35. Safra M, Sas-Chen A, Nir R, Winkler R, Nachshon A, Bar-Yaacov D, et al. The m1A landscape on cytosolic and mitochondrial mRNA at single-base resolution. *Nature.* (2017) 551:251–5. doi: 10.1038/nature24456
- 36. Ontiveros RJ, Shen H, Stoute J, Yanas A, Cui Y, Zhang Y, et al. Coordination of mRNA and tRNA methylations by TRMT10A. *Proc Natl Acad Sci.* (2020) 117:7782–91. doi: 10.1073/pnas.1913448117
- 37. Chen W, Feng P, Song X, Lv H, Lin H. iRNA-m7G: identifying N7-methylguanosine sites by fusing multiple features. *Mol Ther Nucleic Acids.* (2019) 18:269–74. doi: 10.1016/j.omtn.2019.08.022
- 38. Komal S, Zhang L-R, Han S-N. Potential regulatory role of epigenetic RNA methylation in cardiovascular diseases. *Biomed Pharmacother*. (2021) 137:111376. doi: 10.1016/j.biopha.2021.111376
- 39. Ohira T, Suzuki T. Precursors of tRNAs are stabilized by methylguanosine cap structures. Nat Chem Biol. (2016) 12:648–55. doi: 10.1038/nchembio.2117
- 40. Pandolfini L, Barbieri I, Bannister AJ, Hendrick A, Andrews B, Webster N, et al. METTL1 Promotes let-7 MicroRNA Processing via m7G Methylation. *Mol Cell.* (2019) 74:1278–90.e9. doi: 10.1016/j.molcel.2019.03.040
- 41. Volpon I., Culjkovic-Kraljacic B, Osborne MJ, Ramteke A, Sun Q, Niesman A, et al. Importin 8 mediates m7G cap-sensitive nuclear import of the eukaryotic translation initiation factor eIF4E. *Proc Natl Acad Sci.* (2016) 113:5263–8. doi: 10.1073/pnas.1524291113
- 42. Dai Z, Liu H, Liao J, Huang C, Ren X, Zhu W, et al. N7-Methylguanosine tRNA modification enhances oncogenic mRNA translation and promotes intrahepatic cholangiocarcinoma progression. *Mol Cell.* (2021) 81:3339–55.e8. doi: 10.1016/j.molcel.2021.07.003
- 43. Zhang L-S, Liu C, Ma H, Dai Q, Sun H-L, Luo G, et al. Transcriptome-wide mapping of internal N7-methylguanosine methylome in mammalian mRNA. *Mol Cell.* (2019) 74:1304–16.e8. doi: 10.1016/j.molcel.2019.03.036

- 44. Kiriakidou M, Tan GS, Lamprinaki S, De Planell-Saguer M, Nelson PT, Mourelatos Z. An mRNA m7G Cap Binding-like Motif within Human Ago2 Represses Translation. *Cell.* (2007) 129:1141–51. doi: 10.1016/j.cell.2007.05.016
- 45. Jin X, Guan Z, Hu N, He C, Yin P, Gong Z, et al. Structural insight into how WDR4 promotes the tRNA N7-methylguanosine methyltransferase activity of METTL1. *Cell Discov.* (2023) 9(1):65. doi: 10.1038/s41421-023-00562-y
- 46. Li J, Wang L, Hahn Q, Nowak RP, Viennet T, Orellana EA, et al. Structural basis of regulated m7G tRNA modification by METTL1-WDR4. *Nature*. (2023) 613:391–7. doi: 10.1038/s41586-022-05566-4
- 47. Zhang R-K, Li Y, Sun F-L, Zhou Z-H, Xie Y-X, Liu W-J, et al. RNA methyltransferase NSUN2-mediated m5C methylation promotes Cr(VI)-induced Malignant transformation and lung cancer by accelerating metabolism reprogramming. *Environ Int.* (2024) 192:109055. doi: 10.1016/j.envint.2024.109055
- 48. Qi Q, Zhong R, Huang Y, Tang Y, Zhang X-W, Liu C, et al. The RNA M5C methyltransferase NSUN2 promotes progression of hepatocellular carcinoma by enhancing PKM2-mediated glycolysis. *Cell Death Dis.* (2025) 16(1):82. doi: 10.1038/s41419-025-07414-5
- 49. Ye Y, Liu M, Wu F, Ou S, Wang W, Fei J, et al. TRMT6 promotes hepatocellular carcinoma progression through the PI3K/AKT signaling pathway. *Eur J Med Res.* (2023) 28(1):48. doi: 10.1186/s40001-022-00951-1
- 50. Wang Y, Wang J, Li X, Xiong X, Wang J, Zhou Z, et al. N1-methyladenosine methylation in tRNA drives liver tumourigenesis by regulating cholesterol metabolism. *Nat Commun.* (2021) 12(1):6314. doi: 10.1038/s41467-021-26718-6
- 51. Ma J, Han H, Huang Y, Yang C, Zheng S, Cai T, et al. METTL1/WDR4-mediated m7G tRNA modifications and m7G codon usage promote mRNA translation and lung cancer progression. *Mol Ther.* (2021) 29:3422–35. doi: 10.1016/j.ymthe.2021.08.005
- 52. Wang Z, Yu P, Zou Y, Ma J, Han H, Wei W, et al. METTL1/WDR4-mediated tRNA m7G modification and mRNA translation control promote oncogenesis and doxorubicin resistance. *Oncogene*. (2023) 42:1900–12. doi: 10.1038/s41388-023-02695-6
- 53. Han M, Sun H, Zhou Q, Liu J, Hu J, Yuan W, et al. Effects of RNA methylation on Tumor angiogenesis and cancer progression. *Mol Cancer*. (2023) 22(1):198. doi: 10.1186/s12943-023-01879-8
- 54. Jiang X, Liu B, Nie Z, Duan L, Xiong Q, Jin Z, et al. The role of m6A modification in the biological functions and diseases. Signal Transduct Targeted Ther. (2021) 6(1):74. doi: 10.1038/s41392-020-00450-x
- 55. Qiu Y, Tian Z, Miao T-Y, Shen L, Chen J, Li P-F, et al. The METTL3-m6A-YTHDC1-AMIGO2 axis contributes to cell proliferation and migration in esophageal squamous cell carcinoma. *Gene.* (2024) 908:148281. doi: 10.1016/j.gene.2024.148281
- 56. Chen Y, He Y, Li Z, Zhang N, Zhou C, He X, et al. METTL3 facilitates renal cell carcinoma progression by PLOD2 m6A-methylation under prolonged hypoxia. *Cell Death Dis.* (2024) 15(1):62. doi: 10.1038/s41419-023-06411-w
- 57. Jia J, Yu L. METTL3-mediated m6A modification of EPPK1 to promote the development of esophageal cancer through regulating the PI3K/AKT pathway. *Environ Toxicol.* (2024) 39:2830–41. doi: 10.1002/tox.24158
- 58. Huang J, Li H, Yang Z, Liu R, Li Y, Hu Y, et al. SALL4 promotes cancer stem-like cell phenotype and radioresistance in oral squamous cell carcinomas via methyltransferase-like 3-mediated m6A modification. *Cell Death Dis.* (2024) 15 (2):139. doi: 10.1038/s41419-024-06533-9
- 59. Huang Y, Guan Y, Zhang X. METTL3-mediated maturation of miR-99a-5p promotes cell migration and invasion in oral squamous cell carcinoma by targeting ZBTB7A. *Mol Biotechnol.* (2023) 66(8):1942–53. doi: 10.1007/s12033-023-00815-x
- 60. Baoshan Ning YM. LAMA3 promotes tumorigenesis of oral squamous cell carcinoma by METTL3-mediated N6-methyladenosine modification. *Crit ReviewsTM Immunol.* (2024) 44:49–59. doi: 10.1615/CritRevImmunol.2023051066
- 61. Zhou X, Chang L, Liang Q, Zhao R, Xiao Y, Xu Z, et al. The m6A methyltransferase METTL3 drives thyroid cancer progression and lymph node metastasis by targeting LINC00894. *Cancer Cell Int.* (2024) 24(1):47. doi: 10.1186/s12935-024-03240-5
- 62. Zhao K, Chen L, Xie Y, Ren N, Li J, Zhai X, et al. m6A/HOXA10-AS/ITGA6 axis aggravates oxidative resistance and Malignant progression of laryngeal squamous cell carcinoma through regulating Notch and Keap1/Nrf2 pathways. *Cancer Lett.* (2024) 587:216735. doi: 10.1016/j.canlet.2024.216735
- 63. Yan Z, Liang Z, Luo K, Yu L, Chen C, Yu M, et al. METTL3-modified lncRNA DSCAM-AS1 promotes breast cancer progression through inhibiting ferroptosis. *J Bioenerget Biomembranes*. (2024) 56:451–9. doi: 10.1007/s10863-024-10024-z
- 64. Zhou Y, Li W. Methyltransferase-like 3-mediated m6A modification of miR-1908-5p contributes to nasopharyngeal carcinoma progression by targeting homeodomain-only protein homeobox. *Environ Toxicol.* (2023) 39:1631–40. doi: 10.1002/tox.24032
- 65. Zhang H-M, Qi F-F, Wang J, Duan Y-Y, Zhao L-L, Wang Y-D, et al. The m6A Methyltransferase METTL3-Mediated N6-Methyladenosine Modification of DEK mRNA to Promote Gastric Cancer Cell Growth and Metastasis. *Int J Mol Sci.* (2022) 23(12):6451. doi: 10.3390/ijms23126451
- 66. Zang Y, Tian Z, Wang D, Li Y, Zhang W, Ma C, et al. METTL3-mediated N6-methyladenosine modification of STAT5A promotes gastric cancer progression by regulating KLF4. Oncogene. (2024) 43:2338–54. doi: 10.1038/s41388-024-03085-2

- 67. Xiong J, Lian W, Zhao R, Gao K. METTL3/MALAT1/ELAVL1 axis promotes tumor growth in ovarian cancer. *OncoTarg Ther.* (2024) 17:85–97. doi: 10.2147/OTT \$431810
- 68. Ao K, Yin M, Lyu X, Xiao Y, Chen X, Zhong S, et al. METTL3-mediated HSPA9 m6A modification promotes Malignant transformation and inhibits cellular senescence by regulating exosomal mortalin protein in cervical cancer. *Cancer Lett.* (2024) 587:216658. doi: 10.1016/j.canlet.2024.216658
- 69. Guo Y, Bai Y, Wang L, Xu Z, Zhang N, Wang W, et al. METTL3 facilitates the progression of cervical cancer by m6A modification-mediated up-regulation of NEK2. *Sci Rep.* (2024) 14(1):24469. doi: 10.1038/s41598-024-73601-7
- 70. Liu H-T, Zhao Y, Wang H-C, Liu Q-L. METTL3-mediated m6A methylation of SLC38A1 stimulates cervical cancer growth. *Biochem Biophys Res Commun.* (2024) 716:150039. doi: 10.1016/j.bbrc.2024.150039
- 71. Yang R, Yang C, Su D, Song Y, Min J, Qian Z, et al. METTL3-mediated RanGAP1 promotes colorectal cancer progression through the MAPK pathway by recruiting YTHDF1. *Cancer Gene Ther*. (2024) 31:562–73. doi: 10.1038/s41417-024-00731-5
- 72. Chen Z, Zhou J, Wu Y, Chen F, Li J, Tao L, et al. METTL3 promotes cellular senescence of colorectal cancer via modulation of CDKN2B transcription and mRNA stability. *Oncogene*. (2024) 43:976–91. doi: 10.1038/s41388-024-02956-y
- 73. Chen J, Wu H, Zuo T, Wu J, Chen Z. METTL3–mediated N6–methyladenosine modification of MMP9 mRNA promotes colorectal cancer proliferation and migration. Oncol Rep. (2024) 53(1):9. doi: 10.3892/or.2024.8842
- 74. Hou C, Liu J, Liu J, Yao D, Liang F, Qin C, et al. METTL3-induced circ\_0008345 contributes to the progression of colorectal cancer via the microRNA-182-5p/CYP1A2 pathway. *BMC Cancer*. (2024) 24(1):728. doi: 10.1186/s12885-024-12474-5
- 75. Ouyang P, Li K, Xu W, Chen C, Shi Y, Tian Y, et al. METTL3 recruiting M2-type immunosuppressed macrophages by targeting m6A-SNAIL-CXCL2 axis to promote colorectal cancer pulmonary metastasis. *J Exp Clin Cancer Res.* (2024) 43(1):111. doi: 10.1186/s13046-024-03035-6
- 76. Li L, Wang F, Deng Z, Zhang G, Zhu L, Zhao Z, et al. DCLRE1B promotes tumor progression and predicts immunotherapy response through METTL3-mediated m6A modification in pancreatic cancer. *BMC Cancer*. (2023) 23(1):1073. doi: 10.1186/s12885-023-11524-8
- 77. Chen K, Wang Y, Dai X, Luo J, Hu S, Zhou Z, et al. FBXO31 is upregulated by METTL3 to promote pancreatic cancer progression via regulating SIRT2 ubiquitination and degradation. *Cell Death Dis.* (2024) 15(1):37. doi: 10.1038/s41419-024-06425-y
- 78. Liu J, Jiang K. METTL3-mediated maturation of miR-589-5p promotes the Malignant development of liver cancer. J Cell Mol Med. (2022) 26:2505–19. doi: 10.1111/jcmm.16845
- 79. Hu J, Chen K, Hong F, Gao G, Dai X, Yin H. METTL3 facilitates stemness properties and tumorigenicity of cancer stem cells in hepatocellular carcinoma through the SOCS3/JAK2/STAT3 signaling pathway. *Cancer Gene Ther.* (2023) 31:228–36. doi: 10.1038/s41417-023-00697-w
- 80. Li B, Xiong X, Xu J, Peng D, Nie G, Wen N, et al. METTL3-mediated m6A modification of lncRNA TSPAN12 promotes metastasis of hepatocellular carcinoma through SENP1-depentent deSUMOylation of EIF3I. *Oncogene*. (2024) 43:1050–62. doi: 10.1038/s41388-024-02970-0
- 81. Wu J, Wang W, Zheng Y, Deng W, Liu J. Transcription factor RELA promotes hepatocellular carcinoma progression by promoting the transcription of m6A modulator METTL3. *Pathol Res Practice*. (2024) 255:155168. doi: 10.1016/j.prp.2024.155168
- 82. He P, Liu X, Yu G, Wang Y, Wang S, Liu J, et al. METTL3 facilitates prostate cancer progression via inducing HOXC6 m6A modification and stabilizing its expression through IGF2BP2-dependent mechanisms. *Mol Cell Biochem.* (2024) 479:1707–20. doi: 10.1007/s11010-024-05023-y
- 83. Cheng X, Yang H, Chen Y, Zeng Z, Liu Y, Zhou X, et al. METTL3-mediated m6A modification of circGLIS3 promotes prostate cancer progression and represents a potential target for ARSI therapy. *Cell Mol Biol Lett.* (2024) 29(1):109. doi: 10.1186/s11658-024-00628-z
- 84. Yang Z, Hao J, Qiu M, Liu R, Mei H, Zhang Q, et al. The METTL3/miR-196a axis predicts poor prognosis in non-small cell lung cancer. J Cancer. (2024) 15:1603–12. doi: 10.7150/jca.92968
- 85. Qian S, Liu J, Liao W, Wang F. METTL3 promotes non-small-cell lung cancer growth and metastasis by inhibiting FDX1 through copper death-associated pri-miR-21-5p maturation. *Epigenomics*. (2023) 15:1237–55. doi: 10.2217/epi-2023-0230
- 86. Zhang B, Qian R, Li X. METTL3 suppresses invasion of lung cancer via SH3BP5 m6A modification. *Arch Biochem Biophysics*. (2024) 752:109876. doi: 10.1016/j.abb.2023.109876
- 87. Zhou YF, Li JT, Zheng QL, Ren KL, Yi CC. METTL3-mediated m6A methylation of TRAF5 inhibits lung adenocarcinoma cell metastasis via activation of the PI3K/ AKT/NF- $\kappa$ B signaling pathway. Kaohsiung J Med Sci. (2023) 40(2):150–60. doi: 10.1002/kjm2.12791
- 88. Jing Chen BZ. METTL14-Mediated m6a Modification of CDKN2A Promotes the Development of Retinoblastoma by Inhibiting the p53 Pathway. *Crit ReviewsTM Immunol.* (2024) 44:89–98. doi: 10.1615/CritRevImmunol.2023052059

- 89. Hu C, Song S, Zhao S, Xue Z, Zhu X. METTL14 contributes to the progression of nasopharyngeal carcinoma through regulating the stability of AOC1 mRNA. *Hereditas*. (2024) 161(1):20. doi: 10.1186/s41065-024-00317-z
- 90. Li L, Tang Q, Ge J, Wang D, Mo Y, Zhang Y, et al. METTL14 promotes lipid metabolism reprogramming and sustains nasopharyngeal carcinoma progression via enhancing m6A modification of ANKRD22 mRNA. *Clin Trans Med.* (2024) 14(7): e1766. doi: 10.1002/ctm2.1766
- 91. Ji X, Wan X, Sun H, Deng Q, Meng S, Xie B, et al. METTL14 enhances the m6A modification level of lncRNA MSTRG.292666.16 to promote the progression of non-small cell lung cancer. *Cancer Cell Int.* (2024) 24(1):61. doi: 10.1186/s12935-024-03250-3
- 92. Qian S, Liu J, Liao W, Wang F. METTL14 drives growth and metastasis of non-small cell lung cancer by regulating pri-miR-93-5p maturation and TXNIP expression. *Genes Genomics.* (2023) 46:213–29. doi: 10.1007/s13258-023-01436-z
- 93. Ren S, Xiao Y, Yang L, Hu Y. RNA m6A methyltransferase METTL14 promotes the procession of non-small cell lung cancer by targeted CSF1R. *Thorac Cancer*. (2022) 14:254–66. doi: 10.1111/1759-7714.14741
- 94. Wu H, Jiao Y, Guo X, Wu Z, Lv Q. METTL14/miR-29c-3p axis drives aerobic glycolysis to promote triple-negative breast cancer progression though TRIM9-mediated PKM2 ubiquitination. *J Cell Mol Med.* (2024) 28(3):e18112. doi: 10.1111/jcmm.18112
- 95. Xu J, Liu C, Qu K, Zhang J, Liu S, Meng F, et al. m6A methyltransferase METTL14–mediated RP1–228H13.5 promotes the occurrence of liver cancer by targeting hsa–miR–205/ZIK1. *Oncol Rep.* (2024) 51(4):59. doi: 10.3892/or.2024.8718
- 96. Zhang S, Zhang P, Wu A, Xu Z, Huang S, Liu X, et al. Downregulated M6A modification and expression of circRNA\_103239 promoted the progression of glioma by regulating the miR-182-5p/MTSS1 signalling pathway. *J Cancer*. (2023) 14:3508–20. doi: 10.7150/jca.85320
- 97. Li H-B, Huang G, Tu J, Lv D-M, Jin Q-L, Chen J-K, et al. METTL14-mediated epitranscriptome modification of MN1 mRNA promote tumorigenicity and all-transretinoic acid resistance in osteosarcoma. *eBioMedicine*. (2022) 82:104142. doi: 10.1016/j.ebiom.2022.104142
- 98. Jiang L, Zhang Y, Qian J, Zhou X, Ma L, Zhu S, et al. The m6A methyltransferase METTL14 promotes cell proliferation via SETBP1-mediated activation of PI3K-AKT signaling pathway in myelodysplastic neoplasms. *Leukemia*. (2024) 38(10):2246–58. doi: 10.1038/s41375-024-02350-3
- 99. Liang J, Cai H, Hou C, Song F, Jiang Y, Wang Z, et al. METTL14 inhibits Malignant progression of oral squamous cell carcinoma by targeting the autophagy-related gene RB1CC1 in an m6A-IGF2BP2-dependent manner. *Clin Sci.* (2023) 137:1373–89. doi: 10.1042/CS20230219
- 101. Chang M, Cui X, Sun Q, Wang Y, Liu J, Sun Z, et al. Lnc-PLCB1 is stabilized by METTL14 induced m6A modification and inhibits Helicobacter pylori mediated gastric cancer by destabilizing DDX21. *Cancer Lett.* (2024) 588:216746. doi: 10.1016/j.canlet.2024.216746
- 102. Zhang P, Xiang H, Peng Q, Ma L, Weng C, Liu G, et al. METTL14 attenuates cancer stemness by suppressing ATF5/WDR74/ $\beta$ -catenin axis in gastric cancer. *Cancer Sci.* (2024) 116:112–27. doi: 10.1111/cas.16381
- 103. Xu D, Han S, Yue X, Xu X, Huang T. METTL14 suppresses tumor stemness and metastasis of colon cancer cells by modulating m6A-modified SCD1. *Mol Biotechnol.* (2023) 66:2095–105. doi: 10.1007/s12033-023-00843-7
- 104. Sun C-I., Chen J, Xing Z-W, Tao G-S. METTL14 suppresses cancer stem cell phenotype of colorectal cancer via regulating of  $\beta$ -catenin/NANOG. *J Cancer*. (2023) 14:1407–16. doi: 10.7150/jca.82158
- 105. Li L, Zeng J, He S, Yang Y, Wang C. METTL14 decreases FTH1 mRNA stability via m6A methylation to promote sorafenib-induced ferroptosis of cervical cancer. *Cancer Biol Ther.* (2024) 25(1):2349429. doi: 10.1080/15384047.2024.2349429
- 106. Liu C, Gao F, Yang J, Liu C, Tian Z. Wilms' Tumor 1-associating protein promotes nonsmall-cell lung cancer through the expression of carcinoembryonic antigen-related cell adhesion molecule 5. *Am J Clin Oncol.* (2024) 47(10):465–74. doi: 10.1097/COC.0000000000001116
- 107. Tan Y-T, Li T, Wang R-B, Liu Z-K, Ma M-Y, Huang R-Z, et al. WTAP weakens oxaliplatin chemosensitivity of colorectal cancer by preventing PANoptosis. *Cancer Lett.* (2024) 604:217254. doi: 10.1016/j.canlet.2024.217254
- 108. Liu F, Xia F, Wang C, Ren R, Zhang M, Gong J, et al. m6A methyltransferase, WTAP, promotes cancer progression in laryngeal squamous cell carcinoma by regulating PLAU stability. *Ann Clin Lab Sci.* (2023) 53:293–302.
- 109. Zou J, Ma Q, Gao C, Yang M, Wen J, Xu L, et al. WTAP promotes proliferation of esophageal squamous cell carcinoma via m6A-dependent epigenetic promoting of PTP4A1. *Cancer Sci.* (2024) 115:2254–68. doi: 10.1111/cas.15924
- 110. Luo Q, Zhan X, Kuang Y, Sun M, Dong F, Sun E, et al. WTAP promotes oesophageal squamous cell carcinoma development by decreasing CPSF4 expression in an m6A-dependent manner. *Med Oncol.* (2022) 39(12):231. doi: 10.1007/s12032-022-01830-9

- 111. Zheng H, Cao Z, Lv Y, Cai X. WTAP-mediated N6-methyladenine modification of circEEF2 promotes lung adenocarcinoma tumorigenesis by stabilizing CANT1 in an IGF2BP2-dependent manner. *Mol Biotechnol.* (2024) 67 (4):1494–508. doi: 10.1007/s12033-024-01134-5
- 112. Tan M, He Y, Yi J, Chen J, Guo Q, Liao N, et al. WTAP mediates NUPR1 regulation of LCN2 through m6A modification to influence ferroptosis, thereby promoting breast cancer proliferation, migration and invasion. *Biochem Genet*. (2023) 62:876–91. doi: 10.1007/s10528-023-10423-8
- 113. Liu N, Zhang C, Zhang L. WTAP-Involved the m6A Modification of lncRNA FAM83H-AS1 Accelerates the Development of Gastric Cancer. *Mol Biotechnol.* (2023) 66:1883–93. doi: 10.1007/s12033-023-00810-2
- 114. Fukushi A, Kim H-D, Chang Y-C, Kim C-H. Revisited metabolic control and reprogramming cancers by means of the warburg effect in tumor cells. *Int J Mol Sci.* (2022) 23(17):10037. doi: 10.3390/ijms231710037
- 115. Alberghina L. The warburg effect explained: integration of enhanced glycolysis with heterogeneous mitochondria to promote cancer cell proliferation. *Int J Mol Sci.* (2023) 24(21):15787. doi: 10.3390/ijms242115787
- 116. Yu H, Zhao K, Zeng H, Li Z, Chen K, Zhang Z, et al. N6-methyladenosine (m6A) methyltransferase WTAP accelerates the Warburg effect of gastric cancer through regulating HK2 stability. *Biomed Pharmacother*. (2021) 133:111075. doi: 10.1016/j.biopha.2020.111075
- 117. Liu Y, Da M. Wilms tumor 1 associated protein promotes epithelial mesenchymal transition of gastric cancer cells by accelerating TGF- $\beta$  and enhances chemoradiotherapy resistance. *J Cancer Res Clin Oncol.* (2022) 149:3977–88. doi: 10.1007/s00432-022-04320-7
- 118. Huang L, Shao J, Xu X, Hong W, Yu W, Zheng S, et al. WTAP regulates autophagy in colon cancer cells by inhibiting FLNA through N6-methyladenosine. *Cell Adhesion Migration*. (2023) 17:1–13. doi: 10.1080/19336918.2023.2180196
- 119. Ye M, Chen J, Yu P, Hu C, Wang B, Bao J, et al. WTAP activates MAPK signaling through m6A methylation in VEGFA mRNA-mediated by YTHDC1 to promote colorectal cancer development. *FASEB J.* (2023) 37(8):e23090. doi: 10.1096/fi.202300344RRR
- 120. Dai X, Chen K, Xie Y. WTAP mediated the N6-methyladenosine modification of PDK4 to regulate the Malignant behaviors of colorectal cancer cells *in vitro and in vivo. Curr Med Chem.* (2023) 30:3368–81. doi: 10.2174/0929867329666220922102949
- 121. Chong W, Ren H, Chen H, Xu K, Zhu X, Liu Y, et al. Clinical features and molecular landscape of cuproptosis signature-related molecular subtype in gastric cancer. *iMeta*. (2024) 3(3):e190. doi: 10.1002/imt2.190
- 122. Zhou F, Huang L, Li S, Yang W, Chen F, Cai Z, et al. From structural design to delivery: mRNA therapeutics for cancer immunotherapy. *Exploration*. (2023) 4 (2):20210146. doi: 10.1002/EXP.20210146
- 123. Liu QZ, Zhang N, Chen JY, Zhou MJ, Zhou DH, Chen Z, et al. WTAP-induced N6-methyladenosine of PD-L1 blocked T-cell-mediated antitumor activity under hypoxia in colorectal cancer. *Cancer Sci.* (2024) 115:1749–62. doi: 10.1111/cas.16136
- 124. Wang B, Wang Y, Wang W, Wang Z, Zhang Y, Pan X, et al. WTAP/IGF2BP3 mediated m6A modification of the EGRI/PTEN axis regulates the Malignant phenotypes of endometrial cancer stem cells. *J Exp Clin Cancer Res.* (2024) 43 (1):204. doi: 10.1186/s13046-024-03120-w
- 125. Li Q, Wang C, Dong W, Su Y, Ma Z. WTAP facilitates progression of endometrial cancer via CAV-1/NF-κB axis. *Cell Biol Int.* (2021) 45:1269–77. doi: 10.1002/cbin.11570
- 126. Jin W, Yao Y, Fu Y, Lei X, Fu W, Lu Q, et al. WTAP/IGF2BP3-mediated GBE1 expression accelerates the proliferation and enhances stemness in pancreatic cancer cells via upregulating c-Myc. *Cell Mol Biol Lett.* (2024) 29. doi: 10.1186/s11658-024-00611-8
- 127. Zhao P, Han P, Ma Y, Tian P, Li J. Circ\_0082878 contributes to prostate cancer progression via the miR-455-3p/WTAP axis. *Environ Toxicol.* (2023) 39:979–90. doi: 10.1002/tox.24031
- 128. Wang K, Wang G, Li G, Zhang W, Wang Y, Lin X, et al. m6A writer WTAP targets NRF2 to accelerate bladder cancer Malignancy via m6A-dependent ferroptosis regulation. *Apoptosis*. (2023) 28:627–38. doi: 10.1007/s10495-023-01817-5
- 129. Dragonetti M, Turco C, Benedetti A, Goeman F, Forcato M, Scalera S, et al. The IncRNAMALAT1-WTAP axis: a novel layer of EMT regulation in hypoxic triple-negative breast cancer. *Cell Death Discov.* (2024) 10(1):276. doi: 10.1038/s41420-024-02058-4
- 130. Li Z-X, Zheng Z-Q, Yang P-Y, Lin L, Zhou G-Q, Lv J-W, et al. WTAP-mediated m6A modification of lncRNA DIAPH1-AS1 enhances its stability to facilitate nasopharyngeal carcinoma growth and metastasis. Cell Death Diff. (2022) 29:1137–51. doi: 10.1038/s41418-021-00905-w
- 131. Zhou Z, Chen X, Wang H, Ding L, Wang M, Li G, et al. WTAP-dependent N6-methyladenosine methylation of lncRNA TEX41 promotes renal cell carcinoma progression. *Sci Rep.* (2024) 14(1):24742. doi: 10.1038/s41598-024-76326-9
- 132. Wang F, Zhang J, Lin X, Yang L, Zhou Q, Mi X, et al. METTL16 promotes translation and lung tumorigenesis by sequestering cytoplasmic eIF4E2. *Cell Rep.* (2023) 42(3):112150. doi: 10.1016/j.celrep.2023.112150
- 133. Ye F, Wu J, Zhang F. METTL16 epigenetically enhances GPX4 expression via m6A modification to promote breast cancer progression by inhibiting ferroptosis. *Biochem Biophys Res Commun.* (2023) 638:1–6. doi: 10.1016/j.bbrc.2022.10.065

- 134. Xue M, Dong L, Zhang H, Li Y, Qiu K, Zhao Z, et al. METTL16 promotes liver cancer stem cell self-renewal via controlling ribosome biogenesis and mRNA translation. *J Hematol Oncol.* (2024) 17(1):7. doi: 10.1186/s13045-024-01526-9
- 135. Wang J, Xiu M, Wang J, Gao Y, Li Y. METTL16-SENP3-LTF axis confers ferroptosis resistance and facilitates tumorigenesis in hepatocellular carcinoma. *J Hematol Oncol.* (2024) 17(1):78. doi: 10.1186/s13045-024-01599-6
- 136. Wu J, Ouyang P, Huang R, Cui Y, Yang Z, Xu W, et al. METTL16 Promotes Stability of SYNPO2L mRNA and leading to Cancer Cell Lung Metastasis by Secretion of COL10A1 and attract the Cancer-Associated Fibroblasts. *Int J Biol Sci.* (2024) 20:4128–45. doi: 10.7150/iibs.95375
- 137. Xie Y-X, Wang L, Zhou Z-H, Liu W-J, Wang W, Yang J-H, et al. m6A RNA methyltransferase METTL16 induces Cr(VI) carcinogenesis and lung cancer development through glutamine biosynthesis and GLUL expression. *J Hazardous Mater.* (2024) 480:136093. doi: 10.1016/j.jhazmat.2024.136093
- 138. Cheng J, Xu Z, Tan W, He J, Pan B, Zhang Y, et al. METTL16 promotes osteosarcoma progression by downregulating VPS33B in an m6A-dependent manner. *J Cell Physiol.* (2023) 239(3):e31068. doi: 10.1002/jcp.31068
- 139. Wei W, Zhang Z-Y, Shi B, Cai Y, Zhang H-S, Sun C-L, et al. METTL16 promotes glycolytic metabolism reprogramming and colorectal cancer progression. *J Exp Clin Cancer Res.* (2023) 42(1):151. doi: 10.1186/s13046-023-02732-y
- 140. Han L, Dong L, Leung K, Zhao Z, Li Y, Gao L, et al. METTL16 drives leukemogenesis and leukemia stem cell self-renewal by reprogramming BCAA metabolism. *Cell Stem Cell*. (2023) 30:52–68.e13. doi: 10.1016/j.stem.2022.12.006
- 141. Liu N, Zhang J, Chen W, Ma W, Wu T. The RNA methyltransferase METTL16 enhances cholangiocarcinoma growth through PRDM15-mediated FGFR4 expression. *J Exp Clin Cancer Res.* (2023) 42(1):263. doi: 10.1186/s13046-023-02844-5
- 142. Li Q, Wang Y, Meng X, Wang W, Duan F, Chen S, et al. METTL16 inhibits papillary thyroid cancer tumorigenicity through m6A/YTHDC2/SCD1-regulated lipid metabolism. *Cell Mol Life Sci.* (2024) 81(1):81. doi: 10.1007/s00018-024-05146-x
- 143. Yu H, Zhuang J, Zhou Z, Song Q, Lv J, Yang X, et al. METTL16 suppressed the proliferation and cisplatin-chemoresistance of bladder cancer by degrading PMEPA1 mRNA in a m6A manner through autophagy pathway. *Int J Biol Sci.* (2024) 20:1471–91. doi: 10.7150/ijbs.86719
- 144. Li C, Liu J, Lyu Y, Ling S, Luo Y, Lim YP. METTL16 Inhibits the Malignant Progression of Epithelial Ovarian Cancer through the lncRNA MALAT1/β-Catenin Axis. *Analytical Cell Pathol.* (2023) 2023:9952234. doi: 10.1155/2023/9952234
- 145. Park SH, Ju J-S, Woo H, Yun HJ, Lee SB, Kim S-H, et al. The m6A writer RBM15 drives the growth of triple-negative breast cancer cells through the stimulation of serine and glycine metabolism. *Exp Mol Med.* (2024) 56:1373–87. doi: 10.1038/s12276-024-01235-w
- 146. Wang X, Tian L, Li Y, Wang J, Yan B, Yang L, et al. RBM15 facilitates laryngeal squamous cell carcinoma progression by regulating TMBIM6 stability through IGF2BP3 dependent. *J Exp Clin Cancer Res.* (2021) 40(1):80. doi: 10.1186/s13046-021-01871-4
- 147. Zhang H, Li Y, Zhou Y, Xu Q, Liao B, Qiu X, et al. RNA methylase RBM15 facilitates Malignant progression of colorectal cancer through regulating E2F2 in an m6A modification-dependent manner. *J Biochem Mol Toxicol.* (2024) 38(11):e70014. doi: 10.1002/ibt.70014
- 148. Ruixue Wang WT. RBM15-mediated N6-methyl adenosine (m6A) modification of EZH2 drives the epithelial-mesenchymal transition of cervical cancer. Crit Rev Eukaryot Gene Expr. (2024) 34:15–29. doi: 10.1615/CritRevEukaryotGeneExpr.2024052205
- 149. Quan Y, Zhou M, Li J, Yang Y, Guo J, Tang T, et al. The m6A methyltransferase RBM15 affects tumor cell stemness and progression of cervical cancer by regulating the stability of lncRNA HEIH. *Exp Cell Res.* (2024) 436(2):113924. doi: 10.1016/j.yexcr.2024.113924
- 150. Wang M, Qin Y, Ai X, Liu X. RBM15-dependent m6A modification mediates progression of non-small cell lung cancer cells. *Mol Med.* (2024) 30(1):267. doi: 10.1186/s10020-024-01018-z
- 151. Shi S, Wang C, Cai Q, Yang R, Peng M, Liang H, et al. RBM15 drives the progression of lung adenocarcinoma by regulating N6-methyladenosine-mediated LDHA mRNA stability. *Life Sci.* (2024) 358:123146. doi: 10.1016/j.lfs.2024.123146
- 152. Feng J, Li Y, He F, Zhang F. RBM15 silencing promotes ferroptosis by regulating the TGF-β/Smad2 pathway in lung cancer. *Environ Toxicol.* (2023) 38:950–61. doi: 10.1002/tox.23741
- 153. Song Y, Wu Q. RBM15 m6A modification-mediated OTUB2 upregulation promotes cervical cancer progression via the AKT/mTOR signaling. *Environ Toxicol.* (2023) 38:2155–64. doi: 10.1002/tox.23852
- 154. Zhang C, Gu L, Xiao J, Jin F. Knockdown of RBM15 inhibits tumor progression and the JAK-STAT signaling pathway in cervical cancer. *BMC Cancer*. (2023) 23 (1):684. doi: 10.1186/s12885-023-11163-z
- 155. Yuan J, Guan W, Li X, Wang F, Liu H, Xu G. RBM15–mediating MDR1 mRNA m6A methylation regulated by the TGF– $\beta$  signaling pathway in paclitaxel–resistant ovarian cancer. Int J Oncol. (2023) 63(4):112. doi: 10.3892/ijo.2023.5560
- 156. Zhang Z, Mei Y, Hou M. Knockdown RBM15 inhibits colorectal cancer cell proliferation and metastasis via N6-methyladenosine (m6A) modification of myD88 mRNA. *Cancer Biother Radiopharmaceut*. (2022) 37:976–86. doi: 10.1089/cbr.2021.0226

- 157. Zeng X, Chen K, Li L, Tian J, Ruan W, Hu Z, et al. Epigenetic activation of RBM15 promotes clear cell renal cell carcinoma growth, metastasis and macrophage infiltration by regulating the m6A modification of CXCL11. *Free Radical Biol Med.* (2022) 184:135–47. doi: 10.1016/j.freeradbiomed.2022.03.031
- 158. Wang W, He Y, Yao L-C, Yuan Y, Lu C, Xiong L-K, et al. Identification of m6A modification patterns and RBM15 mediated macrophage phagocytosis in pancreatic cancer: An integrative analysis. *Biochim Biophys Acta (BBA) Mol Basis Dis.* (2024) 1870 (7):167304. doi: 10.1016/j.bbadis.2024.167304
- 159. Wu L, Zhou Y, Fu J. KIAA1429 promotes nasopharyngeal carcinoma progression by mediating m6A modification of PTGS2. *Crit ReviewsTM Immunol.* (2023) 43:15–27. doi: 10.1615/CritRevImmunol.2023050249
- 160. Guo W, Wang T, Huai Q, Guo L, Wang X, He J. KIAA1429 regulates lung adenocarcinoma proliferation and metastasis through the PI3K/AKT pathway by modulating ARHGAP30 expression. *Thorac Cancer*. (2024) 15:1397–409. doi: 10.1111/1759-7714.15327
- 161. Xu D, Wang Z, Li F. KIAA1429 Induces m6A Modification of LINC01106 to Enhance the Malignancy of Lung Adenocarcinoma Cells via the JAK/STAT3 Pathway. *Crit ReviewsTM Immunol.* (2024) 44:49–61. doi: 10.1615/CritRevImmunol.2024052728
- 162. Zhang C, Sun Q, Zhang X, Qin N, Pu Z, Gu Y, et al. Gene amplification-driven RNA methyltransferase KIAA1429 promotes tumorigenesis by regulating BTG2 via m6A-YTHDF2-dependent in lung adenocarcinoma. *Cancer Commun.* (2022) 42:609–26. doi: 10.1002/cac2.12325
- 163. Zhao W, Xie Y. KIAA1429 promotes the progression of lung adenocarcinoma by regulating the m6A level of MUC3A. *Pathol Res Practice*. (2021) 217:153284. doi: 10.1016/j.prp.2020.153284
- 164. Ren M, Pan H, Zhou X, Yu M, Ji F. KIAA1429 promotes gastric cancer progression by destabilizing RASD1 mRNA in an m6A-YTHDF2-dependent manner. *J Trans Med.* (2024) 22(1):584. doi: 10.1186/s12967-024-05375-5
- 165. Tang B, Li M, Xu Y, Li X. N6-methyladenosine (m6A) writer KIAA1429 accelerates gastric cancer oxaliplatin chemoresistance by targeting FOXM1. *J Cancer Res Clin Oncol.* (2022) 149:5037–45. doi: 10.1007/s00432-022-04426-y
- 166. Yang D, Chang S, Li F, Ma M, Yang J, Lv X, et al. m6A transferase KIAA1429-stabilized LINC00958 accelerates gastric cancer aerobic glycolysis through targeting GLUT1. *IUBMB Life*. (2021) 73:1325–33. doi: 10.1002/iub.2545
- 167. Shan M, Liu D, Sun L, Yang M, He M, Zhang Y, et al. KIAA1429 facilitates metastasis via m6A-YTHDC1-dependent RND3 down-regulation in hepatocellular carcinoma cells. *Cancer Lett.* (2024) 584:216598. doi: 10.1016/j.canlet.2023.216598
- 168. Liu D, Shan M, Zeng R, He M, Dai X, Lu L, et al. Inhibition of KIAA1429/HK1 axis enhances the sensitivity of liver cancer cells to sorafenib by regulating the Warburg effect. *Biochem Pharmacol.* (2024) 227:116419. doi: 10.1016/j.bcp.2024.116419
- 169. Lu D, Chen A. lncRNA POU6F2-AS1 regulated by KIAA1429 contributes to colorectal cancer progression in an m6A modification manner. *Mol Biotechnol.* (2023) 67:115–22. doi: 10.1007/s12033-023-00986-7
- 170. Ma L, Lin Y, Sun S-W, Xu J, Yu T, Chen W-L, et al. KIAA1429 is a potential prognostic marker in colorectal cancer by promoting the proliferation via downregulating WEE1 expression in an m6A-independent manner. *Oncogene*. (2021) 41:692–703. doi: 10.1038/s41388-021-02066-z
- 171. Ouyang C, Xu G, Xie J, Xie Y, Zhou Y. Silencing of KIAA1429, a N6-methyladenine methyltransferase, inhibits the progression of colon adenocarcinoma via blocking the hypoxia-inducible factor 1 signalling pathway. *J Biochem Mol Toxicol.* (2024) 38(9):e23829. doi: 10.1002/jbt.23829
- 172. Gan L, Zhao S, Gao Y, Qi Y, Su M, Wang A, et al. N6-methyladenosine methyltransferase KIAA1429 promoted ovarian cancer aerobic glycolysis and progression through enhancing ENO1 expression. *Biol Direct.* (2023) 18(1):64. doi: 10.1186/s13062-023-00420-7
- 173. Wu Y, Luo Y, Yao X, Shi X, Xu Z, Re J, et al. KIAA1429 increases FOXM1 expression through YTHDF1-mediated m6A modification to promote aerobic glycolysis and tumorigenesis in multiple myeloma. *Cell Biol Toxicol.* (2024) 40(1):58. doi: 10.1007/s10565-024-09904-2
- 174. Zhang Y, Chen X, Chen H, Zhang Y. ZC3H13 enhances the Malignancy of cervical cancer by regulating m6A modification of CKAP2.  $Crit\ Reviews^{TM}\ Immunol.$  (2023) 43:1–13. doi: 10.1615/CritRevImmunol.2023049342
- 175. Wei Q, Yang Y, Li C, Wang H. ZC3H13-induced the m6A modification of hsa\_circ\_0081723 promotes cervical cancer progression via AMPK/p53 pathway. *J Obstetr Gynaecol Res.* (2024) 50:2286–98. doi: 10.1111/jog.16140
- 176. Huang L, Chen G, He J, Wang P. ZC3H13 reduced DUOX1-mediated ferroptosis in laryngeal squamous cell carcinoma cells through m6A-dependent modification. *Tissue Cell.* (2023) 84:102187. doi: 10.1016/j.tice.2023.102187
- 177. Lin X, Wang F, Chen J, Liu J, Lin Y-B, Li L, et al. N6-methyladenosine modification of CENPK mRNA by ZC3H13 promotes cervical cancer stemness and chemoresistance. *Military Med Res.* (2022) 9(1):19. doi: 10.1186/s40779-022-00378-z
- 178. Wu S, He G, Liu S, Cao Y, Geng C, Pan H. Identification and validation of the N6-methyladenosine RNA methylation regulator ZC3H13 as a novel prognostic marker and potential target for hepatocellular carcinoma. *Int J Med Sci.* (2022) 19:618–30. doi: 10.7150/ijms.69645
- 179. Xie R, Chen W, Lv Y, Xu D, Huang D, Zhou T, et al. Overexpressed ZC3H13 suppresses papillary thyroid carcinoma growth through m6A modification-mediated

- IQGAP1 degradation. J Formosan Med Assoc. (2023) 122:738–46. doi: 10.1016/j.ifma.2022.12.019
- 180. Liu B, Chen M, Liang Y, Mei Z, Sun W, Gao W, et al. ZC3H13 promotes autophagy in bladder cancer through m6A methylation modification of PJA2 and ubiquitination of KSR1. *Hum Cell*. (2024) 38(1):23. doi: 10.1007/s13577-024-01155-x
- 181. Zhang C, Li Y, Wang Y, Chen C. VIRMA Facilitates Triple-Negative Breast Cancer Progression via Increasing m6A-Dependent KIF15 Expression. *Discov Med.* (2023) 35(178):787–95. doi: 10.24976/Discov.Med.202335178.73
- 182. Zheng Z-Q, Huang Z-H, Liang Y-L, Zheng W-H, Xu C, Li Z-X, et al. VIRMA promotes nasopharyngeal carcinoma, tumorigenesis, and metastasis by upregulation of E2F7 in an m6A-dependent manner. *J Biol Chem.* (2023) 299(5):104677. doi: 10.1016/i.jbc.2023.104677
- 183. Xu H, Lin X, Li Z, He X, Li Y, Qiu L, et al. VIRMA facilitates intrahepatic cholangiocarcinoma progression through epigenetic augmentation of TMED2 and PARD3B mRNA stabilization. *J Gastroenterol.* (2023) 58:925–44. doi: 10.1007/s00535-023-02015-5
- 184. Zhu C, Cheng Y, Yu Y, Zhang Y, Ren G. VIRMA promotes the progression of head and neck squamous cell carcinoma by regulating *UBR5* mRNA and m6A levels. *Biomolecul Biomed.* (2024) 24:1244–57. doi: 10.17305/bb.2024.10358
- 185. Xu Y, Chen Y, Yao Y, Xie H, Lu G, Du C, et al. VIRMA contributes to non-small cell lung cancer progression via N6-methyladenosine-dependent DAPK3 post-transcriptional modification. *Cancer Lett.* (2021) 522:142–54. doi: 10.1016/j.canlet.2021.08.027
- 186. Zhang X, An K, Ge X, Sun Y, Wei J, Ren W, et al. NSUN2/YBX1 promotes the progression of breast cancer by enhancing HGH1 mRNA stability through m5C methylation. *Breast Cancer Res.* (2024) 26(1):94. doi: 10.1186/s13058-024-01847-0
- 187. Du X, Cheng C, Yang Y, Fan B, Wang P, Xia H, et al. NSUN2 promotes lung adenocarcinoma progression through stabilizing PIK3R2 mRNA in an m5C-dependent manner. *Mol Carcinogen*. (2024) 63:962–76. doi: 10.1002/mc.23701
- 188. Xing H, Gu X, Liu Y, Xu L, He Y, Xue C. NSUN2 regulates Wnt signaling pathway depending on the m5C RNA modification to promote the progression of hepatocellular carcinoma. *Oncogene*. (2024) 43:3469–82. doi: 10.1038/s41388-024-03184-0
- 189. Niu K, Chen Z, Li M, Ma G, Deng Y, Zhang J, et al. NSUN2 lactylation drives cancer cell resistance to ferroptosis through enhancing GCLC-dependent glutathione synthesis. *Redox Biol.* (2025) 79:103479. doi: 10.1016/j.redox.2024.103479
- 190. Chen B, Deng Y, Hong Y, Fan L, Zhai X, Hu H, et al. Metabolic Recoding of NSUN2-Mediated m5C Modification Promotes the Progression of Colorectal Cancer via the NSUN2/YBX1/m5C-ENO1 Positive Feedback Loop. *Adv Sci.* (2024) 11(28): e2309840. doi: 10.1002/advs.202309840
- 191. Zou S, Huang Y, Yang Z, Zhang J, Meng M, Zhang Y, et al. NSUN2 promotes colorectal cancer progression by enhancing SKIL mRNA stabilization. *Clin Trans Med.* (2024) 14(3):e1621. doi: 10.1002/ctm2.1621
- 192. Yang M, Wei R, Zhang S, Hu S, Liang X, Yang Z, et al. NSUN2 promotes osteosarcoma progression by enhancing the stability of FABP5 mRNA via m5C methylation. *Cell Death Dis.* (2023) 14(2):125. doi: 10.1038/s41419-023-05646-x
- 193. Jin S, Li J, Shen Y, Wu Y, Zhang Z, Ma H. RNA 5-Methylcytosine Regulator NSUN3 promotes tumor progression through regulating immune infiltration in head and neck squamous cell carcinoma. *Oral Dis.* (2022) 30:313–28. doi: 10.1111/odi.14357
- 194. Tang Z, Zhang N, Chen S, Fang J, Tang X, Lou Y, et al. Bipyridine derivatives as NOP2/sun RNA methyltransferase 3 inhibitors for the treatment of colorectal cancer. *J Med Chem.* (2024) 67:13446–73. doi: 10.1021/acs.jmedchem.4c01323
- 195. Wu J, Zhao Q, Chen S, Xu H, Zhang R, Cai D, et al. NSUN4-mediated m5C modification of circERI3 promotes lung cancer development by altering mitochondrial energy metabolism. *Cancer Lett.* (2024) 11:217266. doi: 10.1016/j.canlet.2024.217266
- 196. Zhao Z, Zhou Y, Lv P, Zhou T, Liu H, Xie Y, et al. NSUN4 mediated RNA 5-methylcytosine promotes the Malignant progression of glioma through improving the CDC42 mRNA stabilization. *Cancer Lett.* (2024) 597:217059. doi: 10.1016/j.canlet.2024.217059
- 197. Cui M, Qu F, Wang L, Liu X, Yu J, Tang Z, et al. m5C RNA methyltransferaserelated gene NSUN4 stimulates Malignant progression of hepatocellular carcinoma and can be a prognostic marker. *Cancer Biomarkers*. (2022) 33:389–400. doi: 10.3233/CBM-210154
- 198. Gu X, Li P, Gao X, Ru Y, Xue C, Zhang S, et al. RNA 5-methylcytosine writer NSUN5 promotes hepatocellular carcinoma cell proliferation via a ZBED3-dependent mechanism. *Oncogene*. (2024) 43:624–35. doi: 10.1038/s41388-023-02931-z
- 199. Han H, Zhang C, Shi W, Wang J, Zhao W, Du Y, et al. NSUN5 facilitates hepatocellular carcinoma progression by increasing SMAD3 expression. *Adv Sci.* (2024) 12(2):e2404083. doi: 10.1002/advs.202404083
- 200. Liu S, Liu Y, Zhou Y, Xia G, Liu H, Zeng Y, et al. NSUN5 promotes tumorigenic phenotypes through the WNT signaling pathway and immunosuppression of CD8+ T cells in gastric cancer. Cell Signal. (2024) 124:111475. doi: 10.1016/j.cellsig.2024.111475
- 201. Cui Y, Hu Z, Zhang C. RNA methyltransferase NSUN5 promotes esophageal cancer via 5-methylcytosine modification of METTL1. *Mol Carcinogen.* (2024) 64:399–409. doi: 10.1002/mc.23857
- 202. Zhou J, Kong YS, Vincent KM, Dieters-Castator D, Bukhari AB, Glubrecht D, et al. RNA cytosine methyltransferase NSUN5 promotes protein synthesis and

- tumorigenic phenotypes in glioblastoma. *Mol Oncol.* (2023) 17:1763-83. doi: 10.1002/1878-0261.13434
- 203. Lu Z LB, Kong D, Zhou X, Pei D, Liu D. NSUN6 regulates NM23-H1 expression in an m5C manner to affect epithelial-mesenchymal transition in lung cancer. *Med Princ Pract.* (2024) 33:56–65. doi: 10.1159/000535479
- 204. Cui Y, Lv P, Zhang C. NSUN6 mediates 5-methylcytosine modification of METTL3 and promotes colon adenocarcinoma progression. *J Biochem Mol Toxicol.* (2024) 38(6):e23749. doi: 10.1002/jbt.23749
- 205. Yu M, Ni M, Xu F, Liu C, Chen L, Li J, et al. NSUN6-mediated 5-methylcytosine modification of NDRG1 mRNA promotes radioresistance in cervical cancer. *Mol Cancer*, (2024) 23(1):139. doi: 10.1186/s12943-024-02055-2
- 206. Lai J, Chen L, Li Q, Zhao G, Li X, Guo D, et al. tRNA methyltransferase DNMT2 promotes hepatocellular carcinoma progression and enhances Bortezomib resistance through inhibiting TNFSF10. *Cell Signal.* (2025) 127:111533. doi: 10.1016/j.cellsig.2024.111533
- 207. Wang Q, Zhang Q, Huang Y, Zhang J. m1A regulator TRMT10C predicts poorer survival and contributes to Malignant behavior in gynecological cancers. *DNA Cell Biol.* (2020) 39:1767–78. doi: 10.1089/dna.2020.5624
- 208. Chen J, Li K, Chen J, Wang X, Ling R, Cheng M, et al. Aberrant translation regulated by METTL1/WDR4-mediated tRNA N7-methylguanosine modification drives head and neck squamous cell carcinoma progression. *Cancer Commun.* (2022) 42:223–44. doi: 10.1002/cac2.12273
- 209. Du D, Zhou M, Ju C, Yin J, Wang C, Xu X, et al. METTL1-mediated tRNA m7G methylation and translational dysfunction restricts breast cancer tumorigenesis by fueling cell cycle blockade. *J Exp Clin Cancer Res.* (2024) 43(1):154. doi: 10.1186/s13046-024-03076-x
- 210. Peng W, Fu J, Zhou L, Duan H. METTL1/FOXM1 promotes lung adenocarcinoma progression and gefitinib resistance by inhibiting PTPN13 expression. *Cancer Med.* (2024) 13(13):e7420. doi: 10.1002/cam4.7420
- 211. Wang F, Yang C, Zheng F, Yan Y, Li G, Feng Y, et al. METTL1 mediates PKM m7G modification to regulate CD155 expression and promote immune evasion in colorectal cancer. *J Trans Med.* (2024) 22(1):1161. doi: 10.1186/s12967-023-04783-3
- 212. Dong R, Wang C, Tang B, Cheng Y, Peng X, Yang X, et al. WDR4 promotes HCC pathogenesis through N7-methylguanosine by regulating and interacting with METTL1. *Cell Signal.* (2024) 118:111145. doi: 10.1016/j.cellsig.2024.111145
- 213. Xia P, Zhang H, Xu K, Jiang X, Gao M, Wang G, et al. MYC-targeted WDR4 promotes proliferation, metastasis, and sorafenib resistance by inducing CCNB1 translation in hepatocellular carcinoma. *Cell Death Dis.* (2021) 12(7):691. doi: 10.1038/s41419-021-03973-5
- 214. Liu X, Su K, Sun X, Jiang Y, Wang L, Hu C, et al. Sec62 promotes stemness and chemoresistance of human colorectal cancer through activating Wnt/ $\beta$ -catenin pathway. *J Exp Clin Cancer Res.* (2021) 40(1):132. doi: 10.1186/s13046-021-01934-6
- 215. Lin Z, Niu Y, Wan A, Chen D, Liang H, Chen X, et al. RNA m6A methylation regulates sorafenib resistance in liver cancer through FOXO 3-mediated autophagy. *EMBO J.* (2020) 39(12):e103181. doi: 10.15252/embj.2019103181
- 216. Ye X, Wang L-P, Han C, Hu H, Ni C-M, Qiao G-L, et al. Increased m6A modification of lncRNA DBH-AS1 suppresses pancreatic cancer growth and gemcitabine resistance via the miR-3163/USP44 axis. *Ann Trans Med.* (2022) 10:304–. doi: 10.21037/atm-22-556
- 217. Zhou T, Li S, Xiang D, Liu J, Sun W, Cui X, et al. m6A RNA methylation-mediated HNF3γ reduction renders hepatocellular carcinoma dedifferentiation and sorafenib resistance. Signal Transduct Targeted Ther. (2020) 5(1):296. doi: 10.1038/s41392-020-00299-0
- 218. Zhang C, Ou S, Zhou Y, Liu P, Zhang P, Li Z, et al. m6A methyltransferase METTL14-mediated upregulation of cytidine deaminase promoting gemcitabine resistance in pancreatic cancer. *Front Oncol.* (2021) 11:696371. doi: 10.3389/fonc.2021.696371
- 219. Wang Y, Wei J, Feng L, Li O, Huang L, Zhou S, et al. Aberrant m5C hypermethylation mediates intrinsic resistance to gefitinib through NSUN2/YBX1/QSOX1 axis in EGFR-mutant non-small-cell lung cancer. *Mol Cancer*. (2023) 22(1):81. doi: 10.1186/s12943-023-01780-4
- 220. Yu H, Liu J, Bu X, Ma Z, Yao Y, Li J, et al. Targeting METTL3 reprograms the tumor microenvironment to improve cancer immunotherapy. *Cell Chem Biol.* (2024) 31:776–91.e7. doi: 10.1016/j.chembiol.2023.09.001
- 221. Pan Y, Chen H, Zhang X, Liu W, Ding Y, Huang D, et al. METTL3 drives NAFLD-related hepatocellular carcinoma and is a therapeutic target for boosting immunotherapy. *Cell Rep Med.* (2023) 4(8):101144. doi: 10.1016/j.xcrm.2023.101144
- 222. Wang L, Hui H, Agrawal K, Kang Y, Li N, Tang R, et al. m6A RNA methyltransferases METTL3/14 regulate immune responses to anti-PD-1 therapy. *EMBO J.* (2020) 39(20):e104514. doi: 10.15252/embj.2020104514
- 223. Ning J, Hou X, Hao J, Zhang W, Shi Y, Huang Y, et al. METTL3 inhibition induced by M2 macrophage-derived extracellular vesicles drives anti-PD-1 therapy resistance via M6A-CD70-mediated immune suppression in thyroid cancer. *Cell Death Diff.* (2023) 30:2265–79. doi: 10.1038/s41418-023-01217-x
- 224. Yin H, Zhang X, Yang P, Zhang X, Peng Y, Li D, et al. RNA m6A methylation orchestrates cancer growth and metastasis via macrophage reprogramming. *Nat Commun.* (2021) 12(1):1394. doi: 10.1038/s41467-021-21514-8

- 225. Zheng H, Zheng W-J, Wang Z-G, Tao Y-P, Huang Z-P, Yang L, et al. Decreased expression of programmed death ligand-L1 by seven in absentia homolog 2 in cholangiocarcinoma enhances T-cell-mediated antitumor activity. *Front Immunol.* (2022) 13:845193. doi: 10.3389/fimmu.2022.845193
- 226. Song H, Song J, Cheng M, Zheng M, Wang T, Tian S, et al. METTL3-mediated m6A RNA methylation promotes the anti-tumour immunity of natural killer cells. *Nat Commun.* (2021) 12(1):5522. doi: 10.1038/s41467-021-25803-0
- 227. Liu W-W, Zheng S-Q, Li T, Fei Y-F, Wang C, Zhang S, et al. RNA modifications in cellular metabolism: implications for metabolism-targeted therapy and immunotherapy. *Signal Transduct Targeted Ther.* (2024) 9(1):70. doi: 10.1038/s41392-024-01777-5
- 228. Chen X, Yuan Y, Zhou F, Li L, Pu J, Jiang X. m6A RNA methylation: a pivotal regulator of tumor immunity and a promising target for cancer immunotherapy. *J Trans Med.* (2025) 23(1):245. doi: 10.1186/s12967-025-06221-y
- 229. Li Z, Feng Y, Han H, Jiang X, Chen W, Ma X, et al. A stapled peptide inhibitor targeting the binding interface of N6-adenosine-methyltransferase subunits METTL3 and METTL14 for cancer therapy. *Angewandte Chem Int Ed.* (2024) 63(24):e202402611. doi: 10.1002/anie.202402611
- 230. Yankova E, Blackaby W, Albertella M, Rak J, De Braekeleer E, Tsagkogeorga G, et al. Small-molecule inhibition of METTL3 as a strategy against myeloid leukaemia. *Nature.* (2021) 593:597–601. doi: 10.1038/s41586-021-03536-w
- 231. Chen X, Wang M, Wang H, Yang J, Li X, Zhang R, et al. METTL3 inhibitor suppresses the progression of prostate cancer via IGFBP3/AKT pathway and synergizes

- with PARP inhibitor. *Biomed Pharmacother*. (2024) 179:117366. doi: 10.1016/j.biopha.2024.117366
- 232. Pomaville M, Chennakesavalu M, Wang P, Jiang Z, Sun H-L, Ren P, et al. Small-molecule inhibition of the METTL3/METTL14 complex suppresses neuroblastoma tumor growth and promotes differentiation. *Cell Rep.* (2024) 43 (5):114165. doi: 10.1016/j.celrep.2024.114165
- 233. Liu L, Zhao T, Zheng S, Tang D, Han H, Yang C, et al. METTL3 inhibitor STM2457 impairs tumor progression and enhances sensitivity to anlotinib in OSCC. *Oral Dis.* (2024) 30:4243–54. doi: 10.1111/odi.14864
- 234. Moroz-Omori EV, Huang D, Kumar Bedi R, Cheriyamkunnel SJ, Bochenkova E, Dolbois A, et al. METTL3 inhibitors for epitranscriptomic modulation of cellular processes. *ChemMedChem.* (2021) 16:3035–43. doi: 10.1002/cmdc.202100291
- 235. Dolbois A, Bedi RK, Bochenkova E, Müller A, Moroz-Omori EV, Huang D, et al. 1,4,9-triazaspiro[5.5]undecan-2-one derivatives as potent and selective METTL3 inhibitors. *J Med Chem.* (2021) 64:12738–60. doi: 10.1021/acs.jmedchem.1c00773
- 236. Chen T, Xu Z-G, Luo J, Manne RK, Wang Z, Hsu C-C, et al. NSUN2 is a glucose sensor suppressing cGAS/STING to maintain tumorigenesis and immunotherapy resistance. *Cell Metab.* (2023) 35:1782–98.e8. doi: 10.1016/j.cmet.2023.07.009
- 237. Xue DD, Zhang Y, Shen AR, Tang TT, Liu BC, Lv LL. mRNA therapy: A new frontier in regenerative medicine. *Interdiscip Med.* (2025) 3:1–17. doi: 10.1002/INMD.20240054