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Notch signalling in T cells: bridging tumour immunity and intratumoral cellular crosstalk

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Background: Notch receptor–ligand interaction is ubiquitous and fundamental for coordinating cellular differentiation and determining cell fate for the development of various tissues and organs. Aberrant mutations in the Notch cascade result in various pathophysiological disorders, including cancer. Diverse aspects of carcinogenesis regulated by Notch include the shaping of anti-tumour T-cell immunity through antigen-presenting cell (APC)—T cell interaction and effector functions.

Chief content: Notch depends on juxtacrine and paracrine signalling to influence intercellular communications in the tumour microenvironment. Several preclinical and clinical studies have revealed Notch as a bi-effector molecule, which has a differential effect depending on the immune contexture of the tumour microenvironment. The Notch cascade serves as an effective therapeutic target in preventing off-target cell death and promoting tumour-specific T-cell priming.

Conclusion: This review revolves around Notch crosstalk with respect to the interaction between T-cell populations and other intratumoral cellular components, including professional antigen-presenting cells like dendritic cells, macrophages, B cells, immunosuppressive myeloid-derived suppressor cells, and cancer stem cells. It also summarizes the impact of targeting Notch signalling within intratumoral T cells in combination with traditional oncotherapies.

KEYWORDS

notch, T cells, macrophages, dendritic cells, myeloid derived suppressor cells, B cells, targeted-therapy

Introduction

The immune modulatory functionality of Notch signalling depends on both the involved cell and environmental cues; it becomes further multifaceted within the tumour microenvironment (TME). Tumour intrinsic crosstalk between immune cells, stromal cells, and malignant cells is regulated via Notch-induced reciprocal signalling (juxtacrine and paracrine) networks. Tumour cells often hijack these pathways and reroute them for their own benefit. The Notch signalling pathway plays both oncogenic and tumour-suppressor roles depending upon the type of malignancy. Notch is considered to be an oncogene in multiple malignancies of lymphoid origin (splenic marginal zone lymphoma, T-cell acute lymphoblastic leukaemia, and B-cell chronic lymphocytic leukaemia) (1-3). On the contrary, it is denoted as a tumour suppressor in myeloid malignancies (4, 5). Therefore, to ascertain and design anti-Notch therapeutics, an extensive look at the TME is required. Thus, the aim of this review was to illustrate this labyrinthine Notch signalling network within the TME with special emphasis on T-cell fate, their regulatory mechanisms, and therapeutic applications.

The Notch pathway: receptors, ligands, and downstream signalling

Almost a century ago, Thomas Hunt Morgan and colleagues first discovered the Notch protein in a strain of fruit flies (*Drosophila melanogaster*) where the haplo-insufficiency of the Notch locus resulted in notched wings [R-Morgan, 1917]. This evolutionarily conserved signalling molecule influences differentiation, morphogenesis, proliferation, apoptosis, and cell fate determination (6). A summary of the Notch receptor–ligand interaction controlling immune cell fate is shown in Table 1.

The Notch signalling is activated when any one of the four Notch receptors (Notch1-4) binds to their ligands, Jagged1/2 or Delta-like1-3. In humans, the genes of four Notch receptors are positioned on chromosomes 9, 1, 19, and 6 (7). In particular, Notch is a heterodimeric receptor comprising the Notch extracellular domain (NED), a transmembrane domain, and a Notch intracellular cytoplasmic domain (NICD). NICD harbours several conserved elements, such as protein-protein interaction, transactivation domains, and nuclear localization signals (NLSs) (8). The N-terminal region of Notch possesses a negative regulatory

TABLE 1 Notch signalling in the developmental fate of different immune cells.

Immune cells	Notch receptor	Notch ligand	Developmental stage modulated by the specific immune cell type	References
NK cells	Notch1 and 3	DLL1, Jagged1 and 2	Development of functional NK cells from hematopoietic progenitor cells.	(15–17)
	Notch1 and 2	DLL1 (decidual NK cells) and DLL4 (both peripheral blood and decidual NK cells)	Induce cytotoxicity of NK cells by upregulating IFN- γ secretion.	(18)
	Notch2	Jagged2	DC-mediated cytotoxicity of NK cells is increased.	(19)
	Notch1	DLL1 and 4	Development of NK cells from pro-B cells or common progenitor lymphoid cells.	(20)
Macrophages	Notch1	DLL1 and Jagged1	M0 macrophages are polarized into M1 macrophages with proinflammatory functionalities by inhibiting SIRPα.	(21–23)
	Notch1 and 3	DLL1 and 4	NF-κB-dependent proinflammatory gene expression is upregulated in TLR4-activated macrophages.	(24)
B cells	Notch1	DLL1 and Jagged1	Development of plasma cells occurs, and antibody production of the cells is also increased.	(25, 26)
	Notch2	DLL1	Upon BCR stimulation, follicular B cells are developed into marginal zone B cells.	(27, 28)
	Notch1	DLL1 and 4	Development of B cells from pro-B cells or common progenitor lymphoid cells is inhibited.	(20)
T cells	Notch3	DLL1	Naïve CD4 ⁺ cells developed into Th1 T helper cells.	(29)
	N/A	DLL1 and 4	DC-mediated signalling inhibited Th2 cell development.	(30)
	Notch1	DLL4	Synergistically with TGF- β , it induces Treg cell differentiation by inducing Foxp3 signalling.	(31, 32)

NK, natural killer cells; DC, dendritic cell; TGF-β, transforming growth factor beta; BCR, B cell receptor.

region (NRR) with 36 epidermal growth factor (EGF)-like repeats. The NRR contains 12 cysteine-rich repeats and a site that helps form heterodimers for S2 cleavage (9). Both Notch ligands and receptors are transmembrane proteins involved in cell-to-cell interaction. The Notch receptor interacts with an association region for the recombination signal binding protein for immunoglobulin kappa J region (RBPJ). The RBPJ association module has seven ankyrin repeats (ANKs) and helps in interaction with the nuclear transcription factors (10).

Proline/glutamic acid/serine/threonine-rich motifs (PEST domains) are located in the terminal parts of the intracellular region; these contain signals for degradation and hence are crucial for the stability of NICD (11). Mammalian Notch2–4 have structures resembling Notch1, differing solely in the number of EGF-like repeats, the extent of glycosylation of the EGF-like repeats, and the span of PEST domains. The expression level of Notch receptors on the cell membrane is regulated by endocytosis. During activation, the Notch receptors are first targeted by an ADAM-family metalloprotease (ADAM 10), which targets the NED. It is succeeded by a series of proteolytic cleavages and mediated by the γ -secretase complex and the translocation of NICD to the nucleus to regulate cellular gene expression (12). Once trafficked into the

nucleus, NICD heterodimerizes with RBPJ, a DNA-binding transcription factor, and gives rise to a short-lived nuclear transcription complex, which is supported by co-activators MAML belonging to the Mastermind-like family (MAML1-3) (13, 14).

This leads to the activation of downstream genes like hes (Hairy enhancer of split) and hey (Hairy related). In spite of similarities in the structure, the four Notch ligands have differential effects on various tissue types, which allows high plasticity in the regulation of gene expression (10). Cellular differentiation and communication are mostly regulated by DLL1. DLL4 has been noted to promote metastasis through VEGF secretion (33). JAG1 is reported to promote angiogenesis. However, JAG2 enhances cell proliferation (34, 35). The receptor–ligand interaction Notch is exhibited in Figure 1.

Notch signalling regulates T-cell fate

Studies pertaining to the immune system have proven that Notch signalling influences several steps of T-cell development in central and peripheral lymphoid organs. It is identified to have a

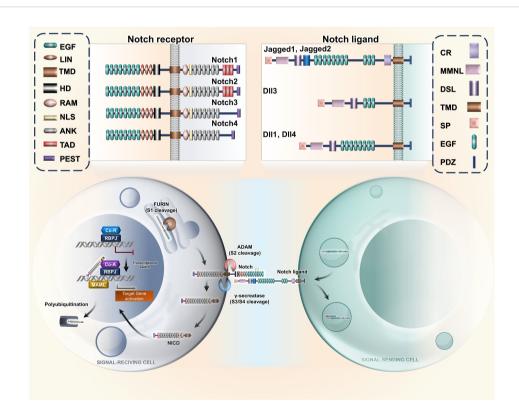


FIGURE 1

Notch receptor–ligand interaction. Representative illustration of interaction between the varied Notch ligands with their corresponding receptors, resulting in cleavage of intracellular domain due to activity of ADAM proteases, resulting in nuclear translocation of NICD. The acronyms or abbreviations have been added to the abbreviation list. EGF, epidermal growth factor; TMD, transmembrane domain; HD, heterodimerization domain; LIN, lineage defective; RAM, RBP-Jk-Associated Module; NLS, nuclear localization signal; ANK, ankyrin repeat; TAD, transactivation domain; PEST, proline glutamine serine threonine; DLL, Delta-like ligand; CR, cysteine-rich; MMNL, module at the N-terminus of Notch ligands; DSL, Delta/Serrate/LAG-2; SP, signal peptide; PDZ, protein-protein interaction module; MAML, Mastermind-like; CoA, coactivator; CoR, corepressor; RBPJ, recombination signal binding protein for immunoglobulin kappa J region; ADAM, A Disintegrin And Metalloprotease; NICD, Notch intracellular domain.

monumental role in T-cell development, differentiation, and maturation. Prior to T-cell receptor (TCR) ligation, Notch receptors are densely expressed in naïve T cells. Both thymopoiesis and T-cell maturation are spatially organized in the thymus (36). The lymphoid progenitors come into contact with overexpressed Notch ligands in the thymic cortex, and they face Notch-associated cues, which determine T-cell lineage commitment. Notch signalling continues till the pre-TCR checkpoint, beyond which it is downregulated (37). In doublepositive (CD4+CD8+) thymocytes, the expression of Notch decreases to avoid the probable intervention in positive and negative selection. TCR stimulation causes CD4+ T cells to differentiate into several subsets. Inversely, JAG1 or JAG2 ligands are considered more important for the Th2 subtype (38). After antigen exposure, CD8+ T cells differentiate into either short-lived effector cells (SLECs) or memory precursor effector cells (MPECs). Notch signalling regulates the differentiation of SLECs following dendritic cell (DC) immunization. The SLEC population dwindles in the absence of Notch signalling. However, Notch is dispensable in the MPEC differentiation process (39).

Tumorigenesis manifests as immune suppression and hinders intrathymic maturation or differentiation of T cells. It mediates a loss in the pool of CD4⁺CD8⁺ immature thymocytes and a steady increase in CD4⁻CD8⁻ progenitors of T cells. A study by Guha and colleagues (40) elaborated that the progression of malignancy inhibits the modulation of Lineage⁻Thy1.2⁺CD25⁺CD44⁺c-Kit low DN2b to Lineage⁻Thy1.2⁺CD25⁺CD44⁻c-Kit⁻DN3 during T-cell maturation. It was also reported that there was a downregulation of Notch1 and downstream targets like Ikaros, irf8, and pu.1, which direct the transition of DN2a towards dendritic cell lineage commitment. Corresponding Notch ligands like DLL1 or DLL4, on CD4⁺ T-cell activation, skew them to the Th1 fate. Moreover, it was also reported that the interaction of interleukin-10 with IL-10 receptor high DN2 thymocytes further drives DC lineage commitment by STAT3 phosphorylation (40).

Notch signalling regulates T-cell functionality

The activation of Notch1/2 in CD8⁺ T cells boosts anti-tumour response, increasing IFN-γ production and reducing tumour load (41). DLL1 (human homolog of Notch ligand) expression on DCs and bone marrow cells magnifies T-cell infiltration in tumours. It also increases IFN-γ production and mouse survivability and decreases tumour burden (42). Moreover, Notch-dependent increased functionality of antigen-specific CD8⁺ tumour-infiltrating lymphocytes (TILs) was also reported. Primarily, the clustered form of DLL1 (c-DLL1) binds and activates Notch (1–4), resulting in Notch-mediated/targeted gene expression, increasing the tumour infiltration of antigen-specific T cells and the regression of tumour (43). By inhibiting or knocking down Notch expression (mainly Notch2), it was found that the differentiation and promotion of the cytolytic function of murine T cells were prohibited both *in vitro* and *in vivo*. These observations specified that Notch2 is decisive for the

anti-tumour reaction of cytotoxic T lymphocyte (CTL) cells. The three complexes comprising phosphorylated CREB1, activated NICD, and transcriptional co-activator p300 enhance Notch transcription by binding to the promoter of the granzyme B gene, thus enhancing its transcription (44). However, many counteractive effects of Notch signalling in intratumoral T cells were also evident. In colorectal carcinoma, Notch cascades inhibit proinflammatory cytokine (IFN-γ, TNFα, IL-1β, IL-6, and IL-8)-dependent noncytolytic anti-tumour function of CD8+ T cells. Apart from increasing the effector immune response of CD8+ T cells, Notch signalling can also hasten T-cell exhaustion. The inhibition of Notch signalling led to a significant decrease in programmed cell death protein 1 (PD-1) expression of CD8⁺ T cells. The activation complex of Notch signalling was observed to accelerate PD-1 transcription in CD8⁺ T cells by binding to its promoter. In colorectal carcinoma, it was observed that the Notch expression elevated the PD-1 expression in CD8⁺ T cells (45). The multifaceted role of Notch signalling in Tcell activity is shown in Table 2.

Notch signalling in T-cell dysfunction

The development and growth of cancer generate dysfunctional or exhausted CD8+ T cells, which assist immunotolerance and correspond with tumour metastasis. This results in failure of functional peripheral blood CD8⁺ T cells, which is revealed by depleting the cytotoxic effects (for example, by perforin/granzyme pathway and Fas/FasL pathway). This involves an increase in the expression of inhibitory immune receptors or regulatory molecules like PD-1, cytotoxic T lymphocyte-associated protein-4 (CTLA-4), and T-cell immunoglobulin and mucin domain-containing-3 (TIM-3), thus creating a tolerogenic environment. The suppression of the Notch signalling pathway in CD8+ T cells from prostate cancer (PCa) patients results in the decreased expression of downstream signalling molecules Hes1 and Hes5. Hes1 maintains the proliferation of progenitor cells, and it is an essential effector of the Notch signalling pathway (46). Notch activation enhances the direct cell killing capability of CD8+ T cells, fostering the expression of cytotoxic molecules while diminishing inhibitory immune regulatory molecules.

It was also observed that due to the distinct patterns of gene activity observed in Notch1 and Notch2 within colorectal carcinoma samples, Notch signalling plays a role in the detriment of CD8⁺ T-cell functions during the development of colorectal carcinoma. The inhibition of Notch signalling reduces PD-1 expression on CD8⁺ T cells in patients with colorectal carcinoma; however, the frequency of CTLA-4⁺ cells within CD8⁺ T cells did not have a significant change in either peripheral blood (healthy individuals and patients) or colorectal carcinoma specimens (45). Notch signalling plays a significant role in T-cell senescence in cancer, influencing various aspects of T-cell function and fate. Dysregulated Notch signalling is often characterized by increased Notch receptor activation and has been linked to the induction of senescence-associated changes in T cells. Senescence-associated secretory phenotype (SASP) is illustrated by the presence of various proinflammatory cytokines, including IL-

TABLE 2 The diverse effects of Notch on T cells in different types of cancer.

Types of malignancy	Modulation of Notch expression	Effect on T cells	Reference
Lung cancer	Overexpression of Notch1 in cancer cells.	Increased glycolysis in cancer cells via Notch1/TAZ axis. It leads to excess lactate production, which inhibits the lactate release of T cells. Due to the intracellular lactate accumulation, T-cell proliferation and cytokine release are abrogated, resulting in reduced cytotoxic T-cell function.	(47)
	Multivalent DLL1-Notch1 interaction.	Increased anti-tumour T-cell response and enhanced efficacy of epidermal growth factor receptor targeted therapy.	(48)
Ovarian cancer	Reduced Notch expression in T cells via restriction of EZH2 by reducing aerobic glycolysis.	Reduced effector T-cell survival and cytokine release. Altogether, T cellmediated antitumour activity is attenuated.	(49)
EG7 thymoma	Notch signalling is stimulated.	Notch2 signalling directly regulates transcription of genes encoding the crucial CTL effector molecules granzyme B and perforin.	(44, 50)
Leukaemia	Increased expression of Notch ligand is observed in mouse model.	DLL4 shows a great capacity to promote the generation of T helper (Th)1 and Th17 CD4+ T cells.	(51)
Hepatocellular carcinoma	Blockade in nuclear translocation of the intracellular motif of Notch2 by overexpression of 1810011010Rik (Tcim)	The cytotoxic effect of CD8 ⁺ T cells was inhibited.	(52)

CTL, cytotoxic T lymphocyte.

1, IL-6, and IL-8. The activated Notch1 intracellular domain (N1ICD) and the conventional Notch1 target, HES1, showed a transient increase during the shift towards senescence. Nevertheless, as senescence reached its full extent, both the cleaved Notch1 intracellular domain and HES1 returned to levels close to their baseline. It was observed that there was a temporary increase in transforming growth factor beta (TGF- β) ligands during both Replicative-Induced Senescence (RIS) and DNA Damage-Induced

Senescence (DDIS). This trend shows the expression pattern of N1ICD. These findings suggest that there is a temporal correlation between Notch signalling and the concurrent induction of TGF- β and proinflammatory cytokines throughout the senescence process (53). Understanding the tangled interplay between Notch signalling and T-cell anergy, exhaustion, and senescence is crucial for unfolding the molecular mechanisms and the therapeutic targets for age-related immune dysfunction associated with aberrant T-cell responses. The impact of Notch signalling in T cell-mediated anti-tumour immunity is showcased in Figure 2.

Notch as a bi-effector molecule in the tumour niche

Several lines of evidence have suggested that Notch can also act as a tumour suppressor by shaping the tumour immune microenvironment. It modifies the immune milieu and stromal interactions. For example, it was demonstrated by Demehri et al. that Notch deletion alters cytokine secretion and interrupts epidermal homeostasis by promoting an inflammatory TME that advances tumour development (54–56). Parmigiani et al. reported that with Notch loss in brain tumour, an immune-suppressive environment was created that ultimately generated conditions favouring tumour progression (57). This highlights that Notch both functions as an oncogenic driver and exerts tumour-suppressive effects through the regulation of the TME.

The aberrant activation of Notch results in the initiation and progression of tumours. In colorectal cancer (CRC), Notch signalling controls the immune contexture and tumour cell morphology, promoting immune evasion and cancer progression (58). In breast cancer cells, Notch binds to JAG1 to secrete macrophage-polarizing cytokines like IL- β or CCL2, resulting in M2-type macrophage recruitment, producing TGF- β cues to tumour cells. Crosstalks such as these may be responsible for drug resistance, which needs further validation (59).

Involvement of Notch signalling in CSC T-cell interaction

In addition to driving the growth of tumours through mesenchymal transition, cancer stem cells (CSCs) play a role in fostering immune evasion within the TME, allowing them to effectively evade the anti-tumour immune responses. Additionally, CSCs with the capacity to generate immunosuppressive molecules, like TGF- β , IL-10, and IL-13, contribute to immune dysfunctions (60). Notch1 plays an important role in governing luminal estrogen receptor (ER⁺) breast cancer and breast cancer stem cells (BCSCs). Interestingly, the levels showed an opposite relationship with the more aggressive triple-negative or basal-like BCSCs and the presence of infiltrating Foxp3⁺ Tregs. These Tregs produced inhibitory cytokines like IL-4 and IL-10, and they also expressed CTLA-4. Enhanced survival in various types of cancer cases was associated with elevated T-cell counts, particularly in activated

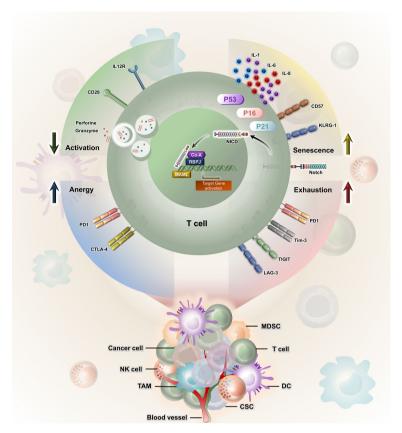


FIGURE 2

Notch signalling has bi-effector functions in T cell-mediated anti-tumour immunity. Notch receptor-ligand interaction controls senescence, exhaustion, anergy, and activation of T cells by regulating the expression of various associated molecules like KLRG1, PD-1, Tim3, LAG-3, CTLA-4, perforin, and granzyme. CSC, cancer stem cell; MDSC, myeloid-derived suppressor cell; DC, dendritic cell; TAM, tumour-associated macrophage; PD-1 1, programmed cell death protein 1; CTLA4, cytotoxic T-cell lymphocyte-associated protein 4; KLRG, killer cell lectin-like receptor subfamily G member: TIGIT. T-cell immunoreceptor with Ig and ITIM domains: TIM3. T-cell immunoglobulin and mucin domain 3.

CD8⁺ CTLs and Th1 cells. Studies have indicated that there is a direct role in regulating gene expression specific to CTLs, including granzyme B, played by both Notch1 and Notch2. Notably, CD8⁺ T cells lacking Notch2, but not those lacking Notch1, demonstrated an inability to proliferate and effectively inhibit tumour growth in mice (61). The Notch signalling-mediated crosstalk between T cells and other intratumoral components is depicted in Figure 3.

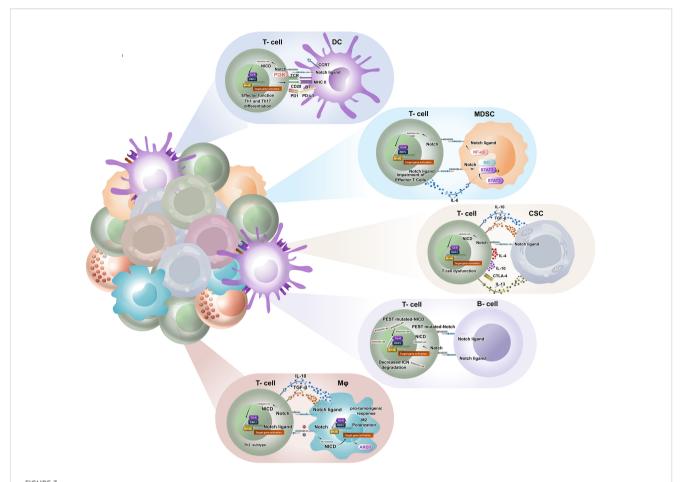
Notch interplay in intratumoral APC-T cell interactions

Naïve T cells receive signals from antigen-presenting cells (APCs) such as DCs, macrophages, and B cells via the release of several cytokines, which have a significant role in both CD4⁺ and CD8⁺ T-cell differentiation.

Notch promotes pro-tumorigenic activity in TAMs

In tumour-resident macrophages, the deletion of Rbpj limited the activity of cytotoxic T cells in B16 melanoma, and Notch activation

improved CD8⁺ T-cell infiltration in Lewis lung carcinoma (LLC) (21, 62). Contrary to the previous observations, the activation of Notch in tumour-associated macrophages (TAMs) elicited protumorigenic response in breast cancer models, including M2 polarization and hindrance of T-cell activity (63). In vivo experiments conducted in a Triple-negative Breast Cancer (TNBC) mouse model reported that intratumoral macrophages, which were educated by tumour-derived Jagged1, inhibited the proliferation and cytotoxicity of T cells via upregulated soluble molecules like CD93 and CD14 (64). A recent study also reported that activated Notch signalling in TAMs led to the upregulated expression of arginase 1, the key driver of immunosuppression, in an engineered mouse model of pancreatic cancer (65). Here, a combinatorial therapy of PD-1 blockade supported by Notch inhibition led to improved intratumoral T-cell infiltration, restriction of tumour size, and tumour cell apoptosis. Contemporary research has also suggested that Notch regulates the switching of macrophage phenotypes within the tumour microenvironment. Notch-deficient macrophages have phenotypes of M2 macrophages with a complex surface marker profile that skews naïve CD4⁺ T cells to produce Th2 subtypes (62). This was further evidenced in osteosarcoma, where the expression of Th2 cytokines like IL-10 and TGF-β was elevated in



Notch signalling mediates bi-directional crosstalk between T cells and other intratumoral components to modulate tumour-immune niche. Notch promotes pro-tumorigenic activity by macrophage switching towards M2 phenotype through regulated secretion of cytokines like IL-10 and TGF-β. Notch signalling is also significant in antigen presentation and intratumoral infiltration of DCs. It directs Th1 and Th17 effector functions against tumour antigen. MDSCs hinder T cell-mediated cytotoxicity through NF-κB or Notch-Stat3-NO cascade. It also promotes intratumoral trafficking of monocytic suppressor cells. Notch upregulation along with TGF-β, IL-10, and IL-13 secretion by CSCs results in T-cell dysfunction. Such CSCs stimulate CTLA-4 expression on effector T cells. There is a huge lacuna in understanding the impact of Notch on B cells in cancer. TGF-β, transforming growth factor beta; DCs, dendritic cells; MDSCs, myeloid-derived suppressor cells; CSCs, cancer stem cells.

Notch1 knockout mice compared to control mice. In contrast, the expression of Th1 cytokines like IL-6, TNF α , and IL-1 β was dampened in Notch1 knockout mice (66). Hence, it is suggested that Notch signalling in macrophages regulates phenotype switching and promotes tumour advancement.

Notch stimulates intratumoral DC-T cell interplay

Notch has been elucidated as a prime signalling pathway in DC–T cell interaction for inducing the effector functions of CD8⁺ T cells. Studies on tumour-infiltrating immune cells in pancreatic and lung cancers have revealed an association between the expression of Notch ligands by tissue-resident DCs and the cytolytic activity of T cells expressing Notch receptors and PD-1. The expression of PD-1 is controlled by several cell-to-cell interactions, including Notch-driven transcriptional control of the PDCD-1 gene. Recent data on murine lung cancer suggest that mice generated with the deletion of

Notch ligand DLL1 in the CD11c lineage promoted tumour growth and increased PD-1 expression on CD8+ T cells, which led to the inhibition of antitumoral T-cell function and long-term T-cell memory responses (52). Intratumoral T-cell infiltration was augmented in mice with the expression of DLL1 on bone marrow and DCs, thereby reducing tumour growth and prolonging mouse survival (53, 60). In mouse bladder cancer, DC subsets with upregulated DLL4 expression during exposure to low doses of antigen have been reported to promote phosphatidylinositol 3-OH kinase (PI3K)-mediated TCR-CD28 interaction, thereby potentiating CD4+ T cells to evoke elevated anti-tumour effect (67). A similar subset of DLL4+ DC population was found in human peripheral blood with inflammatory conditions and was found to aid Th1 and Th17 differentiation (68). Adoptive transfer of Notch-primed DCs restricted the progression of inflammationassociated colorectal carcinogenesis. Notch2 signalling mediated the upregulation of CCR7, promoting DC migration and crosspresentation to effector T cells, which was supported by the fact that RBPJ knockdown in murine DCs showed unrestricted tumour

growth *in vivo* due to poor DC infiltration and subsequent antigen presentation (69). Hence, Notch activation aids DC-mediated T-cell functionality.

Notch in intratumoral MDSC-T cell crosstalk in cancer

Tumours promote the process of immunosuppression and evasion of immunosurveillance by several kinds of suppressor cells, including myeloid-derived suppressor cells (MDSCs), which are represented by a heterogeneous population of immature myeloid cells. Sierra et al. (2015) investigated the importance of Notch1 and Notch2 expression in intratumoral CD8+ T cells of Lewis lung carcinoma. Experimental knockout of Notch1 and Notch2 led to decreased production of IFN-γ and restricted the proliferation and infiltration of activated CD8⁺ T cells (70). MDSCs hindered the expression of Notch1 and Notch2 on T cells via a nitric oxide-dependent cascade. In the same study, it was shown that the transgenic expression of N1ICD in antigen-specific effector T cells had no impact on T-cell activation or proliferation but resulted in increased cytotoxicity and induced a T-cell phenotype with central memory. These effector T cells were resistant to MDSC-mediated tolerogenicity. In a similar collaborative study, they reported that tumour-dwelling MDSCs upregulated the expression of Notch ligand Jagged1 via the NF-KB pathway. In this case, significant prognosis was observed in mouse models of 3LL lung carcinoma and EG-7 lymphoma when treated with anti-Jagged1/2 antibody. The therapeutic response was based on increased CD8 functionality (71). Anti-Jagged therapy in EG-7 tumour intensified the impact of anti-ovalbumin adoptive T-cell therapy (ACT) (72). A study on breast cancer patients suggested that MDSCs augmented tumour progression by hampering the production of effector T-cell cytolytic enzymes like granzyme, perforin, and IFN-γ, as well as by enhancing breast cancer cell stem-like properties. Further research revealed that the mechanism pertained to a crosstalk between STAT3 and Notch pathways. It resulted in the induction of MDSCs to cause IL-6-dependent phosphorylation of STAT3, which led to the activation of Notch via the nitric oxide pathway (73). In a central nervous system cancer model, after oncolytic herpes simplex virus-based therapy, γ-secretase inhibitor (GSI)mediated Notch blockade in myeloid cells reduced CCL2 secretion, thereby preventing the infiltration of immunosuppressive monocytic MDSCs to infect tumour sites and promoting T cellmediated killing (74).

Notch dysregulation in T-cell malignancies

According to prior reports, Notch mutation drives T-cell acute lymphocytic leukaemia (T-ALL). In T-cell malignancies, specifically T-ALL, Notch1 is often mutated, influencing leukemic shift by supporting proliferation and hindering differentiation. Persistent Notch activation results in unregulated proliferation, constrains

differentiation, and improves the survival of leukemic cells through the transcriptional increase of oncogenic targets such as MYC, HES1, and IL7R. Frameshift, nonsense, or alternative splicing mutations, which affect the PEST domain, are sometimes noted in the absence of mutations, disturbing other negative regulatory regions. PEST mutations are mostly observed in chronic lymphocytic leukaemia, mantle cell lymphoma, and peripheral T-cell lymphomas like adult T-cell leukaemia/lymphoma (75).

Notch-mediated oncotherapy involving T cells

Given the importance of Notch in determining immune cell differentiation and consequently moulding the range and variety of immune responses in the TME, several clinicians and researchers have identified Notch as a potential target for therapy. Multiple preclinical studies have unravelled many T-cell therapeutic facets involving the Notch pathway. Some of them are discussed below.

γ -Secretase inhibitors in T cell-mediated oncotherapy

A study conducted by Yu and colleagues (2018) on colorectal cancer indicated that the inhibition of the Notch cascade via N-[N-(3,5-difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester (DAPT), which is a well-known GSI, resulted in improved cytotoxicity of CD8+ T cells infiltrating the TME. DAPT upregulates proinflammatory cytokine production comprising TNF α , IL-1 β , IFN- γ , IL-6, and IL-8 from a tumour-infiltrating effector CD8⁺ T-cell population. DAPT treatment also resulted in the reduced expression of PD-1 exhaustion marker with little impact on cell proliferation. Therefore, Notch blockade improved cytolytic and cytokine-dependent effector functions of T cells in colorectal cancer (22). A recent study assessed the effect of γ secretase inhibitors in T cell-dependent lysis of multiple myeloma (MM) cells. The study disclosed that in ex vivo co-culture setups of MM cells and T cells, GSIs reversed the inhibition of lysis of MM cells induced by soluble B-cell maturation antigen and improved the capacity of BCMAxCD3 bispecific antibodies to lyse MM cells obtained from patients. Furthermore, GSIs also increased T-cell cytotoxicity by increasing the expression of TNFα, CD107a, IL-2, and IFN- γ (76).

Immune checkpoint blockade involving Notch activation

In small cell lung carcinoma (SCLC), the activation of Notch signalling confirmed good clinical prognosis in cohorts with relapsed SCLC receiving immune checkpoint blockade (ICB) therapy (82). In another study on SCLC, the suppression of lysine-specific demethylase 1a (LSD1) led to the activation of Notch signalling. Further investigation suggested that LSD1

inhibition by bomedemstat and consequent Notch activation evoked a better response to anti-PD-1 therapy (77). Contrastingly, in gastric cancer, Notch3 seemed to be inversely correlated with the biomarkers of ICB therapy, including gene expression profiling (GEP), tumour mutational burden (TMB), and innate anti-PD-1 resistance (IPRES) signature, suggesting its importance as a predictive biomarker (78). Combinatorial ICB therapy with GSI restricted tumour progression in triple-negative breast cancer (64). Recent transcriptomic data analysis showed that the increased expression of Notch1/4 enhanced relapse in ERnegative breast cancer. The expression patterns of three intratumoral cells, namely, MDSCs, TAMs, and cancer-associated fibroblasts (CAFs), are known to hinder intratumoral T-cell infiltration, which is essential for the determination of the T-cell exclusion score. In ICB therapy of ER-negative breast cancer, Notch1/4 was synergistically linked with T-cell exclusion score (79). Cytotoxicity-associated molecules like granzyme B, perforin 1, and exhaustion marker PD-1 were upregulated in colorectal carcinoma with Notch mutation (80).

Notch in CAR-iTSCM oncotherapy

A strategy to overcome the scarcity of patient-derived T cells is to generate chimeric antigen receptor (CAR) T cells derived from induced pluripotent stem cells of cancer patients. The production of T cells from induced pluripotent stem cells (iPSCs) is directed by stromal cells that express DLL1 and activate the Notch pathway. Comparable methods have enabled the production of CAR T cells from hematopoietic stem cells; reprogramming factors are added for the induction of pluripotency and have been applied for the proliferation of tumour-specific human T cells. Although this approach facilitates the production of unlimited tumour-specific cytotoxic T lymphocytes, it often results in a low TCR repertoire.

To resolve this, investigators are presently investigating the use of T-stem cell memory (TSCM) cells in the adoptive transfer of T cells,

exploiting their high *in vivo* persistence, self-renewing trait, and multipotency. Adoptive T-cell transfer of CAR T cells produced from TSCM cells initiates more effective anti-tumour responses than those produced from other T-cell subsets (81). Previous experiments conducted by Kondo et al. employed Notch signalling activation to produce induced T memory stem cell (iTSCM) of mouse or human origin. The key traits of TSCM that are also present in iTSCM cells include their ability to show rapid response to antigen re-stimulation and heightened self-renewal efficiency. Moreover, the iTSCM cells also showcase reduced expression of immune checkpoint markers like PD-1 and CTLA-4. The iTSCM cells are produced from intratumoral activated T lymphocytes (81).

Notch inhibition with oncolytic virotherapy

Contemporary research projects the clinical use of oncolytic viruses, specifically herpes simplex virus, as an emerging therapy to kill cancer cells, sparing normal ones. Studies on glioblastoma have revealed that oncolytic herpes simplex virus (oHSV) infection significantly upregulates Notch signalling, stimulating TAMs to produce an increased number of CCL2 and IL-10 and promotes the accumulation of MDSCs in the TME. GSImediated Notch blocking improved CD8+ T cell-oriented antitumour memory response and helped in overcoming the immunosuppressive effect of oHSV therapy. Combinatorial oncolytic virotherapy administered alongside DAPT, GSI, or Notch antibodies showed decreased expression of PD-1 on effector T cells. The treatment of GL261N4-bearing mice receiving combinatorial therapy with depleting antibodies of CD4 and CD8 decreased their survival percentage, emphasizing the significance of tumour-specific T-cell memory response. In glioma mice having decreased Notch expression, cytotoxic T-cell secretions like IFN-γ and granzyme B were compromised (74). The previously described Notch-mediated T-cell oncotherapy is summarized (Table 3).

TABLE 3 Impact of Notch signalling in T cell-mediated oncotherapy.

Type of oncotherapy	Type of malignancy	Notch-mediated effect	Effect of malignancy progression	Reference
γ-Secretase inhibitors (GSIs)	Colorectal cancer	Notch downregulates proinflammatory cytokine production comprising TNF α , IL-1 β , IFN- γ , IL-6, and IL-8 from tumour-infiltrating effector CD8 $^+$ T-cell population.	Notch blockade improved cytolytic and cytokine-dependent effector functions of T cells in colorectal cancer.	(22)
Immune checkpoint blockade (ICB)	SCLC	The inhibition of lysine-specific demethylase 1a (LSD1) led to the activation of Notch signalling. LSD1 suppression by bomedemstat and resulting Notch activation induced better response to anti-PD-1 therapy	Good clinical prognosis in cases of relapsed SCLC receiving ICB therapy.	(77, 82)
Oncolytic herpes simplex virus (oHSV)	Glioblastoma	Upregulation of Notch signalling stimulates TAMs to produce increased number of CCL2 and IL-10.	Decreased Notch expression resulted in compromised cytotoxic T-cell secretions like IFN-γ and granzyme B.	(74)
CAR-iTSCM cell therapy	Multiple myeloma	Notch1 signalling activation is required for increased CAR T-cell proliferation.	Reduced expression of CTLA4 and PD-1.	(81)

SCLC, small cell lung carcinoma; TAMs, tumour-associated macrophages; CAR, chimeric antigen receptor.

Harnessing the versatility of Notch in T cell-based oncotherapy

In order to demonstrate the flexibility of the Notch receptor's activation, here, we discuss synthetic Notch (SynNotch) receptors. SynNotch CAR T cells are an innovative strategy of engineered T cells that integrate SynNotch receptors and CARs on T cells to facilitate target specificity and control proliferation in onco-immunotherapy. SynNotch receptors repurpose the essential proteolytic domain of the innate Notch receptors. In SynNotch receptors, both the extracellular ligand-binding domain and the intracellular transcriptional domain of Notch are substituted with customized modules. Significantly, while the SynNotch system depends on the ligand-mediated proteolytic cleavage mechanism of Notch, it never activates the endogenous Notch signalling pathway. This modular design allows diverse responses upon the recognition of ligand and underscores the enormous potential of Notch-based systems in therapeutic applications (83).

Lim and colleagues (2020) generated the SynNotch system. SynNotch CAR T cells are engineered to recognize multiple tumour antigens (both intracellular and extracellular) and cause explicit tumour cell killing in a heavily heterogeneous cell population of solid tumours (84, 85). In contrast to traditional CAR T cells, which show an immediate response upon identifying the tumour antigen, SynNotch CAR T cells need a two-step activation procedure. The SynNotch receptor is synthesized to recognize an initial priming antigen of the tumour cells, thereby inducing the expression of a chimeric antigen receptor for the recognition of another tumourassociated antigen. This AND-gate logic works like a combinative antigen recognition system, which improves the efficacy of T cellbased oncotherapies. The strategy requires the availability of two separate antigens, one of which is recognized by the SynNotch receptor and the other by the CAR, ensuring that T cells undergo activation only within the context of the tumour microenvironment. This helps in overcoming off-target toxicity (86). These T cells upregulate transcription factors like T-bet, promoting Th1-mediated anti-tumour immunity. Furthermore, they produce T-cell proliferative and polarizing cytokines like IL-2 and IL-12. The SynNotch circuit prompts receptor-specific elimination of tumour cells and stimulates tumour killing via tumour necrosis factor-related apoptosis-inducing ligand (TRAIL). SynNotch diminishes the population of exhausted and anergic T cells and sustains an increased number of T cells in a naïve or memory state (87).

Concluding perspectives

Notch signalling is indispensable for regulating the dynamics of intratumoral cellular interactions. In the tumour microenvironment, Notch acts as a dual-edged sword. Observational and clinical evidence suggests that the tumour microenvironment limits receptor and

ligand-mediated Notch activation, subsequently repressing antitumour T-cell function. Notch convincingly promotes DC-T cell interaction, resulting in improved T-cell priming and subsequent reduction of tumour burden via T-cell cytotoxicity in animal models. The effect of Notch activation on macrophages is context-dependent, with potential pro-tumorigenic outcomes observed in certain cancer models. Many studies have highlighted the importance of Notch signalling in controlling the phenotypic switching of macrophages within tumours, exerting the balance between pro- and anti-tumour immune responses. The activation of Notch, particularly Notch1/2 in CD8⁺ T cells, stimulates a robust anti-tumour response. It has assorted functionalities in the stimulation of effector T cells in an immunosuppressive tumour microenvironment and promotes tumour clearance via cytolytic mediators like granzyme B and IFNy in mouse models. This proposes that targeting the Notch cascade in CD8+ T cells could serve as a promising strategy for boosting antitumour immunity. However, chronic Notch stimulation via APC contributes to PD-1-mediated T-cell exhaustion. Targeted management of the Notch pathway may aid in overcoming MDSCarbitrated immunosuppression and improving T-cell efficacy, signifying possible therapeutic openings in oncotherapy. Although the expression of Notch1 in BCSCs is suspected to be inversely related to T cell-mediated cytotoxicity, the role of Notch in epithelialmesenchymal transition (EMT) via CSC-T cell interaction is substantially dicey and needs further experimental validations for better understanding. Patient data regarding the activity of Notchmediated T-cell response are scarce. Several therapeutic strategies have been developed based on genetic signatures specific to tumour cells in varying kinds of malignancies. There is a sufficient lacuna in the characterization of intratumoral T cells in patients. Even then, an array of Notch-targeted T cell-mediated therapeutic regimes have been experimentally validated as discussed previously. GSIs, CAR T cells, and Notch-associated oncolytic virotherapy have obtained considerable success in preclinical experimental outcomes and initial phases of clinical trials, which abets the first-line use of tumour-specific, economic, and safe Notch therapeutics in patient care. SynNotch CAR T cells are especially fascinating, as their targeted cytotoxic response provides an improved and explicit anti-tumour response. Apart from the assessments of risk and efficiency of Notch therapeutics, it should also be ensured that persistent Notch activation does not result in leukemic mutations of T cells or advanced tumour progression. Notch signalling serves as an oncogenic driver in various malignancies like T-cell acute lymphoblastic leukaemia/lymphomas and peripheral T-cell lymphomas, which complicates its role in cancer biology. While Notch signalling holds a promising therapeutic target for enhancing anti-tumour immunity, its subtle effects on immune cell function and tumour progression should be carefully considered for better clinical translation. In the era of precision medicine, emerging experimental evidence in specific malignancies is aiding substantial progress in the domain of Notch-mediated remedies in T cell-based targeted oncotherapy.

Author contributions

JS: Writing – original draft. PC: Writing – original draft. SRB: Writing – review & editing. MC: Writing – review & editing. AG: Writing – review & editing. PD: Writing – review & editing. JD: Writing – review & editing. GI: Writing – review & editing. AS: Writing – review & editing. SD: Writing – review & editing. NG: Writing – review & editing. RB: Writing – review & editing. AB: Writing – review & editing. SPB: Writing – review & editing.

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References

- 1. Campos-Martín Y, Martínez N, Martínez-López A, Cereceda L, Casado F, Algara P, et al. Clinical and diagnostic relevance of NOTCH2 -and KLF2 -mutations in splenic marginal zone lymphoma. *Haematologica*. (2017) 102:e310–2. doi: 10.3324/haematol.2016.161711
- 2. Koch U, Radtke F. Notch in T-ALL: new players in a complex disease. *Trends Immunol.* (2011) 32:434–42. doi: 10.1016/j.it.2011.06.005
- 3. Willander K, Dutta RK, Ungerbäck J, Gunnarsson R, Juliusson G, Fredrikson M, et al. NOTCH1 mutations influence survival in chronic lymphocytic leukemia patients. *BMC Cancer.* (2013) 13:274. doi: 10.1186/1471-2407-13-274
- 4. Lobry C, Ntziachristos P, Ndiaye-Lobry D, Oh P, Cimmino L, Zhu N, et al. Notch pathway activation targets AML-initiating cell homeostasis and differentiation. *J Exp Med.* (2013) 210:301–19. doi: 10.1084/jem.20121484
- 5. Golfman LS, Kannan S, Hall MA, Zweidler-McKay PA. Notch/HES1 implicated as a tumor suppressor mechanism in human AML *in vivo. Blood.* (2012) 120:3524–4. doi: 10.1182/blood.V120.21.3524.3524
- Artavanis-Tsakonas S, Rand MD, Lake RJ. Notch signaling: cell fate control and signal integration in development. Science (80-). (1999) 284:770-6. doi: 10.1126/ science.284.5415.770
- 7. Kovall RA, Gebelein B, Sprinzak D, Kopan R. The canonical notch signaling pathway: structural and biochemical insights into shape, sugar, and force. *Dev Cell.* (2017) 41:228–41. doi: 10.1016/j.devcel.2017.04.001
- 8. Chillakuri CR, Sheppard D, Lea SM, Handford PA. Notch receptor-ligand binding and activation: Insights from molecular studies. *Semin Cell Dev Biol.* (2012) 23:421–8. doi: 10.1016/j.semcdb.2012.01.009
- 9. Kopan R, Ilagan MXG. The canonical notch signaling pathway: unfolding the activation mechanism. *Cell.* (2009) 137:216–33. doi: 10.1016/j.cell.2009.03.045
- $10.\,$ Meng Y, Bo Z, Feng X, Yang X, Handford PA. The notch signaling pathway: mechanistic insights in health and disease. Engineering.~(2024)~34:212-32. doi: 10.1016/ j.eng.2023.11.011

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- 11. Bhanushali AA, Babu S, Thangapandi VR, Pillai R, Chheda P, Das BR. Mutations in the HD and PEST domain of notch-1 receptor in T-cell acute lymphoblastic leukemia: report of novel mutations from Indian population. *Oncol Res Featur Preclin Clin Cancer Ther*. (2010) 19:99–104. doi: 10.3727/096504010X12864748215007
- 12. Zhou B, Lin W, Long Y, Yang Y, Zhang H, Wu K, et al. Notch signaling pathway: architecture, disease, and therapeutics. *Signal Transduct Target Ther*. (2022) 7:95. doi: 10.1038/s41392-022-00934-y
- 13. Bray SJ. Notch signalling: a simple pathway becomes complex. *Nat Rev Mol Cell Biol.* (2006) 7:678–89. doi: 10.1038/nrm2009
- 14. Friedrich T, Ferrante F, Pioger L, Nist A, Stiewe T, Andrau JC, et al. Notch-dependent and -independent functions of transcription factor RBPJ. *Nucleic Acids Res.* (2022) 50:7925–37. doi: 10.1093/nar/gkac601
- 15. DeHart SL, Heikens MJ, Tsai S. Jagged2 promotes the development of natural killer cells and the establishment of functional natural killer cell lines. *Blood.* (2005) 105:3521–7. doi: 10.1182/blood-2004-11-4237
- 16. Neves H, Weerkamp F, Gomes AC, Naber BAE, Gameiro P, Becker JD, et al. Effects of delta1 and jagged1 on early human hematopoiesis: correlation with expression of notch signaling-related genes in CD34 + Cells. *Stem Cells.* (2006) 24:1328–37. doi: 10.1634/stemcells.2005-0207
- 17. Beck RC, Padival M, Yeh D, Ralston J, Cooke KR, Lowe JB. The notch ligands jagged2, delta1, and delta4 induce differentiation and expansion of functional human NK cells from CD34+ Cord blood hematopoietic progenitor cells. *Biol Blood Marrow Transpl.* (2009) 15:1026–37. doi: 10.1016/j.bbmt.2009.06.002
- 18. Manaster I, Gazit R, Goldman-Wohl D, Stern-Ginossar N, Mizrahi S, Yagel S, et al. Notch activation enhances IFN γ secretion by human peripheral blood and decidual NK cells. *J Reprod Immunol.* (2010) 84:1–7. doi: 10.1016/j.jri.2009.10.009
- 19. Kijima M, Yamaguchi T, Ishifune C, Maekawa Y, Koyanagi A, Yagita H, et al. Dendritic cell-mediated NK cell activation is controlled by Jagged2–Notch interaction. *Proc Natl Acad Sci.* (2008) 105:7010–5. doi: 10.1073/pnas.0709919105

- 20. Benne C, Lelievre JD, Balbo M, Henry A, Sakano S, Levy Y. Notch increases T/NK potential of human hematopoietic progenitors and inhibits B cell differentiation at a pro-B stage. *Stem Cells.* (2009) 27:1676–85. doi: 10.1002/stem.94
- Zhao JL, Huang F, He F, Gao CC, Liang SQ, Ma PF, et al. Forced activation of notch in macrophages represses tumor growth by upregulating miR-125a and disabling tumor-associated macrophages. *Cancer Res.* (2016) 76:1403–15. doi: 10.1158/0008-5472.CAN-15-2019
- 22. Guo S, Liu J, Zhang Y, Liang S, Jiang X, Lin Y. Notch signaling regulates M1-type polarization in macrophages by inhibiting signal regulatory protein α (SIRP α). Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi. (2021) 37:673–8.
- 23. Monsalve E, Pérez MA, Rubio A, Ruiz-Hidalgo MJ, Baladrón V, García-Ramírez JJ, et al. Notch-1 up-regulation and signaling following macrophage activation modulates gene expression patterns known to affect antigen-presenting capacity and cytotoxic activity. *J Immunol.* (2006) 176:5362–73. doi: 10.4049/jimmunol.176.9.5362
- 24. López-López S, Monsalve EM, Romero de Ávila MJ, González-Gómez J, Hernández de León N, Ruiz-Marcos F, et al. NOTCH3 signaling is essential for NF-κB activation in TLR-activated macrophages. *Sci Rep.* (2020) 10(1):14839. doi: 10.1038/s41598-020-71810-4
- 25. Kang J, Kim W, Park S. Notch1 is an important mediator for enhancing of B-cell activation and antibody secretion by N otch ligand. Immunology. (2014) 143:550–9. doi: 10.1111/imm.12333
- 26. Zhu G, Wang X, Xiao H, Liu X, Fang Y, Zhai B, et al. Both Notch1 and its ligands in B cells promote antibody production. *Mol Immunol.* (2017) 91:17–23. doi: 10.1016/j.molimm.2017.08.021
- 27. Hozumi K, Negishi N, Suzuki D, Abe N, Sotomaru Y, Tamaoki N, et al. Deltalike 1 is necessary for the generation of marginal zone B cells but not T cells *in vivo. Nat Immunol.* (2004) 5:638–44. doi: 10.1038/ni1075
- 28. Hammad H, Vanderkerken M, Pouliot P, Deswarte K, Toussaint W, Vergote K, et al. Transitional B cells commit to marginal zone B cell fate by Taok3-mediated surface expression of ADAM10. *Nat Immunol.* (2017) 18:313–20. doi: 10.1038/ni.3657
- 29. Maekawa Y, Tsukumo Si, Chiba S, Hirai H, Hayashi Y, Okada H, et al. Delta1-notch3 interactions bias the functional differentiation of activated CD4+ T cells. *Immunity.* (2003) 19:549–59. doi: 10.1016/S1074-7613(03)00270-X
- 30. Sun J, Krawczyk CJ, Pearce EJ. Suppression of th2 cell development by notch ligands delta1 and delta4. J Immunol. (2008) 180:1655–61. doi: 10.4049/j jimmunol.180.3.1655
- 31. Samon JB, Champhekar A, Minter LM, Telfer JC, Miele L, Fauq A, et al. Notch1 and TGFbeta1 cooperatively regulate Foxp3 expression and the maintenance of peripheral regulatory T cells. *Blood.* (2008) 112:1813–21. doi: 10.1182/blood-2008-03-144980
- 32. Ting HA, de Almeida Nagata D, Rasky AJ, Malinczak CA, Maillard IP, Schaller MA, et al. Notch ligand Delta-like 4 induces epigenetic regulation of Treg cell differentiation and function in viral infection. *Mucosal Immunol.* (2018) 11:1524–36. doi: 10.1038/s41385-018-0052-1
- 33. Srivastava M, Murriel C, Yun R, Mayes E, Jie HB, Axelrod F, et al. Co-targeting of delta-like ligand 4 (DLL4) and vascular endothelial growth factor a (VEGF) with programmed death 1 (PD1) blockade inhibits tumor growth and facilitates anti-tumor immune responses. *J Immunother Cancer*. (2015) 3:P373. doi: 10.1186/2051-1426-3-22.233
- 34. Pitulescu ME, Schmidt I, Giaimo BD, Antoine T, Berkenfeld F, Ferrante F, et al. Dll4 and Notch signalling couples sprouting angiogenesis and artery formation. *Nat Cell Biol.* (2017) 19:915–27. doi: 10.1038/ncb3555
- 35. Yustein JT, Liu YC, Gao P, Jie C, Le A, Vuica-Ross M, et al. Induction of ectopic Myc target gene JAG2 augments hypoxic growth and tumorigenesis in a human B-cell model. *Proc Natl Acad Sci.* (2010) 107:3534–9. doi: 10.1073/pnas.0901230107
- 36. Amsen D, Helbig C, Backer RA. Notch in T cell differentiation: all things considered. *Trends Immunol.* (2015) 36:802–14. doi: 10.1016/j.it.2015.10.007
- 37. Wolfer A, Wilson A, Nemir M, MacDonald HR, Radtke F. Inactivation of notch1 impairs VDJ β Rearrangement and allows pre-TCR-independent survival of early $\alpha\beta$ Lineage thymocytes. *Immunity*. (2002) 16:869–79. doi: 10.1016/S1074-7613 (02)00330-8
- 38. Amsen D, Blander JM, Lee GR, Tanigaki K, Honjo T, Flavell RA. Instruction of distinct CD4 T helper cell fates by different notch ligands on antigen-presenting cells. *Cell.* (2004) 117:515–26. doi: 10.1016/S0092-8674(04)00451-9
- 39. Mathieu M, Duval F, Daudelin JF, Labrecque N. The notch signaling pathway controls short-lived effector CD8+ T cell differentiation but is dispensable for memory generation. *J Immunol.* (2015) 194:5654–62. doi: 10.4049/jimmunol.1402837
- 40. Guha I, Bhuniya A, Shukla D, Patidar A, Nandi P, Saha A, et al. Tumor arrests DN2 to DN3 pro T cell transition and promotes its conversion to thymic dendritic cells by reciprocally regulating notch1 and ikaros signaling. *Front Immunol.* (2020) 11. doi: 10.3389/fimmu.2020.00898
- 41. Tsukumo Si, Yasutomo K. Regulation of CD8+ T cells and antitumor immunity by notch signaling. Front Immunol. (2018) 9. doi: 10.3389/fimmu.2018.00101
- 42. Tchekneva EE, Goruganthu MUL, Uzhachenko RV, Thomas PL, Antonucci A, Chekneva I, et al. Determinant roles of dendritic cell-expressed Notch Delta-like and Jagged ligands on anti-tumor T cell immunity. *J Immunother Cancer*. (2019) 7:95. doi: 10.1186/s40425-019-0566-4

- 43. Huang Y, Lin L, Shanker A, Malhotra A, Yang L, Dikov MM, et al. Resuscitating cancer immunosurveillance: selective stimulation of DLL1-notch signaling in T cells rescues T-cell function and inhibits tumor growth. *Cancer Res.* (2011) 71:6122–31. doi: 10.1158/0008-5472.CAN-10-4366
- 44. Maekawa Y, Minato Y, Ishifune C, Kurihara T, Kitamura A, Kojima H, et al. Notch2 integrates signaling by the transcription factors RBP-J and CREB1 to promote T cell cytotoxicity. *Nat Immunol.* (2008) 9:1140–7. doi: 10.1038/ni.1649
- 45. Yu W, Wang Y, Guo P. Notch signaling pathway dampens tumor-infiltrating CD8+ T cells activity in patients with colorectal carcinoma. *BioMed Pharmacother*. (2018) 97:535–42. doi: 10.1016/j.biopha.2017.10.143
- 46. Xie M, Fu X, Jiang K. Notch1/TAZ axis promotes aerobic glycolysis and immune escape in lung cancer. *Cell Death Dis.* (2021) 12:832. doi: 10.1038/s41419-021-04124-6
- 47. Biktasova AK, Dudimah DF, Uzhachenko RV, Park K, Akhter A, Arasada RR, et al. Multivalent forms of the notch ligand DLL-1 enhance antitumor T-cell immunity in lung cancer and improve efficacy of EGFR-targeted therapy. *Cancer Res.* (2015) 75:4728–41. doi: 10.1158/0008-5472.CAN-14-1154
- 48. Zhao E, Maj T, Kryczek I, Li W, Wu K, Zhao L, et al. Cancer mediates effector T cell dysfunction by targeting microRNAs and EZH2 via glycolysis restriction. *Nat Immunol.* (2016) 17:95–103. doi: 10.1038/ni.3313
- 49. Sugimoto K, Maekawa Y, Kitamura A, Nishida J, Koyanagi A, Yagita H, et al. Notch2 signaling is required for potent antitumor immunity *in vivo. J Immunol.* (2010) 184:4673–8. doi: 10.4049/jimmunol.0903661
- 50. Meng L, Hu S, Wang J, He S, Zhang Y. DLL4+ dendritic cells: Key regulators of Notch Signaling in effector T cell responses. *Pharmacol Res.* (2016) 113:449–57. doi: 10.1016/j.phrs.2016.09.001
- 51. Dai K, Huang L. Huang Y bing, Chen Z bing, Yang L hua, Jiang Y an. 1810011010 Rik Inhibits the Antitumor Effect of Intratumoral CD8+ T Cells through Suppression of Notch2 Pathway in a Murine Hepatocellular Carcinoma Model. Front Immunol. (2017) 8. doi: 10.3389/fimmu.2017.00320
- 52. Wang RX, Li YF, Hao CH, Fang W. Increased Notch receptors induces CD8(+) T cell exhaustion in patients with prostate cancer. *Zhonghua Yi Xue Za Zhi.* (2020) 100 (34):2669–74. doi: 10.3760/cma.j.cn112137-20200509-01468
- 53. Hoare M, Ito Y, Kang TW, Weekes MP, Matheson NJ, Patten DA, et al. NOTCH1 mediates a switch between two distinct secretomes during senescence. *Nat Cell Biol.* (2016) 18:979–92. doi: 10.1038/ncb3397
- 54. Demehri S, Turkoz A, Manivasagam S, Yockey LJ, Turkoz M, Kopan R. Elevated epidermal thymic stromal lymphopoietin levels establish an antitumor environment in the skin. *Cancer Cell.* (2012) 22:494–505. doi: 10.1016/j.ccr.2012.08.017
- 55. Demehri S, Turkoz A, Kopan R. Epidermal notch1 loss promotes skin tumorigenesis by impacting the stromal microenvironment. *Cancer Cell.* (2009) 16:55–66. doi: 10.1016/j.ccr.2009.05.016
- 56. Demehri S, Liu Z, Lee J, Lin MH, Crosby SD, Roberts CJ, et al. Notch-deficient skin induces a lethal systemic B-lymphoproliferative disorder by secreting TSLP, a sentinel for epidermal integrity. *PloS Biol.* (2008) 6(1):e123. doi: 10.1016/j.ccr.2009.05.016
- 57. Parmigiani E, Ivanek R, Rolando C, Hafen K, Turchinovich G, Lehmann FM, et al. Interferon- γ resistance and immune evasion in glioma develop via Notch-regulated co-evolution of Malignant and immune cells. *Dev Cell.* (2022) 57:1847–1865.e9. doi: 10.1016/j.devcel.2022.06.006
- 58. Sun J, Chen Y, Xu Z, Wang W, Li P. Notch signaling in the tumor immune microenvironment of colorectal cancer: mechanisms and therapeutic opportunities. *J Transl Med.* (2025) 23:315. doi: 10.1186/s12967-025-06282-z
- 59. Shen Q, Cohen B, Zheng W, Rahbar R, Martin B, Murakami K, et al. Notch shapes the innate immunophenotype in breast cancer. *Cancer Discov.* (2017) 7:1320–35. doi: 10.1158/2159-8290.CD-17-0037
- 60. Yu G, Gong J. Targeting CSC-immune cell crosstalk to overcome chemoresistance and enhance immunotherapy efficacy. *Front Immunol.* (2025) 16:1620807. doi: 10.3389/fimmu.2025.1620807
- 61. Janghorban M, Xin L, Rosen JM, Zhang XHF. Notch signaling as a regulator of the tumor immune response: to target or not to target? *Front Immunol.* (2018) 9. doi: 10.3389/fimmu.2018.01649
- 62. Wang YC, He F, Feng F, Liu XW, Dong GY, Qin HY, et al. Notch signaling determines the M1 versus M2 polarization of macrophages in antitumor immune responses. *Cancer Res.* (2010) 70:4840–9. doi: 10.1158/0008-5472.CAN-10-0269
- 63. Liu H, Wang J, Zhang M, Xuan Q, Wang Z, Lian X, et al. Jagged1 promotes aromatase inhibitor resistance by modulating tumor-associated macrophage differentiation in breast cancer patients. *Breast Cancer Res Treat*. (2017) 166:95–107. doi: 10.1007/s10549-017-4394-2
- $64.\,$ Meng J, Jiang Yz, Zhao S, Tao Y, Zhang T, Wang X, et al. Tumor-derived Jagged1 promotes cancer progression through immune evasion. Cell Rep. (2022) 38:110492. doi: 10.1016/j.celrep.2022.110492
- 65. Yan W, Menjivar RE, Bonilla ME, Steele NG, Kemp SB, Du W, et al. Notch signaling regulates immunosuppressive tumor-associated macrophage function in pancreatic cancer. *Cancer Immunol Res.* (2024) 12:91–106. doi: 10.1158/2326-6066.CIR-23-0037
- 66. Ren S, Zhang X, Hu Y, Wu J, Ju Y, Sun X, et al. Blocking the Notch signal transduction pathway promotes tumor growth in osteosarcoma by affecting

polarization of TAM to M2 phenotype. Ann Transl Med. (2020) 8:1057-7. doi: 10.21037/atm-20-3881

- 67. Laky K, Evans S, Perez-Diez A, Fowlkes BJ. Notch signaling regulates antigen sensitivity of naive CD4+ T cells by tuning co-stimulation. *Immunity*. (2015) 42:80–94. doi: 10.1016/j.immuni.2014.12.027
- 68. Meng L, Bai Z, He S, Mochizuki K, Liu Y, Purushe J, et al. The notch ligand DLL4 defines a capability of human dendritic cells in regulating th1 and Th17 differentiation. *J Immunol.* (2016) 196:1070–80. doi: 10.4049/jimmunol.1501310
- 69. Wang L, Yu S, Chan ER, Chen KY, Liu C, Che D, et al. Notch-regulated dendritic cells restrain inflammation-associated colorectal carcinogenesis. *Cancer Immunol Res.* (2021) 9:348–61. doi: 10.1158/2326-6066.CIR-20-0428
- 70. Sierra RA, Thevenot P, Raber PL, Cui Y, Parsons C, Ochoa AC, et al. Rescue of notch-1 signaling in antigen-specific CD8+ T cells overcomes tumor-induced T-cell suppression and enhances immunotherapy in cancer. *Cancer Immunol Res.* (2014) 2:800–11. doi: 10.1158/2326-6066.CIR-14-0021
- 71. Hossain F, Majumder S, Ucar DA, Rodriguez PC, Golde TE, Minter LM, et al. Notch signaling in myeloid cells as a regulator of tumor immune responses. *Front Immunol.* (2018) 9. doi: 10.3389/fimmu.2018.01288
- 72. Sierra RA, Trillo-Tinoco J, Mohamed E, Yu L, Achyut BR, Arbab A, et al. Antijagged immunotherapy inhibits MDSCs and overcomes tumor-induced tolerance. *Cancer Res.* (2017) 77:5628–38. doi: 10.1158/0008-5472.CAN-17-0357
- 73. Peng D, Tanikawa T, Li W, Zhao L, Vatan L, Szeliga W, et al. Myeloid-derived suppressor cells endow stem-like qualities to breast cancer cells through IL6/STAT3 and NO/NOTCH cross-talk signaling. *Cancer Res.* (2016) 76:3156–65. doi: 10.1158/0008-5472.CAN-15-2528
- 74. Otani Y, Yoo JY, Lewis CT, Chao S, Swanner J, Shimizu T, et al. NOTCH-Induced MDSC Recruitment after oHSV Virotherapy in CNS Cancer Models Modulates Antitumor Immunotherapy. *Clin Cancer Res.* (2022) 28:1460–73. doi: 10.1158/1078-0432.CCR-21-2347
- 75. Di Ianni M, Baldoni S, Rosati E, Ciurnelli R, Cavalli L, Martelli MF, et al. A new genetic lesion in B-CLL: a NOTCH1 PEST domain mutation. *Br J Haematol.* (2009) 146:689–91. doi: 10.1111/j.1365-2141.2009.07816.x
- 76. Chen H, Yu T, Lin L, Xing L, Cho SF, Wen K, et al. γ-secretase inhibitors augment efficacy of BCMA-targeting bispecific antibodies against multiple myeloma

- cells without impairing T-cell activation and differentiation. *Blood Cancer J.* (2022) 12:118. doi: 10.1038/s41408-022-00716-3
- 77. Hiatt JB, Sandborg H, Garrison SM, Arnold HU, Liao SY, Norton JP, et al. Inhibition of LSD1 with bomedemstat sensitizes small cell lung cancer to immune checkpoint blockade and T-cell killing. *Clin Cancer Res.* (2022) 28:4551–64. doi: 10.1158/1078-0432.CCR-22-1128
- 78. Cui Y, Li Q, Li W, Wang Y, Lv F, Shi X, et al. NOTCH3 is a prognostic factor and is correlated with immune tolerance in gastric cancer. *Front Oncol.* (2021) 10. doi: 10.3389/fonc.2020.574937
- 79. Liu D, Hofman P. Expression of NOTCH1, NOTCH4, HLA-DMA and HLA-DRA is synergistically associated with T cell exclusion, immune checkpoint blockade efficacy and recurrence risk in ER-negative breast cancer. *Cell Oncol.* (2022) 45:463–77. doi: 10.1007/s13402-022-00677-6
- 80. Wang F, Long J, Li L, Zhao Z, Wei F, Yao Y, et al. Mutations in the notch signalling pathway are associated with enhanced anti-tumour immunity in colorectal cancer. *J Cell Mol Med.* (2020) 24:12176–87. doi: 10.1111/jcmm.15867
- 81. Kondo T, Ando M, Nagai N, Tomisato W, Srirat T, Liu B, et al. The NOTCH-FOXM1 axis plays a key role in mitochondrial biogenesis in the induction of human stem cell memory–like CAR-T cells. *Cancer Res.* (2020) 80:471–83. doi: 10.1158/0008-5472.CAN-19-1196
- 82. Roper N, Velez MJ, Chiappori A, Kim YS, Wei JS, Sindiri S, et al. Notch signaling and efficacy of PD-1/PD-L1 blockade in relapsed small cell lung cancer. *Nat Commun.* (2021) 12:3880. doi: 10.1038/s41467-021-24164-y
- 83. Roybal KT, Williams JZ, Morsut L, Rupp LJ, Kolinko I, Choe JH, et al. Engineering T cells with customized therapeutic response programs using synthetic notch receptors. *Cell.* (2016) 167:419–432.e16. doi: 10.1016/j.cell.2016.09.011
- 84. Williams JZ, Allen GM, Shah D, Sterin IS, Kim KH, Garcia VP, et al. Precise T cell recognition programs designed by transcriptionally linking multiple receptors. *Science* (80-). (2020) 370:1099–104. doi: 10.1126/science.abc6270
- 85. Saffern M, Samstein R. Taking CAR T cells up a synthetic Notch. *Nat Rev Immunol.* (2021) 21:135–5. doi: 10.1038/s41577-021-00514-1
- 86. Hyrenius-Wittsten A, Su Y, Park M, Garcia JM, Alavi J, Perry N, et al. SynNotch CAR circuits enhance solid tumor recognition and promote persistent antitumor activity in mouse models. *Sci Transl Med.* (2021) 13(591):eabd8836. doi: 10.1126/scitranslmed.abd8836

Glossary

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ACT	adoptive T-cell therapy	MDSCs	myeloid-derived suppressor cells
ADAM	protein family ("A disintegrin and metalloprotease")	MET	mesenchymal-epithelial transition
ANK	ankyrin repeat	MHC-II	major histocompatibility complex II
APC	antigen-presenting cells	MMP9	matrix metalloproteinase 9
BCR	B cell receptor	MPEC	Memory Precursor and Short-Lived Effector cells
CAR	chimeric antigen receptor	MRK 560	γ -secretase inhibitor
CCR7	C-C chemokine receptor type 7	MYC	myelocytomatosis oncogene
CCL2	C-C chemokine ligand 2	NED	Notch extracellular domain
CD4+	cluster of differentiation 4+	NF-I	neurofibromatosis type 1
CD8+	cluster of differentiation 8+	NF-κB	nuclear factor kappa B
cMYC	cellular myelocytomatosis oncogene	NICD	Notch intracellular cytoplasmic domain
CSC	cancer stem cell	NK	natural killer
CTL	cytotoxic T lymphocytes	NLS	nuclear localization signal
CTLA4	cytotoxic T-cell lymphocyte-associated protein 4	NPC	neural precursor cells
DAPT	N-[N-(3,5-difluorophenacetyl)-L-alanyl]-S-phenylglycine t-	NRR	negative regulatory region
DD14	butyl ester	NSCLC	non-small cell lung cancer
DDIS	DNA Damage-Induced Senescence	oHSV	oncolytic herpes simplex virus
DLL	Delta-like ligand	OXPhos	oxidative phosphorylation
EGF	epidermal growth factor	PD-L1	programmed death ligand 1
EGFR	epidermal growth factor receptor	PD-1	programmed cell death protein 1
EMT	epithelial-mesenchymal transition	PECAM	platelet endothelial cell adhesion molecule
EOMES EOC	eomesodermin epithelial ovarian cancer	PEST	a polypeptide sequence rich in proline (P), glutamic acid (E), serine (S), and threonine (T)
ER	estrogen receptor	PI3K/AKT/mTORC1	phosphoinositide-3-kinase/protein kinase B/mammalian
ERK	extracellular signal-regulated kinase	cascade	target of rapamycin complex 1
EZH2	enhancer of zeste homolog 2	PTCL	peripheral T-cell lymphoma
FOXA2	forkhead box protein A2	RBPJ	recombination signal binding protein for immunoglobulin
GATA3	GATA binding protein 3	DIC	kappa J region
$GAT\alpha$	GGA and Tom1 domain	RIS	Replicative-Induced Senescence
GEP	gene expression profiling	RTK SASP	receptor tyrosine kinase
GSI	gamma secretase inhibitors	SLEC	senescence-associated secretory phenotype short-lived effector cells
hes	Hairy enhancer of split	STAT3	signal transducer and activator of transcription 3
hey	Hairy related	T ALL	T-cell acute lymphoblastic leukaemia/lymphomas
HES4	Hes family BHLH transcription factor 4	TAM	tumour-associated macrophage
HNSCC	head and neck squamous cell carcinoma	TBX2	T-box transcription factor 2
ICB	immune checkpoint blockade	TCIM	traditional complementary and integrative medicine
ICI	immune checkpoint inhibitor	TCR	T-cell receptor
IFN-γ	interferon gamma	TNBC	Triple-negative breast cancer
IL-6/8	interleukin-6/8	TGF-β	transforming growth factor beta
iPSCs	induced pluripotent stem cells	TIGIT	T-cell immunoreceptor with Ig and ITIM domains
iTSCM	induced T memory stem cells	TIL	tumour-infiltrating lymphocytes
JAG	Jagged1	TIM3	T-cell immunoglobulin and mucin domain 3
KLK6	Kallikrein-related peptidase 6	ТМВ	tumour mutational burden
KLRG	Killer cell lectin-like receptor subfamily G member 1	TME	tumour microenvironment
LLC	Lewis lung carcinoma	TNFα	tumour necrosis factor alpha
LSD1	demethylase 1a	TRAIL	tumour necrosis factor-related apoptosis-inducing ligand
MAML	Mastermind-like	VEGF	vascular endothelial growth factor
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